

#### UNIVERSIDAD DE LA FRONTERA

### Facultad de Ingeniería, Ciencias y Administración

Programa de Doctorado en Ciencias de Recursos Naturales

Effect of Al and Cd on oxidative stress and production of secondary metabolites of phenolic nature of highbush blueberry (*Vaccinium corymbosum* L.) cultivated *in vitro*.

**Doctoral Thesis** 

**In Partial Fulfillment** 

Of the Requirements for the Degree

**Doctor of Sciences in Natural Resources** 

by

Karen de los Ángeles Manquián Cerda

**TEMUCO-CHILE** 

2014

# "Effect of Al and Cd on oxidative stress and production of secondary metabolites of phenolic nature of highbush blueberry (*Vaccinium corymbosum* L.) cultivated *in vitro*"

Tesis presentada como parte de los requisitos para optar al grado de

Doctor en Ciencias de Recursos Naturales, aprobada por el

### COMITÉ DE TESIS

Director Programa de Postgrado en	Dirección de Postgrado
Ciencias de Recursos Naturales	Universidad de La Frontera
Prof. tutor, Dr. Mauricio Escudey C.	Co-tutor Dr. Gustavo Zúñiga
Dra. Miren Alberdi.	Dr. Alejandro Urzúa
Du Andrés Ovins	
Dr. Andrés Quiroz	

Esta Tesis esta dedicada a Dios... Gracias por tu fidelidad.

#### Agradecimientos / Acknowledgments

Quisiera agradecer en primer lugar a Nicolás por acompañarme en estos años y darme su apoyo incondicional durante toda esta etapa. También a mis amigos, en especial a Luis y Simonet quienes me han ayudado y acompañado, y obviamente a mi familia quienes hacen que los domingos no sean fomingos.

Agradezco a mi tutor, el Dr. Mauricio Escudey por permitirme desarrollar esta tesis en su laboratorio y por contribuir de manera significativa en mi formación científica y profesional mediante su preocupación y confiar en mi trabajo.

Agradezco a mi co-tutor, el Dr. Gustavo Zúñiga por guiarme y apoyar el dasarrollo de esta tesis, además de estar siempre presente cuando necesitaba de su ayuda, y de manera especial al laboratorio de fisiología vegetal por hacerme sentir parte del grupo.

Agradezco a la comisión evaluadora y de manera especial al Dr. Alejandro Urzúa por su tiempo.

Al Programa de Doctorado en Ciencias de Recursos Naturales, el cual me entregó las herramientas necesarias para mi formación científica, brindándome una mirada multidiciplinaria de la ciencia.

Quisiera agradecer de manera especial a la Dra. María de la Luz Mora, por creer en mí, haberme apoyado en mi estadía en el doctorado y brindarme siempre un consejo cuando lo necesite.

A la Universidad de la Frontera por el apoyo y financiamiento entregado, ya que sin su ayuda este trabajo no habría sido posible de realizar.

# **INDEX**

FIGURE INDEX	7
TABLES INDEX	11
RESUMEN	13
ABSTRACT	16
ABBREVIATION	19
OUTLINE DE LA TESIS	20
GENERAL INTRODUCTION	22
HYPOTHESIS	30
GENERAL OBJETIVES	30
SPECIFIC OBJETIVES	30
Chapter 1.	
The Responses of Plants to Trace Elements. A Review Focusing on	Aluminium and
Cadmium	31
Chapter 2.	
Effect of Aluminum on Antioxidant Activity and Phenolic Compour	nds Content in in
vitro Cultured Blueherries	75

# Chapter 3.

Effect of the Availability of Cd <sup>2+</sup> on the non-enzymatic Antioxidant Response and the		
nenolic Compounds Profile of Blueberry (Vaccinium Corymbosum L.) Plantl	lets	
ıltivated in vitro	.95	
napter 4.		
xidative Stress and Production of Compounds of Phenolic Origin in Blueber	rry	
antlets Cultivated in vitro under a Combined Cadmium and Aluminum Treatme	nt	
	125	
SCUSIÓN GENERAL1	l56	
ENERAL CONCLUSIONS1	l <b>6</b> 7	
TITURAS PROVECCIONES	170	

# FIGURE INDEX

Chapter 1. The Responses of Plants to Trace Elements. A Review Focusing on
Aluminium and Cadmium.
Fig.1. Different ways for high metal concentrations to protect plants against biotic
(Taken from Poschenrieder et al., 2006)
Fig. 2. Contribution of apoplast and symplast to Cd uptake by roots of maize (Zm)
and high- (Nc-H) and low- (Nc-L) Cd accumulating ecotypes of Noccaea caerulescens
(Taken from Redjala <i>et al.</i> , 2009)42
Fig. 3. Lateral transport of minerals (micronutrients) and water in roots (Taken
White, 2001)43
Chapter 2. Effect of Aluminum on Antioxidant Activity and Phenolic Compounds
Content in in vitro Cultured Blueberries.
Fig. 1. Effect of the Al concentration and exposure time on the content of MDA in
plantlets of <i>V. corymbosum</i> L. Each value is a mean of three samples ± 1 s.d83
Fig. 2. Antioxidant capacity of extracts of plantlets of V. corymbosum L. treated with
Al. DPPH assay (A) and FRAP assay (B). Each value is a mean of three samples ± 1
s.d
Fig. 3. Variation in the content of total phenolic compounds (TPC) in plantlets of $V_{ij}$
corymbosum L. treated with Al. Each value is a mean of three samples ± 1 s.d
Identification of phenolic compound by HPLC-DAD85

Fig. 4. HPLC profiles of phenolic compound in blueberries cultivated in vitro at 314
nm (A) and 254 nm (B). Blue line: control; Red line: 200μM Al <sup>3+</sup> . In both cases
evaluation was done after 14 days of treatment86
Fig. 5. Variation in the content of chlorogenic acids (up) and ellagic acid (bottom), in
function of exposure time and the concentration of Al in plantlets of V. corymbosum
L. Each value is a mean of three samples ± 1 s.d
Chapter 3. Effect of the Availability of Cd2+ on the non-enzymatic Antioxidant
Response and the Phenolic Compounds Profile of Blueberry (Vaccinium Corymbosum
L.) Plantlets Cultivated in vitro.
Fig. 1.Effect of Cd concentration on oxidative stress in blueberries measured as
MDA. Data are mean $\pm$ sd; n = 6. Different letters (lower case) indicate statistical
differences among means from treatments and time after ANOVA and Tukey HSD
post hoc test. The dashed line indicates the level of MDA measured for control when
initiating treatment103
Fig. 2. Effect of Cd concentration on antioxidant activity of ethanolic extracts. (a)
DPPH free radical scavenging (DPPH consumed), (b) FRAP test (ascorbic acid
equivalents). Data are mean $\pm$ sd; n = 6. Different letters (lower case) indicate
statistical differences among means from treatments and time after ANOVA and
Tukey HSD post hoc test. The dashed line indicates the level of % DPPH and FRAP
value measured for control when initiating treatment105
Fig. 3. Effect of Cd concentration on TPC variation in Vaccinium corymbosum L.
Values are means $\pm$ sd; $n = 6$ . Different letters (lower case) indicate statistical
differences among means from treatments and time after ANOVA and Tukey HSD

post hoc test. The dashed line indicates the level of TPC measured for control when
initiating treatment107
Fig. 4. Relationship between the lipid peroxidation and FRAP (a), MDA and TPC (b)
and FRAP and TPC (c). Pearson's coefficients are indicated112
Fig. 5. Representation of derivative of the antioxidant response ( $\Delta TPC$ , $\Delta FRAP$ ,
$\Delta$ DPPH) and the damage ( $\Delta$ MDA) as a function of Cd concentrations in the growth
medium
Chapter 4. Oxidative stress and production of compounds of phenolic origin in
blueberry plantlets cultivated in vitro under a combined cadmium and aluminum
treatment.
Fig. 1. DPPH scavenging, the result is expressed in % of consumed DPPH. Each
column represent the mean $\pm$ standard deviation, (n= 6; p<0.05). A different lower
case letters indicate statistically significant differences (Tukey, $P < 0.05$ ) between same
day and different treatment. A different upper case letters indicate differences
(Tukey, $P \leq 0.05$ ) between different day and same treatment. The segmented line
shows the % of DPPH consumed for the t=0 days control136
Fig. 2. Ferric reducing/Antioxidant power (FRAP) the results as expressed in ascorbic
acid equivalents (mean $\pm$ standard deviation, n= 6; p<0.05). A different lower case
letters indicate statistically significant differences (Tukey, $P < 0.05$ ) between same day
and different treatment. A different upper case letters indicate differences (Tukey, P ≤
0.05) between different day and same treatment. The segmented line shows the FRAP
value for the t=0 days control

Fig. 3. Effects of the different stress treatments on MDA contents (mean $\pm$ standard
deviation, n=6; p<0.05). A different lower case letter indicate statistically significant
differences (Tukey, p< 0.05) between same day and different treatment. A different
upper case letters indicate differences (Tukey, $P \leq 0.05$ ) between different day and
same treatment. Insert: Normalized data with respect to control
$(MDA_{sample}/MDA_{control})$ . The segmented line shows the MDA level for the t=0 days
control
Fig. 4. Effect of different stress treatments on the total content of phenolic (TPC)
compounds (mean ± standard deviation). A different lower case letter indicates
statistically significant differences (Tukey, $P < 0.05$ ) between same day and different
treatment. A different upper case letters indicate differences (Tukey, $P \leq 0.05$ )
between different day and same treatment. The segmented line shows the TPC for the
t=0 days control140
Fig. 5. Chromatogram of blueberry extracts of 14 days measured at 314 nm, for the
treatment 100Al/100Cd (a) and control (b). The numbers correspond to identified
phenolic compounds (1) chlorogenic acid, (2) Rutin, (3) ellagic acid and (4) Quercetin.
Fig. 6. Variation of (a) chlorogenic and (b) elagic acid content (mean ± standard
deviation) as a function of time and the concentration of the different treatments in
blueberries (the graphs have different scales. A different lower case letter indicates
statistically significant differences (Tukey, $P < 0.05$ ) between same day and different
treatment. A different upper case letter indicates differences (Tukev, $P < 0.05$ )

between different day and same treatment. The segmented line shows the elagic and
chlorogenic acids concentration for the t=0 days control142
Fig. 7. Relations between lipid peroxidation (MDA) and the antioxidant response
measured by FRAP, DPPH, Chlorogenic acid, and ellagic acid addition ratio DPPH
and chlorogenic acid Pearson's coefficients are indicated144

#### TABLES INDEX

Chapter 1. The Responses of Plants to Trace Elements. A Review Focusing or
Aluminium and Cadmium.
Table 1. Essential nutrient elements showing element, symbol and primary forms used
by plants
Table 2. Main sites of production of reactive oxygen species (ROS) in plants and the
respective detoxification mechanisms ('scavengers')48
Chapter 3. Effect of the Availability of Cd2+ on the non-enzymatic Antioxidan
Response and the Phenolic Compounds Profile of Blueberry (Vaccinium Corymbosum
L.) Plantlets Cultivated in vitro.
Table 1. Phenolic compounds in ethanolic extracts for control and Cd exposed
blueberry plantlets109
Table 2. Percent distribution of Cd in the Lloyd-McCown medium culture, for a
concentration of 50 and 100 $\mu M$ of total Cd (Cd_T) added (Cd-anion means Cd
complexed by the corresponding anion, Parker et al., 1995)113

Chapter 4. Oxidative stress and production of compounds of phenolic origin in blueberry plantlets cultivated in vitro under a combined cadmium and aluminum treatment.

Table 1. Percent distribution of Al and Cd in Lloyd-McCown cult	ure medium for the
different conditions studied ("complexed with" means Al and Cd	complexed by the
respective anion, Parker et al., 1995)	134
Table 2. Molar concentration and activity of Al and Cd in Lloy	d-McCown culture
medium for the different treatments applied	135

#### **RESUMEN**

El estudio de los efectos de los metales pesados en plantas ha sido ampliamente descrito en la literatura, en donde se ha investigado su efecto en diferentes parámetros fisiológicos y bioquímicos. Sin embargo existe escasa información sobre la posibilidad de modular el perfil de compuestos fenólicos con respuesta antioxidante u otras propiedades de interés científico, a partir una modificación de la composición de un medio de cultivo o de la solución del suelo. En este contexto elementos presentes de manera natural en el suelo o incorporados por actividades antropogénicas (fertilización, aplicación de enmiendas, entre otras), como son los casos del Al y Cd, surgen como alternativa de estudio y análisis para desarrollar metodologías de manejos de cultivos de interés comercial en diferentes matrices (suelo, solución nutritiva y/o cultivos *in vitro*), potenciando la producción de principios bio-activos con propiedades farmacéutica, cosmetológicas y alimenticias.

La investigación desarrollada en esta tesis se centró en los efectos que puede causar la incorporación de elementos trazas como Al y Cd, en el estrés oxidativo, la capacidad antioxidante y el perfil de compuestos fenólicos, de plántulas de arándano cultivadas *in vitro*. El desarrollo de este estudio significó evaluar el comportamiento del arándano, cuando este es sometido a diferentes concentraciones de Al y Cd en tratamientos independientes y combinados, a fin de determinar cómo estos elementos producen cambios en la actividad antioxidante, el perfil de compuestos fenólicos y el estrés oxidativo en plántulas de arándano. Para evaluar la actividad antioxidante se utilizaron técnicas como DPPH y FRAP, mientras que para cuantificar e identificar el perfil de compuestos fenólicos se empleó el uso de HPLC y LC-MS.

En general la aplicación de Al y Cd de manera independiente o combinada modificó significativamente los parámetros fisiológicos anteriormente mencionados, siendo posible observar la capacidad que posee las plántulas de arándano, para contrarrestar los efectos negativos de ambos metales en su desarrollo, con diferentes mecanismos de respuesta.

La aplicación de diferentes concentraciones de Al al medio de cultivo (100 y 200 μM), permitió evaluar el contenido de malodialdehido (MDA), la capacidad antioxidante y el contenido de compuestos fenólicos. Luego de 3 semanas de estudio, se observó un mayor contenido de MDA en los tejidos de las plántulas alcanzando un peak a los 14 días de exposición. En el mismo periodo de tiempo aumentó la respuesta antioxidante y se incrementó el contenido de los ácidos clorogénico y elágico (determinados por HPLC), que corresponden a los principales compuestos fenólicos presentes en el arándano.

En un ensayo de tres semanas de duración, se evaluó la respuesta antioxidante y la producción de compuestos fenólicos del arándano, debido a una aplicación inicial de Cd (50-100 μM) al medio de cultivo. Se observaron cambios significativos en la respuesta antioxidante mediante el ensayo FRAP y en el contenido de MDA, parámetro que aumentó durante el estudio, indicando que el Cd<sup>2+</sup> generó un daño oxidativo a las plántulas cultivadas *in vitro*. El principal compuesto fenólico identificado fue el ácido clorogénico, compuesto que vio aumentada su abundancia por efecto de la adición de Cd<sup>2+</sup>, también fue posible identificar otros compuestos de origen fenólico como el ácido vallínico y Quercetina (determinados por LC-MS). Es indudable que las variaciones en el perfil de compuestos fenólicos, se debe a la presencia de Cd<sup>2+</sup> en el medio de cultivo.

Se evalúo el efecto combinado (Al + Cd) en el estrés oxidativo, capacidad antioxidante y concentración de compuestos fenólicos en plántulas de arándano cultivadas *in vitro* durante 4 semanas. Los resultados indicaron que la aplicación conjunta de Al + Cd produjo estrés oxidativo, un incremento en la capacidad antioxidantes y en la concentración total de compuestos fenólicos. Los principales compuestos fenólicos que incrementaron su concentración después de la aplicación de los metales son los ácidos clorogénico y elágico.

De los resultados obtenidos en los diferentes estudios realizados en esta tesis podemos concluir que elementos trazas como Al y Cd inducen estrés oxidativo tanto de manera independiente como combinados, la cual correlaciona significativamente con la concentración total de compuestos fenolicos, lo que sugiere que estos compuestos tienen un rol importante en la respuesta frente al daño por estrés oxidativo. De acuerdo a lo anterior, la aplicación de elementos traza al medio de crecimiento produce cambios en el perfil y concentración de compuestos fenólicos, los cuales podrían ser modulados para aumentar la concentración de productos de interés cuando la planta se encuentra en un medio de crecimiento, controlando la adición de pequeñas concentraciones de elementos como Al y Cd.

#### **ABSTRACT**

The study of the effects of heavy metals in plants has been extensively described in the literature, where the effect has been investigated in various physiological and biochemical parameters. However, there is scant information on the possibility of modulating the profile of phenolic compounds with antioxidant response and other properties of scientific interest, from a change in the composition of culture medium or soil solution. In this context elements naturally present in soil or incorporated by anthropogenic activities (fertilization, application of amendments, etc.), as in the case of Al and Cd, emerge as an alternative study and analysis to develop methodologies for managements crop commercial interest in different matrices (soil, nutrient solution and/or *in vitro* culture), fostering the production of bioactive principles with pharmaceutical, nutritional and cosmetological properties.

The research developed in this thesis focused on the effects that can cause the incorporation of trace elements such as Al and Cd, on oxidative stress, antioxidant capacity and profile of phenolic compounds of blueberry seedlings cultured *in vitro*. The development of this study meant to evaluate the behavior of the blueberry, when this is subjected to different concentrations of Al and Cd in combination separate treatments and to determine how these factors cause changes in the antioxidant activity, the profile of phenolic compounds and oxidative stress in seedlings of blueberry. To evaluate the antioxidant activity was measured by using DPPH and FRAP, whereas to quantify and identify the profile of phenolic compounds using HPLC and LC-MS were used.

In general the application of Al and Cd independently or in combination significantly change the above physiological parameters, being possible to observe the capacity that blueberry seedling to counter act the negative effects of both metals in its development, with different mechanisms of response.

The application of different Al concentrations in the culture medium (100-200  $\mu$ M), and its relation with the content of Malondialdehyde (MDA), the antioxidant capacity and phenolics content was studied. After 3 weeks of study, a higher content of MDA was observed in tissues of seedlings reaching a peak at 14 days of exposure. In the same period of time increased the antioxidant response and the content of chlorogenic and ellagic acids (determined by HPLC), corresponding to the main phenolic compounds in blueberry increased.

In a three-week test period, the antioxidant response and the production of phenolic compounds was evaluated blueberry, due to initial application of Cd (50-100 mM) in the culture medium. Significant changes were observed in antioxidant response by FRAP assay and the content of MDA, parameter increased during the study, indicating that Cd<sup>2+</sup> generated oxidative damage to plantlets grown *in vitro*. The phenolic compound was identified principal chlorogenic acid, compound abundance was increased due to the addition of Cd<sup>2+</sup>, it was also possible to identify other phenolic compounds as vallinic acid source and quercetin (determined by LC-MS). It is clear that the profile of phenolic compounds change with the presence of Cd<sup>2+</sup> in the culture medium.

The combined effect of (Al +Cd) was evaluated in the oxidative stress, antioxidant capacity and concentration of phenolic compounds in blueberries plantlets cultured *in vitro* 

for 4 weeks. The results indicated that the combined effect of Al+Cd produced oxidative stress, an increase in antioxidant capacity and the total concentration of phenolic compounds. The major phenolic compounds that increased the concentration after the application of the metals are ellagic and chlorogenic acids.

From the results obtained indifferent studies in this thesis we can conclude that trace elements such as Al and Cd induce oxidative stress both independently and in combination, which correlated significantly with the total concentration of phenolic compounds, suggesting that these compounds have an important role in the response to oxidative stress damage. According to the above application of trace elements to the growth medium causes changes in the profile and concentration of phenolic compounds, which could be modulated to increase the concentration of products of interest when the plant is in a growth medium, controlling adding small concentrations of elements such as Al, and Cd.

#### **ABBREVIATIONS**

Al Aluminium element

Al<sup>3+</sup> Aluminium ion

≡Al-OH Aluminol

APX Ascorbate peroxidase

CAT Catalase

Cd Cadmiun element

Cd<sup>2+</sup> Cadmiun ion

CEC Cationic exchange capacity

DNA Desoxyribonucleic acid

DPPH 1,1-diphenyl-2-picrylhydrazyl

EDTA Etilendiamintetracetic acid

≡Fe-OH Ferrol

FRAP Ferric reducing antioxidant power

GSH Glutathione HM Heavy Metal

HPLC High performance liquid chromatography

H<sub>2</sub>O<sub>2</sub> Peroxide hidrogen

LC-MS Liquid Chromatography Mass Spectrometry

MDA Malondialdehyde

MRM Multiple reactions monitoring

OM Organic Matter

PAL Phenylalanine ammonia lyase

POD Peroxidase

ROS Reactive oxigen species
SOD Superoxide Dismutasa

≡Si-OH Silanol

TPC Total Phenolic Content

TPTZ Tripyridyltriazine
TEs Trace Elements

TSP Triple superphosphate

#### **OUTLINE DE LA TESIS**

La investigación reportada en esta tesis fue realizada en los laboratorios de Fisiología y Biotecnología Vegetal y Fisicoquímica de Suelos en Facultad de Química y Biología, Universidad de Santiago de Chile.

El capítulo I de esta tesis proporciona una visión general de los efectos de elementos trazas, principalmente Al<sup>3+</sup> y Cd<sup>2+</sup>, sobre diferentes parámetros fisiológicos determinados en distintos tipos de plantas y como propiedades fisicoquímicas del suelo, tales como, pH, contenido de arcillas, materia orgánica y óxidos de Fe, pueden modificar la disponibilidad de estos elementos, llegando incluso a una condición de fitotoxicidad. Este trabajo se preparó para ser próximamente enviado a *South African Journal of Botany* (**The Responses of Plants to Trace Elements. A Review Focusing on Aluminium and Cadmium**).

Las modificaciones en el perfil metabólico y cambios en parámetros fisiológicos, enfocado a estrés oxidativo y respuesta antioxidante, por cambios en el contenido de Al<sup>3+</sup>, se resumen en el capítulo II, de estos resultados se publicó un artículo en el *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas* (Effect of aluminum on antioxidant activity and phenolic compounds content in *in vitro* cultured blueberries).

El capítulo III se estudió el comportamiento del arándano sometido a diferentes concentraciones de Cd<sup>2+</sup> en un lapsus de tiempo de 3 semanas. Se estableció que este elemento genera especies reactivas de oxígeno (ROS por sus siglas en inglés), aunque el mecanismo es diferente a lo observado para el Al<sup>3+</sup> (capítulo II). Este trabajo sera enviado a la revista *Molecules*, bajo el título de "Effect of the availability of Cd<sup>2+</sup> on the non-enzymatic antioxidant response and the phenolic compounds profile of blueberry (*Vaccinium corymbosum* L.) plantlets cultivated *in vitro*".

El capítulo IV describe el efecto en la peroxidación lipídica (estrés oxidativo) y respuesta antioxidante del arándano cuando es sometido a medios de crecimientos enriquecidos con Al y Cd (sistemas binarios). Los resultados obtenidos serán enviado al journal *Environmental and Experimental Botany*.

#### **GENERAL INTRODUCTION**

The soil is the most important factor in a plant's growth and development. Depending on its origin and its physical, chemical and biological characteristics, it conditions the disposition and concentration not only of nutrients but also contaminants, which makes most of the plant's nutritive flows.

Acid soils are found throughout the world. It is estimated that about 40% of the world's soils are being used for agriculture and of that about 12% have pH below 5.5 (Guo et al., 2007). The soils in the central and south zones of Chile are of volcanic origin, which are characterized by having a high capacity of chemical elements retention of agricultural and environmental importance (nutrients, HM, pesticides). This type of soil has pH values under 6.0, as well as, a low concentration of interchangeable basis (Ca<sup>2+</sup>, Mg<sup>2+</sup>, K<sup>+</sup>, Na<sup>+</sup>), and as consequence has a high level of phytotoxic aluminum (Mora et al., 2002, 2009). In addition, this high acidity is accelerated by the high pluviometry in the zones/areas and the use of fertilizers with acid reaction (urea, ammonia phosphate). The use of fertilizers as source of nitrogen, phosphorus and potassium, among those that stand out as the most used which are urea, triple superphosphate (TSP), diamonic monoammonium phosphate, are important sources of trace elements. Various research has shown that the use of fertilizers, are the most important sources for incorporation of As, Zn, Cd, Pb, Cr Ni, Se, V and Cu to the soil (Chang and Page, 2000; Nicholson et al., 2003; Williams et al., 1973; Otero et al., 2005). By this, it can be shown that there is evidence of high contamination of trace elements to the soils via different ways, which are also affecting the organisms and plants that grow in these soils.

Amongst the elements that are often named because of how they affect the soils and the waters, include cadmium (Cd) for its high toxicity and mobility, which is shown in the soil/plant system (Sanita and Gabbrielli., 1999). The industrial, the agricultural activity with phosphatic fertilizers and biosolids have enriched the soils with this element (Zheljazkov *et al.*, 2005). In Chile, Bonomelli *et al.*, 2002 analyzed the presence of Cd in phosphatic fertilizers imported to Chile during the years 1999-2000, finding a high metal content in TSP type fertilizers. Similar results were obtained by Molina *et al.*, (2009) who found high concentrations of Cd in this type of fertilizers, concluding that the prolonged use of phosphatic fertilizers can increase the levels of HM, as well as, Cd in soils used for agriculture (Molina *et al.*, 2009).

In plants, the exposure to cadmium induced various photo-toxicity symptoms, such as chlorosis, biomass reduction, inhibition of root elongation, which kills them (Milone *et al.*, 2003; Lux *et al.*, 2011). It has been discovered that the presence of Cd generates free radicals which damage the plants tissue and depending on its concentration and the type of plant that is exposed to cadmium, this can inhibit or stimulate activity in various anti-oxidant enzymes, before the symptoms of toxicity are even visible (Fu and Huang., 2001; Liu *et al.*, 2007). Another side effect of the presence of cadmium is the fact that it is related to disruption in the capture and distribution of nutrients in the plants (Sandalio *et al.*, 2001; Rascio *et al.*, 2010).

The concentration of heavy metals affects the plants in different ways when they develop in an environment that is contaminated, because some plants possess a high tolerance to heavy metals and others that are more sensitive to them. The heavy metals induce the generation of species reactive to oxygen (ROS) which produce damage at the cell membrane level, lipid oxidation, proteins and DNA (Boscolo *et al.*, 2003; Arora *et al.*, 2002)

It has been described that plants show strategies that allows them to control the

species reactive to oxygen. One of these strategies is based in the production of secondary metabolites with anti-oxidant properties. The secondary metabolites are synthesized by plant as a defense mechanism against biotic stress. However, it is possible that when a plant is affected by the presence of a high concentration of heavy metals, when these metabolites play a role in the physiological function of said plant (Close and McArthur, 2002; Keilig *et al.*, 2009).

Of the different bioactive secondary metabolites, we can find the phenolic compounds, which make up a vast group of metabolites, with different activity and chemical structure. Nowadays, these compounds require more attention due to their contribution to health, how many of the beneficial properties found in plants and vegetables, especially linked to anti-oxidant activities, are tightly linked to the presence and content of secondary metabolites of phenolic nature (Vanisree *et al.*, 2004; Jung *et al.*, 2007; Seeram *et al.*, 2005).

The antioxidant activity shown by phenolic compounds, is of interest from the technological and nutritional point of view, for example the phenolic compounds intervene as natural antioxidants in food, which reduces the use of synthetic additives and from the nutritional point of view the antioxidant compounds are associated with the role of protecting against cancer, cardiovascular problems and the aging process (Sakihama *et al.*, 2002; Niggeweg *et al.*, 2004; Prasad *et al.*, 2009).

The higher demand of plants with antioxidant properties and bioactive compounds has generated an increase in production, beginning to be an important part in today's agricultural production, which means that plants are being cultivated in places where soil handling, which in part means that fertilizers and pesticides are being used and applied in order to improve the fertility and property of the soils.

The blueberry is a minor fruit tree native to North America, considered among the berries group, which was introduced in Chile in the nineteen eighties. In Chile, the farming of blueberry is mainly concentrated between the VI and the X regions. The tall blueberry (*Vaccinium corymbosum* L) grows very well in soils with pH between 4.8 and 5.5, as well as, it is very well valued in different countries by its antioxidant and antibiotic properties. Due to the fact that in the last few years this species has positioned itself in the country's agriculture, it is believed that it is important to cultivate and handle the growth conditions for this plant, so it will allow us to evaluate the factors directly related to the production of secondary metabolites and its antioxidant properties.

Therefore, considering the development that the blueberry has had in our country and the importance that it has generated in the last few years due to the contributions this fruit provides to our health because of its antioxidant properties, it is important to research the effects that handling the soil has (by incorporating Cd and Al in mono and multicomponent system) on the production of secondary metabolites of phenolic nature, by cultivating plants thru *in vitro*, conditioning the previously mentioned factors, in order to determine its real effect and to be able to modulate the plant in a line of interest.

#### REFERENCES

- 1. Arora A., Sairam R. K., and Srivastava G.C., 2002. Oxidative stress and antioxidative system in plants. Current Science 82, 10-25.
- 2. Boscolo, P., Menossi, M., Jorgea, R., 2003. Aluminum-induced oxidative stress in maize. Phytochemistry 62, 181-189

- 3. Chang, A.C. and Page, A.L., 2000. Trace element slowly accumulating, depleting in soils. California Agriculture 54, 49-55.
- 4. Fu, J., Huang, B., 2001. Involvement of antioxidants and lipid peroxidation in the adaptation of two cool-season grasses to localized drought stress. Environmental and Experimental Botany 45, 105-114.
- Guo, T. R., Zhang, G.P.and Zhang, Y.H., 2007. Physiological change in barley plants under combined toxicity of aluminum, copper and cadmium. Colloids and Surface B: Biointerfaces 57, 182-188.
- Jung, M.J., Heo, S., Wang, M.H., 2008. Free radical scavenging and total phenolic contents from methanolic extracts of *Ulmus davidiana*. Food Chemistry 108, 482-487.
- 7. Keilig. K., Ludwuig-Muller, J., 2009. Effect of flavonoids on heavy metal tolerance in *Arabidopsis thaliana* seedlings. Botanical Studies 50, 311-318.
- 8. Liu, X. L., Zhang, S. Z., Shan, X. Q., Christie, P., 2007. Combined toxicity of cadmium and arsenate to wheat seedlings and plant uptake and antioxidative enzyme responses to cadmium and arsenate co-contamination. Ecotoxicolology and Environmental Safety 68, 305–313.
- 9. Lux, A., Martinka1, M., Vaculík, M., White, P., 2011. Root responses to cadmium in therhizosphere: a review. Journal of Experimental Botany 62, 21–37.

- 10. Milone, M.T., Sgherri, C., Clijsters, H., Navari-Izzo, F., 2003. Antioxidative responses of wheat treated with realistic concentration of cadmium. Environmental and Experimental Botany 50, 265-276
- 11. Molina, M., Aburto, F., Calderón, R., Escudey, M., Cazanga, M., 2009. Trace element composition of selected fertilizers used in Chile with special regard to phosphorus fertilizers. Soil and Sediment Contamination 18, 497-511
- 12. Mora, M., Cartes, P., Demanet, R., Cornforth I. S., 2002. Effect of lime and gypsum on pasture growth and composition on acid Andisol in Chile, South America. Communications in Soil Science and Plant Analysis 33, 2069-2081.
- 13. Mora, M.L., Rosas, A., Ribera, A., Rengel, Z., 2009. Differential tolerance to Mn toxicity in perennial ryegrass genotypes: involvement of antioxidative enzymes and root exudation of carboxylates. Plant Soil 320, 79-89.
- 14. Nicholson, F.A., Smith, S., Alloway, B. J., Carlton-Smith, C., Chambers, B.J., 2003. An inventory of heavy metals inputs to agricultural soils in England and Wales. The Science of the Total Environment 311, 205-219.
- 15. Niggeweg, R., Michael, A.J., Martin, C., 2004. Engineering plants with increased levels of the antioxidant chlorogenic acid. Nature Biothecnology 22, 746-54.
- 16. Rascio, N., and Navari-Izzo, F., 2011. Heavy metal hyperaccumulating plants: How and why do they do it? And what makes them so interesting? Plant Science 180, 169-181.

- 17. Sandalio, L.M., Dalurzo, H.C., Gomez, M., Romero-Puertas, M.C., del Río, L.A., 2001. Cadmium-induced changes in the growth and oxidative metabolism of pea plants. Journal of Experimental Botany 52, 2115-2126.
- Sanitá di Toppi, L., Gabbrielli, R., 1999. Response to cadmium in higher plants.
   Environmental and Experimental Botany 41, 105-130.
- 19. Smeets, K., Cuypers, A., Lambrechts, A., Semane, B., Hoet, P., Laerve, A.V., Vangronsveld, J., 2005. Induction of oxidative stress and antioxidative mechanisms in Phaseolus Vulgaris after Cd application. Journal Plant Physiology and Biochemistry 43, 437-444.
- Otero, N., Vitoria, L., Soler, A., Canals, A., 2005. Fertiliser Characterisation:
   Major, Trace and Rare Earth Elements. Applied Geochemistry 20, 1473-1488.
- 21. Prasad, K. N., Yang, B., Yang, S., Chen, Y., Zhao, M., Ashraf, M., Jiang, Y., 2009. Identification of phenolic compounds and appraisal of antioxidant and antityrosinase activities from litchi (*Litchi sinensis Sonn.*) seeds. Food Chemistry 116, 1–7.
- 22. Sakihama, Y., Cohen, M.F., Grace, S.F., Yamasaki H., 2002. Plant phenolic antioxidant and prooxidant activities: phenolics-induced oxidative damage mediated by metals in plants. Toxicology 177, 67–80.
- 23. Seeram, N.P., Lee, R., Sheuller, S., Heber, D., 2005. Identification of phenolic compound in strawberries by liquid chromatography ionization mass spectroscopy. Food Chemistry 97, 1-11.

- 24. Vanisree, M., Hsin-Sheng, T., 2004. Plant Cell Cultures An Alternative and Efficient Source for the Production of Biologically Important Secondary Metabolites. International Journal of Applied Science and Engineering 2, 29-48.
- 25. Williams, C.H., David, D.J., 1973. The Effect of Superphosphate on the Cadmium Content of Soils and Plants. Australian Journal of Soil Research 11, 43-56.
- 26. Zheljazkov, V. D., Craker, L. E., Baoshan, X., 2005. Effects of Cd, Pd, Cu on growth and essential oil contents in dill, pepermint, and basil. Environmental and Experimental Botany 58, 9-16.

#### **HYPOTHESIS**

Taking into account the high phytotoxic Al contents found in the acid soils and the toxicity that shows Cd on the plants, both metals will induce oxidative stress in the blueberry plants, stress characterized by incresing levels of secondary metabolites of phenolic nature.

#### GENERAL OBJECTIVE

To evaluate, through *in vitro* cultivation, the impact of Cd and A1 in the production of secondary metabolites of phenolic origin in blueberry.

#### **SPECIFIC OBJETIVES**

- 1) To evaluate the effects of Cd and A1, in the oxidative status in *Vaccinium* corymbosum cultivated in vitro
- 2) To determine the effect of Cd and Al in the antioxidant capacity of ethanolic extracts in the cultivation of *V. corymbosum* cultivated *in vitro*.
- 3) To determine the effect of Cd and Al in the phenolic metabolic in ethanolic extract of *V. corymbosum* cultivated *in vitro*.
- 4) To establish the combined effect of Al-Cd toxicity over the oxidative stress and production of secondary metabolites in the different crops.

# **Chapter 1**

The Response of Plants to Trace Elements. A Review

Focusing on Aluminium and Cadmium

#### **Abstract**

Contamination with trace elements has increased in recent times as a direct consequence of industrialization and technification at the world level. The different sources of trace elements incorporation in the environment include production sectors such as mining, agriculture, and industry which alter the normal conditions of these elements in the soil and irrigation water, consequently increasing the total content and potentially the bioavailability of heavy metals to plants. Many factors affect trace elements bioavailability in soil, being the most relevant; the total metal concentration, pH, redox conditions, organic matter, clays and hydrated oxides.

Trace elements concentration affects in different ways the plants that grow in a contaminated environment, because there are plants that have high tolerance for heavy metals and others that are sensitive to them. Heavy metals induce the generation of reactive oxygen species (ROS) that cause damage at the cell membrane, lipid oxidation, protein, and DNA levels. It has been described that plants have strategies that allow them to control ROS. One of these strategies is based on the production of secondary metabolites having anti-oxidant properties. The phenolic compound are synthesized by the plants as a method of defense against biotic stress, but it is also possible that they play a role in the physiologic functioning of the plants when they are in the presence of a high concentration of heavy metals. This review shows the complexity of the toxicity Mechanisms of trace elements, such as changes in the availability of free species of heavy metals can induce oxidative stress by multiple mechanisms, from the generation of ROS until inhibition of some enzymes

Keywords: Heavy metals, Lipid peroxidation, Phenolic Compound, ROS, Soil, Plant

#### Introduction

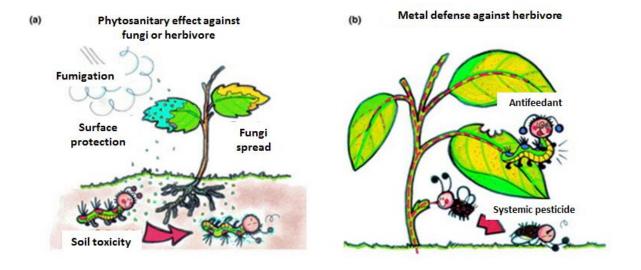
Soil contamination by heavy metals (HM) is considered one of the main problems affecting both developing and developed countries (Chang and Page 2000). In recent years, the soil composition has changed due to man-caused activities. Industrial, agricultural residues and mining activities have led to an increase in the concentration of heavy metals (HM) in soils (Li *et al.*, 2008, Boularbah *et al.*, 2006).

The HM as Cr, Cd, Pb and others are called trace elements (TE), because the quantities that are in traces in environmental matrices. These elements are not biodegradable and they easily accumulate in the soil, becoming harmful to plants, animals and humans when their concentrations are above a critical level. Among the different trace elements we can find a group of them which are essential micronutrients for plants (Fe, Mn, Cu, Co, Zn, Ni, Mo), due to their involvement in various metabolic processes (Salt *et al.*, 1998; Schutzendubel and Polle 2002). Then, there are those that are considered as non-essential (Pb, Cr, Cd, Hg, Al, etc.) and they are potentially toxic to plants (Schutzendubel and Polle 2002). The behavior and speciation of the HM in soil depends on various factors such as organic matter, moisture, aeration, biological activity, redox potential, pH and others, which determine the shape and therefore the bioavailability and mobility of HM (Bonomelli *et al.*, 2002).

The heavy metals can physiologically inhibit processes such as respiration, photosynthesis, cell elongation, capture water and nutrients. When a plant has physiological or visible symptoms of toxicity by HM, it is the last link in a chain of events that caused an increase of these contaminants to phytotoxic levels in the environment (Scora and Chang, 1997). These events are conditioned in two steps: 1) the incorporation of heavy metals in groundwater and soil through various natural and anthropogenic sources, standing out as

the most important sources of pollution to highly productive sectors such as agriculture, mining and industry (Li *et al.*, 2008, Boularbah *et al.*, 2006) and 2) the deployment by the plant's mechanisms of absorption of nutrients (radical exudates), which indirectly incorporates HM and trace elements being these translocated to various parts of the plants, altering its different states and physiological processes (Larcher, 2003; Meyer *et al.*, 2000; Poschenrieder *et al.*, 2006) (Figure 1).

Plants absorb essential and non-essential elements for its life cycle from soils, in response to the concentration gradients induced by selective ion absorption by the roots or by diffusion of ions in the soil. The levels of accumulation of elements in plants differ within the same species or between species (McGrath *et al.*, 2002).



**Figure 1.** Different ways for high metal concentrations to protect plants against biotic stress. (a) Metal-containing compounds are used by farmers in phytosanitary treatments as leaf sprays or soil amendments. (b) According to the metal defense hypothesis, the high metal concentrations that accumulate in leaf tissues of metal hyper accumulators either deter (antifeedant) or intoxicate (plant systemic pesticide) herbivores or pathogens. (Taken from Poschenrieder *et al.*, 2006).

Plants may adopt different strategies against the presence of metals in the environment (Baker, 1981; Gunsé *et al.*, 2003). Some based their resistance to metals with the strategy of an efficient exclusion of metal, restricting transport to the aerial part. Others accumulate the metal in the aerial part in a non-toxic way to the plant. The exclusion is more characteristic of sensitive and tolerant species to metals, and accumulation is more common in species that always appear in contaminated soils (Baker, 1981).

Among the mechanisms to deal with the toxicity by HM, are those that limit uptake of metals by the roots, helping the plant to tolerate some amount of toxic elements using organic acids exudation which chelate metal in the rhizosphere, preventing this to be absorbed by the root (Schutzendubel and Polle, 2002). Tolerance to heavy metals is also reinforced by the action of mycorrhizal fungi present in some types of plants (Schutzendubel and Polle, 2002).

The heavy metals can be transported via apoplast and being immobilized on the cell walls. The HM becomes a threat to the plants, mainly when they reach the cell cytosol. Therefore, the ability of root cells to control the transport of heavy metals through the membranes determines tolerance by plants.

Three molecular mechanisms associated with the toxicity of heavy metals have been described: the production of reactive species to the oxygen (ROS), the blocking of essential functional groups and the displacement of metal ions in biomolecules (Schützendübel and Polle, 2002).

The objectives of this work are showing the impact and the effect of heavy metals in plants, analyzing the incorporation of these contaminants routes to the different substrates and its impact on the production of secondary metabolites related with the degree of tolerance from a general perspective.

#### **SOIL AND METALS**

#### Factors affecting metal availability

The trace elements pools present in soils are not fully available for the growth of plants. The total traces elements pool present in the soils can be classified into four categories; 1) the solubilized elements in the soil solution, 2) those which form precipitates, 3) the slurp in clays, oxides of Fe and organic matter and 4) those who are part of the soil parent material, these fractions are in a constant dynamic equilibrium with each other (Norvell, 1991; Davis and Leckie, 1978). These equilibria may be strongly influenced by the physicochemical characteristics of the soil, where the plant availability of nutrients, micronutrients and trace elements in general depend on the total content of these present in soil, pH and clay Fe oxide and organic matter, in addition to the redox conditions (Fageria *et al.*, 1991; Marschner, 1995; Whitehead, 2000).

Research on volcanic soils treated with mud and sludge ashes and oriented to determine the retention capacity of heavy metals (Cu, Zn, Cd, Pb, Mo and Mn), showed that the percentage of heavy metals that are retained in the soil varies between 96.0 to 99.9% (Escudey *et al.*, 2007; Pinochet *et al.*, 2002).

However in the rhizosphere (region of soil that is subject to the influence of the roots and is characterized by an area of high microbial activity), chemical and biological conditions are quite different from the "bulk" of the soil, therefore the bioavailability of heavy metals depend on the physicochemical nature of the soil, which governs both the sorption and mobility of these elements in depth and the strategies of plants to capture nutrients (Table 1), mainly through root exudates, which include short-chain organic acids (Legrand *et al.*,

2005; Dakora and Phillips, 2002, Lexmond, 1980; Sauve *et al.*, 1996; McBride *et al.*, 1997; Sauve *et al.*, 1997; Peijnenburg *et al.*, 2000).

**Table 1**.Essential nutrient elements used by plants: element, symbol and primary chemical forms.

Element	Symbol	Primary Forms Used by Plants
NON-MINERAL ELEMENTS		
		$\mathrm{CO}_{2(\mathrm{g})}$
Carbon		-16/
Hydrogen	H	$ m H_2O$ $_{ m (l)},  m H^+$
Oxygen	O	$H_2O_{(l)}, O_{2(g)}$
MINERAL ELEMENTS		
Nitrogen	N	$\mathrm{NH_4}^+,\mathrm{NO_3}^-$
Phosphorous	P	$HPO_4^{2-}, H_2PO_4^{-}$
Potassium	K	$K^{+}$
SECONDARY NUTRIENTS		
Calcium	Ca	$\mathrm{Ca}^{2+}$ $\mathrm{Mg}^{2+}$ $\mathrm{SO_4}^{2-}$
Magnesium	Mg	$\mathrm{Mg}^{2+}$
Sulfur	S	$\mathrm{SO_4}^{2^-}$
MICRONUTRIENTS		
Iron	<del></del> Fe	$Fe^{3+}, Fe^{2+}$
Manganese	Mn	$\mathrm{Mn}^{2+}$
Zinc	Zn	$\mathrm{Zn}^{2+}$
Copper	Cu	$\mathrm{Cu}^{2+}$
Boron	В	B(OH) <sub>3</sub> <sup>0</sup> (Boric Acid)
Molybdenum	Mo	$\mathrm{MoO_4}^{2^-}$
Chlorine	Cl	Cl <sup>-</sup>

### <u>pH</u>

The balance between the different trace elements speciation, its solubility, adsorption or exchange occurring with the sites present in the soil solid phase, it is intimately related to the soil pH value (Olomu *et al.*, 1973; Kalbasi *et al.*, 1978; Cavallaro and McBride, 1984; Sauve *et al.*, 1997). Several studies have shown that minor variations in this parameter

significantly affect the bioavailability of trace elements (Turner, 1994; McBride *et al.*, 1997), such as Mn, Cu, Zn, Cd and the in which the percentage of their free species are strongly affected by the pH of the soil (Fergus, 1954; McGrath *et al.*, 1988; Turner, 1994; Kalbasi *et al.*, 1978; McBride, 1982; Bar-Tal *et al.*, 1989 ;Msaky and Calvet, 1990; Sauve *et al.*, 1997; Jeffery and Uren, 1983). Studies in Arachishypogaea (peanut), Phaseolusvulgaris (beans) and Vignaun-guiculata (Cowpea) grown in contaminated soils with Zn<sup>2+</sup>, and Mn<sup>2+</sup> a phytotoxic response was observed, with a significant increase in these elements shoots, as pH decreased (Parker *et al.*, 2001; Davis-Carter and Shuman, 1993; Fergus, 1954; Vega *et al.*, 1992).

Other sensitive to pH changes in soil metals are Al<sup>3 +</sup> and Cd<sup>2 +</sup>, which are also affected in their speciation, solubility, complexation and adsorption (Payne and Pickering, 1975; Msaky and Calvet, 1990; Reddy *et al.*, 1995, Tolrá *et al.*, 2005; Guo *et al.*, 2007). In the case of Al, it is noteworthy that this metal is a constituent of the inorganic fraction of the soil, forming part of aluminosilicates (Escudey *et al.*, 2007).

The availability of  $Al^{3+}$  in soil solution is favored under conditions of pH less than 5.5, where the predominant species are  $Al^{3+}$ ,  $Al(OH)^{2+}$ ,  $Al(OH)^{2+}$ , but above pH 7.5 the dominant species is  $Al(OH)_4^-$  (McBride, 1994). Under conditions of pH less than 5.5 the Al rapidly alter the plant growth affecting mainly the root elongation, which hinder the capture of water and nutrients, whereby a concentration of free Al is considered one of the major factors limiting crop growth in acid soils (Tolrá *et al.*, 2005; Guo *et al.*, 2007).

The application of fertilizers such as triple superphosphate (TSP), has been one of the main inputs of Cd in soils (Molina *et al.*, 2010; Cordero *et al.*, 2004). Cadmium availability

strongly decreases at pH values above 6, where it starts to precipitate as hydroxide (Barančiková *et al.*, 2004). Cadmium also forms precipitates with HPO<sub>3</sub><sup>2-</sup> and CO<sub>3</sub><sup>2-</sup>, while variations in the concentrations of Ca<sup>2+</sup> and Cl<sup>-</sup> are impacting the availability in soil (Dudley *et al.*, 1988; 1991). In general in acid conditions the solubility increases Cd, where the ionic species (Cd<sup>2+</sup>) of this element predominates, however the potential reduction of the Cd<sup>2+</sup> is unable to generate ROS through Fenton reaction (Cuypers *et al.*, 2010). Studies on soybean cultivated in nutrient media enriched with Cd, determined that this metal is phytotoxic inducing oxidative stress by multiple mechanisms (Balestrasse *et al.*, 2004; Sandalio *et al.*, 2001; Balestrasse *et al.*, 2001).

# Organic matter

The type and content of organic matter (OM) play a key role in complexation of metal ions present in soil solution, affecting availability for plants. This complexation can occur by different functional groups presenting the organic matter (Baker and Senft, 1995), such as:

- Nitrogenous aromatic groups, for example pyrrole and indole.
- Aromatic groups: as naphthalene and benzene.
- Reactive groups: primarily responsible for the properties of organic matter: hydroxyl, carboxyl, amino.

The interaction of the organic matter with the Al<sup>3+</sup>, has been extensively studied (Stevenson, 1991; Garrido and Matus, 2012). Bioavailability and potential Al toxicity in soils and waters are highly dependent on chemical interactions with natural organic matter. There are generally two types of binding mechanisms in which interact different forms of Al with OM; those of electrostatic type occurring mainly between Al<sup>3+</sup> and dissociated

functional groups of organic matter, mainly -COOH, -OH and-NH<sub>2</sub>, besides the Al can form specific bonds with one or more specific functional groups through coordination with aromatic groups (Tam, 1987).

There are few studies that account for the interaction of Cd with OM compared to other heavy metals such as Cu. However, OM also affects the bioavailability of Cd in soils (Barančiková *et al.*, 2004). Soils with high content in OM are able to absorb up to 30 times more than clay soils, where the main interactions occur with Cd ionizable groups of fulvic acids (Barančiková *et al.*, 2004). It has been reported that this element has a high affinity for sulfhydryl groups, hydroxyl and those ligands that contain nitrogen (Benavides *et al.*, 2005).

# Clays and hydrous oxides

Clay and hydrated oxides of Al and Fe have ionizable sites silanol ( $\equiv$ Si-OH), aluminol ( $\equiv$ Al-OH) and ferrol ( $\equiv$ Fe-OH) which play an important role in the availability of metals present in the soil solution, through different mechanisms where the processes of sorption (adsorption, absorption and precipitation) (Miller *et al.*, 1987; Pampura *et al.*, 1993; Kalbasi *et al.*, 1978; Basta and Tabatabai, 1992; Martínez and McBride, 1998). Despite the scarce studies that consider the interactions that occur between the inorganic fraction of soil and Al<sup>3+</sup> and Cd<sup>2+</sup>, it is possible to establish that increases in clay levels and hydrated oxides, by anthropogenic or natural factors, provides more places for the adsorption of these, thus reducing their availability (Ghanem and Mikkelsen, 1988; Barrow, 1993; Qiao and Ho, 1996).

### **METALS AND PLANTS**

Plants have evolved a number of mechanisms for obtaining nutrients present in the soil and translocating each of its components, but these processes are not specific, and therefore the plant also includes non-essential elements (Redjala *et al.*, 2009).

### Uptake mechanisms

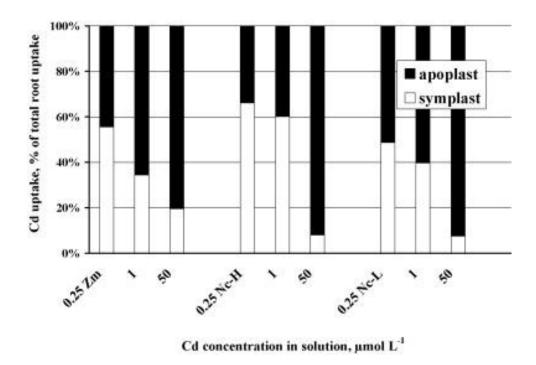
The roots are responsible for the uptake of essential and non-essential elements in plants. This process involves a sequence of steps, not fully understood, allowing the transfer of these from the soil solution to the root surface and then into the root cells (Laurie and Manthey, 1994; Fan *et al.*, 1997). The complexity of the absorption processes of elements present in the soil, due to the constant changes that occur between the rhizosphere soil solution and microorganisms (Laurie and Manthey, 1994).

### Bio-available metals supply for plant uptake.

Plants are able to influence the solubility and speciation of metals that are found in the rhizospheric area, either by the exudation of chelating agents or pH variations agents (Welch, 1995; Fan *et al.*, 1997). The uptake of the elements inside the plants can occur through the xylem and phloem, where the composition and plants age have an important role in ions movement speed and its toxicity (Fan *et al.*, 1997).

### Mechanisms of incorporation of Al and Cd in plants

Incorporation mechanisms of essential and nonessential elements within the plant occur through water transportation (Redjala *et al.*, 2009) (Figure 2).



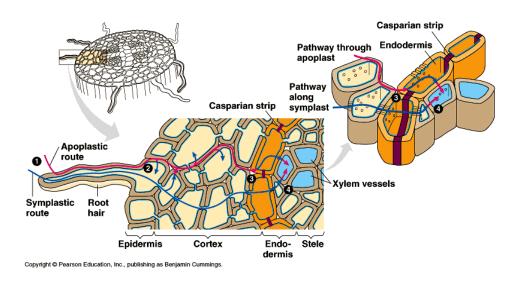
**Figure 2**. Contribution of apoplast and symplast to Cd uptake by roots of maize (Zm) and high- (Nc-H) and low- (Nc-L) Cd accumulating ecotypes of *Noccaea caerulescens* (Taken from Redjala *et al.*, 2009).

This step may occur through apoplast or symplast pathway (Figure 3).

- The apoplast pathway refers to the passage of water transversely through the cell walls and intercellular spaces.
- The symplast pathway refers to the passage of water into tissues transversely through cell walls and cytoplasm of cells.

In the case of aluminum, plants compartmentalize in vacuoles, once it was absorbed, and thereby reduce its phytotoxic effect. Silva *et al.*, 2002, suggests that the main aluminum transport mechanism occurs via apoplast (extracellular) where the net negative charge of the roots caused by the deprotonation of the carboxylic groups of polygalacturonic acid, the

main component of the cell wall and the outer surface of the membrane, plasma, leads to increased cationic exchange capacity (CEC) of the roots, facilitating the incorporation of Al (Valencia *et al.*, 2012).



**Figure 3.** Lateral transport of minerals (micronutrients) and water in roots (Taken from White, 2001)

However, inside the plant, cells have a lower content of pectin and high degree of methylation, which reduces the CEC and the binding of Al<sup>3+</sup> to the cell wall (Silva *et al.*, 2002), having as a consequence a lower accumulation of Al<sup>3+</sup> to the interior of the cell (via symplast). In soils with high contents of free Al, the roots of the plants are characterized by a low CEC, being this adaptation a defense mechanism (Blamey *et al.*, 1990).

Studies in *Pisium sativum* L., suggest that Cd is accumulated in descending order in roots, stems, leaves, fruits and seeds (Blumm, 1977). In general, once that Cd enters to the root can move through the xylem via the apoplast and/or symplast forming complexes with nitrogen and sulfur donors compounds present in macromolecules and low molecular weight ligands, which were possibly biosynthesized in response to the presence of Cd<sup>2+</sup>

(Clemens *et al.*, 2002). Since Cd can enter the plant via apoplast and / or symplast, this element can accumulate in other organelles compared to what happens with Al<sup>3+</sup> (Godbold and Huttermann, 1985; Frausto da Silva and Williams, 2001), the symplastic pathways of Cd incorporation in plants are closely linked with those carrying Zn<sup>2+</sup> and Ca<sup>2+</sup>, so that variations in the concentrations of these nutrients (Huang *et al.*, 1996; Guerinot *et al.*, 2000).

## Effect of heavy metals in plants: phytotoxicity symptoms

Elements such as Cu, Zn, Fe, Mn, Mo, Ni and Co are essential micronutrients to metabolic development of plants, but an excess of them in soils can be toxic for their development (Monni *et al.*, 2000). These elements along with Cr, Cd metals, Pb and others, are called trace elements, because the quantities that are in traces (10 mg kg<sup>-1</sup> or mg ml<sup>-1</sup>) or ultra traces (1 μg kg<sup>-1</sup> or μg L<sup>-1</sup>) in environmental matrices (Monni *et al.*, 2000).

The most widespread visual evidence of the toxicity of metals is a reduction in the growth of plants as the metal toxicity increases. However, as different metals have different sites of action within the plant, the response of the toxicity in visual form differs among metals.

# Effects of cadmium on plants

The Cd is highly toxic to humans, animals and plants. The Cd limits in agricultural soil is 100 mg kg<sup>-1</sup> of soil. In the plants exposure to Cd produces various symptoms of phytotoxicity, such as chlorosis, reduction of the biomass, inhibition of roots elongation and even death of the plant (Sanita di Toppi and Gabbrielli, 1999; Milone *et al.*, 2003; Guo *et al.*, 2008).

The presence of Cd in the growth medium of the plant interferes with incorporation, transport and uptake of essential elements such as Ca, Mg, K, and P, causing imbalances in the plant metabolism (Sandalio *et al.*, 2001; Li *et al.*, 2008). The Cd also reduces the absorption of nitrates and their transport from the root to the stem, and also to inhibit nitrate reductase activity in stems (Gouia *et al.*, 2000). Plants exposed to Cd show changes in stomatal aperture, photosynthesis and transpiration (Sandalio *et al.*, 2001). Studies have shown that phytotoxicity by Cd produces chlorosis due to a deficiency of Fe, PO<sub>4</sub><sup>3</sup>-or reduced transport of Mn (Liu *et al.*, 2007; Benavides *et al.*, 2005; Gaballah *et al.*, 2012).

The photosynthetic apparatus is one of the target sites of Cd action in plants. The Cd can directly or indirectly interact with different components of the photosynthetic apparatus and can reduce the efficiency of electron transport inhibiting chlorophyll biosynthesis, leading to a reduction of CO<sub>2</sub> assimilation (Maksymiec *et al.*, 2007; Wang *et al.*, 2006). Other effects on the metabolism of the plant due to Cd are reduced ATPase activity of the plasma membrane, changes in the functionality of the plasma membrane (Fodor *et al.*, 1995), which has been observed in roots of sunflower and imbalances in chloroplast metabolism, inhibiting chlorophyll synthesis and reducing the activity of enzymes involved in CO<sub>2</sub> fixation (Sandalio *et al.*, 2001; Astolfi *et al.*, 2005; Maksymiec *et al.*, 2007). It has been found that the presence of Cd in plants increases ROS production, causing oxidative stress and increased lipid peroxidation in leaves and roots (Smeets *et al.*, 2005; Guo *et al.*, 2007; Cherif *et al.*, 2011).

#### Effect of Aluminum on the plant

Phytotoxicity by Al<sup>3+</sup>, is shown especially in acid soils, where it is one of the main factors affecting culture production. The first symptoms of phytotoxicity by Al<sup>3+</sup> are reduced elongation and cell division of roots in plants, which is reflected in a decrease in the absorption of nutrients and water (Miyasaka *et al.*, 1991). The concentration of Al<sup>3+</sup> in plants grown under the presence of metal is greater in roots than in the aerial part. Symptoms of phytotoxicity of Al<sup>3+</sup> have been observed in *Triticum aestivum*, where a decrease in root length after one hour exposure of the roots to the metal, together with an increase in the diameter of the root tips was observed (Ryan *et al.*, 1993).

Severe phytotoxicity Al<sup>3+</sup> reduces and damages the roots, making the plant susceptible to water stress and nutritional deficiencies. Among the nutritional effects, include blocking calcium and potassium channels, which cause long-term mineral deficiencies. It has been reported that phytotoxic concentrations of Al decrease the flow of oxygen in the roots; interfere with certain enzymes involved in the production and deposition of constituent polysaccharides of the cell wall, altering Donnan free space in cell wall and forms highly stable complex with amino acids and nucleic acids (Rout *et al.*, 2001).

The Al<sup>3+</sup> interferes with the absorption and transport of nutrients such as Ca, Mg, K, P, B, Fe and Cu (Keltjens and Tan, 1998; Lukaszewski and Blevins, 1996; Taylor *et al.*, 1998; Lidon *et al.*, 1992; Guo *et al.*, 2004, 2007; Olivares *et al.*, 2009). The decrease in the uptake of cations may be related to the inhibition of root growth and the tolerance of cultures to Al<sup>3+</sup>. Giannakoula *et al.*, 2008; studied sensitive lines of rice and tolerant to

aluminum, finding those lines sensitive to accumulate more of that the tolerant line, which largely retains the concentration of Ca, Mg and K.

### Oxidative stress

Aerobic organisms are continuously exposed to the generation of reactive oxygen species (ROS). Many of the lethal processes undergone by plants under adverse conditions are mediated by ROS, which are generated in different cellular compartments as a result of malfunction of normal metabolic pathways and physiological processes (Arora *et al.*, 2002, Fu and Huang, 2001) (Table 2).

Within the group of ROS we can find free radicals (chemical species with one or more unpaired electrons in its outer orbit) and superoxide anion (·O<sub>2</sub><sup>-</sup>) and hydroxyl radical (·OH) and generating molecules of ROS such as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), and singlet oxygen (¹O<sub>2</sub>) (Arora *et al.*, 2002). ROS are produced in various parts of the cell, some of the sites of ROS production and components of the antioxidant system of the cell are summarized in Table 2 (Arora *et al.*, 2002). Excessive generation of ROS occurs under stress conditions such as UV light, water and salt stress, pathogen attack, injury, cold, heavy metals (Merwin *et al.*, 1995; Ercal *et al.*, 2001). Research has shown that ROS produce damage to cell membranes by inducing lipid peroxidation, besides cause damage to the level of DNA, proteins, lipids and chlorophyll (Boscolo *et al.*, 2003; Cakmak *et al.*, 1991; Guo *et al.*, 2007; Gaballah *et al.*, 2012).

**Table 2**. Main sites of production of reactive oxygen species (ROS) in plants and the respective detoxification mechanisms ('scavengers').

Reactive Oxygen Species (ROS)	<b>ROS Production</b>	Scavenger
Hydrogen peroxide	Cellular compartments (chloroplasts, mitochondria, cytosol) Peroxisome (photorespiration)	Catalase, Ascorbate peroxidases, Glutathione
Superoxide Radical	Cellular compartments (chloroplasts, mitochondria, cytosol)	Superoxide dismutases, Ascorbic Acid, Glutathione
Singlet oxygen	Thylakoid	α-Tocopherol, Carotenoids
Hydroxyl radical	Several Sites (eg. Haber- Weiss Reaction in chloroplast)	Ascorbic Acid

Plants growing in soils with HM suffer from reduced growth affecting culture yields. At the molecular level, the oxidative stress is widely studied as a key sign of plant stress. This process is described as an imbalance between ROS and antioxidant systems (Boscolo *et al.*, 2003). Dietz *et al.*, (1999) propose a relationship between the toxicity of metals, redox homeostasis and antioxidant capacity in plants. Depending on the chemical and behavior of markers in biological systems, the toxicity may be attributed to properties of the following mechanisms: (1) interfering with the functional sites in proteins, (2) transport of essential elements, with consequent disturbance of the enzymatic functions, and (3) an onset of ROS production (Dietz *et al.*, 1999).

Metals such as  $\text{Fe}^{2+}$  and  $\text{Cu}^+$ , can convert the molecule  $H_2O_2$  in  $\cdot$  OH radical through the Fenton type reaction.

$$H_2O_2 + Fe^{2+} (Cu^+) \rightarrow Fe^{3+} (Cu^{2+}) + \cdot OH + OH^- (Eq.1)$$

Other metals are not involved in the Fenton reaction, however; also generate ROS, as for example the Hg<sup>2+</sup> it is believed that it can inhibit antioxidant enzymes, particularly the enzyme glutathione reductase, tending as a result the accumulation of ROS (Mithofer *et al.*, 2004). Cadmium does not cause directly ROS through fenton reaction, however can cause indirectly oxidative stress by activating the enzyme Lipoxygenase, which stimulates lipid peroxidation (Keunen *et al.*, 2011). Other studies suggest that the metal may act as pro-oxidant, by reducing the pool of glutathione (GSH), necessary for the synthesis of phytochelatins (Ercal *et al.*, 2001).

The Al<sup>3+</sup> is a phytotoxic element for plants which also cause ROS when entering to this, various research suggests that it may induce oxidative stress, due to the high affinity of this element by the components of the Phospholipid membrane, mainly phosphate and carboxylic groups, which leads to its rigidification (Devi *et al.*, 2003; Ma *et al.*, 1998; Ryan *et al.*, 2001). This phenomenon causes an increase in the amount of reactive oxygen species (ROS), which affect the physiological functioning of plants, and may even induce cell death (Yamamoto *et al.*, 2002; Boscoso *et al.*, 2003; Corrales *et al.*, 2008). The production of ROS various damage to cellular components such as lipid peroxidation, the oxidation of proteins and DNA damage (Munné-Bosch and Alegre, 2002; Barlett and Stadtman, 1997; Ercal *et al.*, 2001).

The Malondialdehyde (MDA) is one of the resulting products of lipid peroxidation and its content accounts for the level of damage that occurs in the lipid membrane in the cell membrane. From here, it is that the MDA is an important physiological indicator of the degree of oxidative stress in the plant (Heath and Parker, 1968).

Several studies have shown that the Cd is an inducer trace element of oxidative stress, which is reflected in an increase in lipid peroxidation. Ge et al., found an increase in the content of MDA in poplar cultivars grown in hydroponic solution exposed to 50 and 100 µM of Cd (Ge *et al.*, 2012). Treatments with Cd exposure, produced lipid peroxidation in *Pisum sativum* (Lozano *et al.*, 1997; Metwally *et al.*, 2005; Pandey and Kumar, 2012; Gaballah *et al.*, 2012), in different cultivars of barley (Guo *et al.*, 2007; Wu *et al.*, 2003), in peanuts (*Arachis hypogaea*), similar results were reported in a study in tomato exposed to different concentrations of Cd (Kumar et al., 2008; Dong *et al.*, 2006).

In the case of plants subjected to treatment with Al, have reported similar behaviors to those of Cd. Studies on barley and quince determined significant increases in lipid peroxidation, in those treatments containing Al (Guo *et al.*, 2007; Wulff *et al.*, 2010). Aluminum generates oxidative damage during variable exposure time, Yamamoto *et al.*, found an increase of lipid peroxidation in pea plants after exposure for 4 hours with Al (Yamamoto *et al.*, 2001), while that Çakmak and Horst (1991), observed in soybeans plants, significant increases in the content of MDA after 48 h of exposure.

To resist oxidative stress, plants can induce a series of reactions catalyzed by detoxifying antioxidant enzymes, where are found, superoxide dismutase (SOD EC 1.15.1.1), Peroxidase (POD EC 1.11.1.7), ascorbate peroxidase (APX EC 1.11.1.11) and

catalase (CAT EC 1.11.1.6) (Garratt *et al.*, 2002), in addition to non-enzymatic low molecular weight antioxidant compounds as ascorbic acid, glutathione, tocopherol, and pigments such as carotenoids (Rama-Devi and Prasad, 1998). These three enzymes maintain the balance between the production of free radicals and eliminate these products, avoiding the effects of free radicals in cells (Gaballah and Rady, 2012). Studies have shown that the activity of these enzymes increases with stress concentration of heavy metals such as Cd and Al (Guo *et al.*, 2004; Gaballah and Rady, 2012; Ribeiro *et al.*, 2012). Therefore the tolerance of some plants, though the presence of heavy metals increases can be attributed in part to the action of the SOD, POD, CAT and other enzymes (Dixit *et al.*, 2001; Lagriffoul *et al.*, 1998; Zhang *et al.*, 2004).

Lactuca sativa exposed for two weeks at 100 μM Cd was increased enzyme activity of SOD and POD, suggesting that both enzymes act in combination to reduce the impact of Cd toxicity, especially in young leaves (Monteiro *et al.*, 2009). However, the amount of enzymes in plants is reduced, so that their role in antioxidant protection and combat the negative effects of oxidative stress induced by heavy metals appears to be limited. Several studies have shown that exposures to high concentrations of Cd decreased antioxidant capacity, mainly of CAT enzyme (Fodor, 2002; Chaoui *et al.*, 1997; Monteiro *et al.*, 2009).

The effects of Al on the activity of antioxidant enzymes have also been investigated by several authors (Meriga *et al.*, 2004; Guo *et al.*, 2007; Boscolo *et al.*, 2003). The induction of SOD activity was observed in *Zea mays* treated with Al<sup>3+</sup> (36 µM) for 48 hours, while this element did not cause activation of the CAT enzyme (Boscolo *et al.*, 2003). *Artemisia annua* L plants treated with soil in the presence of 10 mM of Al<sup>3+</sup>, an increase in the enzymatic activity of CAT, and SOD was observed (Aftab *et al.*, 2010).

Similar results were obtained in rice plants, an increase in antioxidant enzymes CAT, and SOD in the presence of Al<sup>3+</sup> (Meriga *et al.*, 2004, Ribeiro *et al.*, 2012).

## Phenolic compound and antioxidant defenses and protective role of heavy metals

The phenolic compounds are secondary metabolites synthesized by plants as a defense measure against various stresses and act as protective agents against pathogens, being secreted as a defense mechanism to stress conditions, such as infection, UV radiation, among others (Li *et al.*, 2000; Close and McArthur, 2002). These compounds are characterized by containing one or more aromatic rings (C6), together with one or more hydroxyl groups (Balasundram *et al.*, 2006; Ignat *et al.*, 2011).

The phenolic compounds have diverse functions in plants. Research has shown an increase in the amount of phenolic compounds under different environmental conditions and stresses. Research has shown that plants increase the synthesis of phenolic compounds as a strategy to control ROS. From this point, it is possible to consider that phenolic compounds play an antioxidant role or protective role when they grow in environments of stress such as the presence of HM.

Induction of biosynthesis of phenolic compounds has been observed in wheat seedlings in response to toxicity by Ni and corn in response to exposure of Al (Kováčik *et al.*, 2009, 2010). In a *Phaseolus vulgaris* study, it was exposed to Cd<sup>2+</sup> accumulating phenolic compounds (Milone *et al.*, 2003). Irtelli and Navari-Izzo, (2006) found a decrease of phenolic compounds such as chlorogenic acid in *Brassica juncea*, exposed to Cd.

Studies reported on the effect of Cu in the production of phenolic compounds in *Matricaria chamomilla*, showed changes in the contents of 11 phenolic compounds such as

chlorogenic acid, caffeic acid, vallinic acid, among others, which varied depending on the metal concentration (Kováčik *et al.*, 2008).

It has been investigated that the production of phenolic compounds, help the plant to have a better tolerance to metals such as Al<sup>3+</sup>, which has allowed us to develop the hypothesis that phenolic compounds have a role in detoxification when acting by complexing in a stable way Al<sup>3+</sup> in plants, in addition to the antioxidant capacity that phenolic compounds have, helping Al tolerance in plants such as corn (Tolrá *et al.*, 2005, 2009; Rout *et al.*, 2001).

### Concluding remarks and future trends

The trace element stress is one of the main problems affecting the agricultural productivity of soils and plants. Tolerance to these elements varies between species, which has led to different plants grow well on soils enriched by these elements. Scientific research has helped to elucidate the most important negative effects that these elements produce to cultures, showing that many of the first visible symptoms of phytotoxicity are similar between metals or in many cases similar to symptoms of nutrient deficiency. The chemical speciation of these elements in the soil solution is related to pH, organic matter, clays present, which may increase their availability to plants, increasing the toxicity of these. Elements such as Cd and Al are highly toxic due to their mobility in soil, in addition to being in the case of Al the biggest problem in acid soils system.

Different investigations have shown that the presence of trace elements in general induces ROS which negatively affects plants. The mechanisms to combat these species are highly known and studied to date, which include the enzymatic and non-enzymatic

mechanisms. It is in this sense that the study of non-enzymatic mechanisms such as the production of phenolic compounds suggests to be a promising approach for the study of the detoxification of plant with tolerant and sensitive genotypes.

Although the phenolic compounds are synthesized in normal conditions by the plant, an alteration in the conditions of growth and development, such as the presence of heavy metals, causes changes in producing these compounds. There is not a clear signal of what are the involved routes that correlate the synthesis of phenolic compounds and the presence of heavy metals.

Despite numerous studies on heavy metals or trace elements-plant interaction and synthesis of phenolic compounds, there are still steps to understand the phenomenon, due to the absence of studies on a complementary or synergistic effect of various parameters or agents that participate in periods of plant growth and development.

#### Referencias.

- Aftab, T., Masroor, M., Khan, A., Idrees, M., Naeem, M. Moinuddin, M., 2010.
   Effects of aluminium exposures on growth, photosynthetic efficiency, lipid peroxidation, antioxidant enzymes and artemisinin content of *Artemisia annua* L.
   Journal of Phytology 2, 23–37.
- 2. Arora, A., Sairam, R.K., Srivastava, G.C., 2002. Oxidative stress and antioxidative system in plants. Current Science 82, 1227-1238.
- 3. Apel, K., Hirt, H., 2004. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. Annul Review of Plant Biology 55, 373-399.

- 4. Astolfi, S., Zuchi, S., Passera, C., 2005. Effect of cadmium on H<sup>+</sup> ATPase activity of plasma membrane vesicles isolated from roots of different S-supplied maize (*Zea mays* L.) plants. Plant Science 169, 361-368.
- Baker, D.E., Senft, J.P., 1995. Copper. In 'Heavy Metals inSoils'. 2nd. edn. (ed. Alloway, B.J.). pp. 179-205. Blackie Academic and Professional, London.
- 6. Baker, J.F., Burrows, N.L., Keohane, A.E., Filippis, L.F., 1995. Chemical root pruning of kangaroo paw (Anigozanthos flavidus) by selected heavy metal carbonates. Scientia Horticulturae 62, 245-253.
- 7. Baker, A.J.M., 1981. Accumulators and exhuders-strategies in response of plants to heavy metals. Journal ofPlant Nutrition 3, 643-654.
- 8. Balasundram, N., Sundram, K., Samman, S., 2006. Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. Food Chemistry 99, 191-203.
- 9. Balestrasse, K.B., Gallego, S.M., Tomaro, M.L., 2004. Cadmium-induced senescence in nodules of soybean (*Glycine max* L.) plants. Plant and Soil 262, 373-381.
- 10. Barančíková, G., Madaras, M., Rybár, O., 2004. Crop contamination by selected trace elements. Journal of Soils and Sediments 4, 37-42.
- 11. Barlett, B., Stadtman, E., 1997. Protein oxidation in aging, disease, and oxidative stress. The Journal of Biological Chemistry 272, 20313-20316.

- 12. Bar-Tal, A., Baryosef, B., Chen, Y., 1988. Effects of fulvic-acid and pH on zinc sorption on montmorillonite. Soil Science 146, 367-373.
- Basta, N.T., Tabatabai, M.A., 1992. Effect of cropping systems on adsorption of metals by soils: II. Effect of pH. Soil Science 153, 195-204.
- Benavides, M.P., Gallego, S.M., Tomaro, M., 2005. Cadmium toxicity in plants.
   Brazilian Journal of Plant Physiology 17, 21-34.
- 15. Blamey, F.P.C., Wheeler, D.M., Edmeades, D.C., Christie, R.A., 1990.
  Independence of differential aluminum tolerance in Lotus on changes in rhizosphere
  pH or excretion of organic ligands. Journal of Plant Nutrition 13, 713-728.
- 16. Bonomelli, C., Bonilla, C., Valenzuela, A., Saavedra, N.,2002. Presencia de Cadmio en fertilizantes fosfatados de diferente procedencia comercializados en Chile, segunda temporada. Simiente 72, 9-16.
- 17. Boscolo, P., Menossi, M. and Jorgea, R., 2003. Aluminum-induced oxidative stress in maize. Phytochemistry 62, 181-189.
- 18. Boularbah, A., Schwartz, C., Bitton, G., Morel, J., 2006. Heavy metal contamination from mining sites in South Morocco: 1. Use of a biotest to assess metal toxicity of tailings and soils. Chemosphere 63, 802-810.
- 19. Cakmak, I., Horst, W.J., 1991. Effect of aluminum on lipid peroxidation, superoxide dismutase catalase and peroxidase activities in roots tips of soybean (*Glicine max* L). Plant Physiology 83, 463-468.

- 20. Cavallaro, N., McBride, M.B., 1984. Zinc and copper sorption and fixation by and acid soil clay: Effect of selective dissolutions. Soil Science Society of America Journal 48, 1050-1054.
- 21. Chang, A.C., Page, A.L., 2000. Trace element slowly accumulating, depleting in soils. California Agriculture 54, 49-55.
- 22. Chaoui, A., Mazhoudi, S., Ghorbal, M.H., Elferjani, E., 1997. Cadmium and zinc induction of lipid peroxidation and effects on antioxidant enzyme activities in vean (*Phaseolus vulgaris* L.). Plant Science 127, 139–147.
- 23. Cherif, J., Mediouni, Ch., Ammar W. B., Jemal, F., 2011. Interactions of zinc and cadmium toxicity in their eects on growth and inantioxidative systems in tomato plants (*Solanum lycopersicum*). Journal of Environmental Sciences 23, 837–844.
- 24. Clemens, S., Palmgren, M.G., Krämer, U., 2002. Along way ahead: understanding and engineering plant metal accumulation. Plant Science 7, 309-315.
- 25. Close, D. C., McArthur, C., 2002. Rethinking the role of many plant phenolics—Protection from photodamage not herbivores? Oikos 99, 166-172.
- 26. Cordero, B., Lodeiro, P., Herrero, R., Esteban Sastre de Vicente, M. 2004. Biosorption of cadmium by Fucus spiralis. Environmental Chemistry 1, 180-187.
- 27. Corrales, I., Poschenrieder, C., Barcelo, J., 2008. Boron induced amelioration of aluminum toxicity in a monocot and a dicot species. Journal ofPlant Physiology 165, 504-513.

- 28. Cuypers, A., Plusquin, M., Remans, T., Jozefczak, M., Keunen, E., Gielen, H., Opdenakker, K., Nair, A.R., Munters, E., Artois, T.J., Nawrot, T., Vangronsveld, J., Smeets, K., 2010. Cadmium stress: an oxidative challenge. Biometals 23, 927-940.
- 29. Davis, J.A., Leckie, J.O., 1978. Surface ionization and complexation at the oxide/water interface. II. Surface properties of amorphous iron oxyhydroxide and adsorption of metal ions. Journal of Colloid and Interface Science 67, 90-107.
- 30. Davis-Carter, J.G., Shuman, L.M., 1993. Influence of texture and pH of kaolinitic soils on zinc fractions and zinc uptake by peanuts. Soil Science 155, 376-384.
- 31. Devi, R. S., Yamamoto, Y., Matsumoto, H., 2003. An intracellular mechanism of aluminum tolerance associated with high antioxidant status in cultured tobacco cells. Journal of Inorganic Biochemistry 97, 59-68.
- 32. Dietz, K.J., Baier, M., Kramer, U., 1999. Free radical and active oxigen species as mediators of heavy metal toxicity in plants. In: Prasad M.N.V. Hagemeyer J. (eds.), Heavy Metal Stress in Plants. Springer, Berlin, pp 73-97.
- 33. Dixit, V., Pandey, V., Shyam, R., 2001. Differential oxidative responses to cadmium in roots and leaves of pea (*Pisum sativum* L). Journal of Experimental Botany 52, 1101-1109.
- 34. Dong, J., Wu, F., Zhang, G., 2006. Influence of cadmium on antioxidant capacity and four microelement concentrations in tomato seedlings (*Lycopersicon esculentum*). Chemosphere 64, 1659-1666.

- 35. Dudley, L. M., McLean, J. E., Furst, T. H., Jurinak, J. J., 1991. Sorption of Cd and Cu from an acid mine waste extract by two calcareous soils: column studies. Soil Science 151,121-135.
- 36. Dudley, L. M., McNeal, B. L., Baham, J. E., Coray C. S., ChengH. H., 1987. Characterization of soluble organic compounds and complexation of copper, nickel, and zinc in extracts of sludge-amended soils. Journal of Environmental Quality 16, 341-348.
- 37. Ercal, N., Gurer-Orhan, H., Aykin-Burns, N., 2001. Toxic metal and oxidative stress part I: mechanisms involved in metal induced oxidative damage. Current Topic in Medicinal Chemistry 1, 529-539.
- 38. Escudey, M., Förster, J.E., Becerra, J.P., Quinteros, M., Torres, J., Arancibia, N., Galindo, G., Chang, A.C., 2007. Disposal of domestic sludge and sludge ash on volcanic soils. Journal of Hazardous Materials 139, 550-555.
- 39. Fageria, N.K., Baligar, V.C., Jones, C.A., 1991. Growth and Mineral Nutrition of Field Crops. Marcel Dekker, New York.
- 40. Fan, T.W.M., Lane, A.N., Pedler, J., Crowley, D., Higashi, R.M., 1997. Comprehensive analysis of organic ligands in whole root exudates using nuclear magnetic resonance and gas chromatography mass spectrometry. Analytical Biochemistry 251, 57-68.
- 41. Fergus, I.F., 1954. Manganese toxicity in an acid soil. Queensland Journal of Agricultural Science 11, 15-21.

- 42. Fodor, F., 2002. Physiological responses of vascular plants to heavy metals. In: Prasad MNV, Strzalka K (eds) Physiology and biochemistry of metal toxicity and tolerance in plants. Kluwer Academic Publisher, Dortrech, pp 149–177
- 43. Fodor, A., Szabó-Nagy, A., Erdei, L., 1995. The effects of cadmium on the fluidity and H<sup>+</sup>-ATPase activity of plasma membrane from sunflower and wheat roots.

  Journal of Plant Physiology 14, 787-792.
- 44. Frausto da Silva, J.J.R., Williams, R.J.P. The Biological Chemistry of the Elements: The Inorganic Chemistry of Life. 2nd edn. Oxford: Oxford University Press; 2001.
- 45. Fu, J., Huang, B., 2001. Involvement of antioxidants and lipid peroxidation in the adaptation of two cool-season grasses to localized drought stress. Environental and. Experimental Botany 45, 105–114.
- 46. Gaballah, M., Rady, M.M., 2012. Salicylic Acid Mitigated Cadmium Toxicity by Attenuating the Oxidative Stress in Pea (*Pisum sativum* L.) Plants. International Journal of Biological, Ecological and Environmental Sciences 4, 159-165.
- 47. Garratt, L.C., Janagoudar, B.S., Lowe, K.C., Anthony, P., Power, J.B., Davey, M.R., 2002. Salinity tolerance and antioxidant status in cotton cultures. Free Radical Biology and Medicine 33, 502-511.
- 48. Garrido, E., Matus, F., 2012. Are organo-mineral complexes and allophane content determinant factors for the carbon level in Chilean volcanic soils? Catena 92, 106-112.

- 49. Ge, W., Jiao, Y.Q., Sun, B.L., Qin, R., Jiang, W.S., Liu, D.H., 2012. Cadmium-mediated oxidative stress and ultrastructural changes in root cells of poplar cultivars. South African Journal of Botany 83, 98–108.
- 50. Ghanem, S.A. and Mikkelsen, D.S., 1988. Sorption of zinc on iron hydrous oxide. Soil Science 146, 15-21.
- 51. Giannakoula, A., Moustakas, M., Mylona, P., Papadakis, I., Yupsanis, T., 2008. Aluminum tolerance in maize is correlated with increased levels of mineral nutrients, carbohydrates and proline, and decreased levels of lipid peroxidation and Al accumulation. Journal of Plant Physiology 165, 385-396.
- 52. Gunsé, B., Garzón, T., Barceló, J., 2003. Study of aluminum toxicity by means of vital staining profiles in four cultivars of *Phaseolus vulgaris* L. Journal of Plant Physiology 160, 1447-1450.
- 53. Godbold, D.L., Huttermann, A., 1985. Effect of zinc, cadmium and mercury on root elongation of Picea abies (Karst.) seedlings, and the significance of these metals to forest die-back. Environmental Pollution (series A) 38, 375-381.
- 54. Gouia, H., Ghorbal, M.H., Meyer, C., 2000. Effects of cadmium on activity of nitrate reductase and on other enzymes of nitrate assimilation pathway in bean. Plant Physiology and Biochemistry 38,629-638.
- 55. Guo, T. R., Zhang, G.P.and Zhang, Y.H., 2007. Physiological change in barley plants under combined toxicity of aluminum, copper and cadmium. Colloids and Surface B: Biointerfaces 57, 182-188.

- 56. Guo, T., Zhang, G., Zhou, M., Wu, F., Chen, J., 2004. Effects of aluminum and cadmium toxicity on growth and antioxidant enzyme activities of two barley genotypes with different Al resistance. Plant and Soil 258, 241-248.
- 57. Heath, R., Parker, L., 1968. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. Archives Biochemistry and Biophysics 125, 189-198.
- 58. Huang, J.W., Pellet, D.M., Papernik, L.A. and Kochian, L.V., 1996. Aluminum interactions with voltage dependent calcium transport in plasma membrane vesicles isolated from roots of aluminum-sensitive and -resistant wheat cultivars. Plant Physiology 110, 561-569.
- 59. Ignat, I., Volf, I., Popa, V. I., 2011. A critical review of methods for characterisation of polyphenolic compounds in fruits and vegetables. Food Chemistry 126, 1821-1835.
- 60. Irtelli, B., Navari-Izzo, F., 2006. Influence of sodium nitrilotriacetate (NTA) and citric acid on phenolic and organic acids in *Brassica juncea* L grown in excess of cadmium. Chemosphere 65, 1348-1354.
- 61. Jeffery, J., Uren, N.C., 1983. Copper and zinc species in the soil solution and the effects of soil pH. Australian Journal of Soil Research 21, 479-488.
- 62. Kalbasi, M., Racz, G.J. and Loewen Rudgers, L.A., 1978. Mechanism of zinc adsorption by iron and aluminum oxides. Soil Science 125, 146-150.

- 63. Keltjens, W.G. and Van Beusichem, M.L., 1998. Phytochelatins as biomarkers for heavy metal toxicity in maize: Single metal effects of copper and cadmium. Journal of Plant Nutrition 21, 634-648.
- 64. Keunen, E., remans, T., Bohler, S., Vangronsveld, J., Cuypers, A., 2011. Metal-Induced oxidative stress and plant mitocondria. Molecular Sciences 12, 6894-6918.
- 65. Kovácik, J., Grúz, J., Backor, M., Tomko, J., Strnad, M., Repcák, M., 2008.

  Phenolic compounds composition and physiological attributes of Matricaria chamomillagrown in copper excess. Environmental and Experimental Botany 62, 145-152.
- 66. Kovácik, J., Klejdus, B., Backor, M., 2009. Phenolic metabolism of *matricaria chamomilla* plants exposed to nickel. Journal of plant physiology 166, 1460-1464.
- 67. Kovácik, J., Klejdus, B., Hedbavny, J., 2010. Effect of aluminium uptake on physiology, phenols and amino acids in *Matricaria chamomilla* plants. Journal of Hazardous Materials 178, 949–955.
- 68. Kumar, S., Mehta, U. J., Hazra, S., 2008. Accumulation of cadmium in growing peanut (*Arachis hypogaea* L.) seedlings-Its effect on lipid peroxidation and on the antioxidative enzymes catalase and guaiacol peroxidase. Journal of Plant Nutritionand Soil Science 171, 440–447.
- 69. Lagriffoul, A., Mocquot, B., Mench, M., 1998. Cadmium toxicity effects on growth, mineral and chlorophyll contents, and activities of stress related enzymes in young maize plants (*Zea mays* L.). Plant and Soil 200, 241-250.

- 70. Larcher W., 2004. Ecophysiology and Stress Physiology of Functional Groups Physiological. Plant Ecology 77, 365-366.
- 71. Laurie, S.H. and Manthey, J.A., 1994. The chemistry and role of metal ion chelation in plant uptake processes. In Biochemistry of Metal Micronutrients in the Rhizosphere. (ed. Manthey, J.A., Crowley, D.E. and Luster, D.G.). pp. 165-182. Lewis Publishers, Boca Raton.
- 72. Lexmond, T.M., 1980. The effect of soil pH on copper toxicity to forage maize grown under field conditions. Netherlands Journal of Agricultural Science 28, 164-183.
- 73. Li, C., Berninger, F., Koskela, J. and Sonninen, E., 2000. Drought responses of Eucalyptus microtheca provenances depend on seasonality of rainfall in their place of origin. Australian Journal of Plant Physiology 27, 231-238.
- 74. Li, M., Zhang, L.J. Tao L., Li., W., 2008. Ecophysiological responses of Jussiaea rapens to cadmium exposure. Aquatic Botany 88, 347-352.
- 75. Lidon, F.C. and Henriques, F.S. 1992. Copper toxicity in rice: Diagnostic criteria and effect on tissue manganese andiron. Soil Science 154, 130-135.
- 76. Liu, X.L., Zhang, S.Z., Shan, X.Q., Christie, P., 2007. Combined toxicity of cadmium and arsenate to wheat seedlings and plant uptake and antioxidative enzyme responses to cadmium and arsenate co-contamination. Ecotoxicology and Environmental Safety 68, 305–313.

- 77. Lozano-Rodriguez, E., Hernàndez, L.E., Bonay, P., Carpena-Ruiz, R.O., 1997. Distributions of cadmium in shoot and root tissues. Journal of Experimental Botany 48, 123-128.
- 78. Lukaszewski, K. M., Blevins, D. G., 1996. Root growth inhibition in boron-deficient or aluminum-stressed squash may be a result of impaired ascorbate metabolism. Plant Physiology 112, 1135-1140.
- 79. Ma, J.F., Hiradate, S., Matsumoto, H., 1998. High aluminum resistance to buckwheat. II. Oxalic acid detoxifies aluminum internally. Plant Physiology 117, 753-759.
- 80. Maksymiec, W., Wojcik, M., Krupa, Z., 2007. Variation in oxidative stress and photochemical activity in Arabidopsis thaliana leaves subjected to cadmium and excess copper in the presence or absence of jasmonate and ascorbate. Chemosphere 66, 421-427.
- 81. Meriga, B., Reddy, B. K., Rao, K. R., Reddy, L. A., Kishor P. B. K., 2004. Aluminium-induced production of oxygen radicals, lipid peroxidation and DNA damage in seedlings of rice (*Oryza sativa*). Journal of Plant Physiology 161, 63-68.
- 82. Milone, M.T., Sgherri, C., Clijsters, H., Navari-Izzo, F., 2003. Antioxidative responses of wheat treated with realistic concentration of cadmium. Environmental and Experimental Botany 50, 265-276.

- 83. Miyasaka, S.C., Buta, J.G., Howell, R.K., Foy, C.D., 1991. Mechanism of aluminumtolerance in snapbeans: root exudation of citric acid. Plant Physiology 196, 737-743.
- 84. Marschner, H., 1995. 'Mineral Nutrition of Higher Plants.' 2<sup>nd</sup> edn. Academic Press, London.
- 85. McBride, M., Sauve, S., Hendershot, W., 1997. Solubility control of Cu, Zn, Cd and Pb in contaminated soils. European Journal of Soil Science 48, 337-346.
- 86. McBride, M.B., 1982. Electron spin resonance investigation of Mn<sup>2+</sup> complexation in natural and synthetic organics. Soil Science Society of America Journal 46, 1137-1143.
- 87. McGrath, S.P., Sanders, J.R., Shalaby, M.H., 1988. The effects of soil organic matter levels on soil solution concentrations and extractabilities of manganese zinc and copper. Geoderma 42, 177-188.
- 88. McGrath, S.P., Zhao, F.J., 2002. Phytoextraction of metals and metalloids from contaminated soils. Current Opinion in Biotechnology 14, 277-282.
- 89. Metwally, A., Safronova, V. I., Belimov, A., Dietz, K.J., 2005. Genotypic variation of the response to cadmium toxicity in *Pisum sativum* L. Journal of Experimental Botany 56, 167-178.
- 90. Merwin, M.L., Martin, J.A., Westfall, R.D., 1995. Provenance and progeny variation in growth and frost tolerance of Casuarina cunninghamiana in California, USA. Forest Ecology and Management 79, 161-171.

- 91. Meyer A., Tempé J. and P. Costantino., 2000. Hairy root; a molecular overview. Functional analysis of Agrobacterium rhizogenes T-DNA genes. In: G. Stacey and N.T. Keen, Editors, Plant Microbe Interactions, APS Press, St. Paul, pp. 93–139.
- 92. Miller, W.P., Martens, D.C. and Zelazny, L.W., 1987. Short-term transformations of copper in copper-amended soils. Journal of Environmental Quality 16, 176-180.
- 93. Milone, M.T., Sgherri, C., Clijsters, H., Navari-Izzo, F., 2003. Antioxidative responses of wheat treated with realistic concentration of cadmium. Environmetal and Experimental Botany 50, 265-276.
- 94. Mithofer, A., Schulze, B., Boland, W., 2004. Biotic and heavy metal stress response in plants: evidence for common signals. FEBS letter, 566, 1-5.
- 95. Molina, M., Manquian-Cerda, K. and Escudey, M., 2010. Sorption and selectivity sequences of Cd, Cu, Ni, Pb, and Zn in single- and multi-component systems in a cultivated Chilean mollisol. Soils and Sediments Contamination 19, 405-418.
- 96. Monni, S., Salemma, M., Millar, N., 2000. The tolerance of Empetrum nigrum to copper and nickel. Environmental Pollution 109, 221-229.
- 97. Monteiro, M.S., Santos, C., Soares, A.M.V.M., Mann, R.M., 2009. Assessment of biomarkers of cadmium stress in lettuce. Ecotoxicology and Environmental Safety 72, 811–818.
- 98. Msaky, J.J. and Calvet, R., 1990. Adsorption behaviour of copper and zinc in soils: influence of pH on adsorption characteristics. Soil Science 150, 513-522.

- 99. Munné-Bosch, S., Alegre, L., 2002. Plant aging increases oxidative stress in chloroplasts. Planta 214, 608-615.
- 100. Norvell, W.A., 1991. Reactions of metal chelates in soils and nutrient solutions. In 'Micronutrients in Agriculture'. 2<sup>nd</sup>.edn. (ed. Mortvedt, J.J., Cox, F.R., Shuman, L.M. and Welch, R.M.). pp.187-227. Soil Science Society of America, Madison
- 101. Olivares, E., Pena, E., Marcano, E., Mostacero, J., Aguiar, G., Benitez, M., Rengifo, E., 2009. Aluminum accumulation and its relationship with mineral plant nutrients in 12 pteridophytes from Venezuela. Environmental and Experimental Botany 65, 132-141.
- 102. Olomu, M.O., Racz, G.J. and Cho, C.M., 1973. Effect of flooding on the Eh, pH, and concentrations of Fe and Mn in several Manitoba soils. Soil Science Society of America Proceedings 37, 220-224.
- 103. Otero, N., Vitoria, L., Soler, A., Canals, A., 2005. Fertiliser Characterisation: Major, Trace and Rare Earth Elements. Applied Geochemistry 20, 1473-1488.
- 104. Pampura, T.B., Pinskiy, D.L., Ostroumov, V.G., Gershevich, V.D., Bashkin, V.N., 1993. Experimental study of the buffer capacity of a Chernozem contaminated with copper and zinc. Eurasian Soil Science 25, 27-38.
- 105. Pandey, N., Kumar, G., 2012. Studies on antioxidative enzymes induced by cadmium in pea plants (*Pisum sativum*). Journal of Environmental Botany 33, 201-206.

- 106. Parker, D.R., Pedlar, J.F., Ahnstrom, Z.A.S., Resketo, M., 2001. Reevaluating the free-ion activity model of trace element metal toxicity towards higher plants: experimental evidence with copper and zinc. Environmental toxicology and chemistry 20, 899-906.
- 107. Payne, K., Pickering, W.F., 1975.Influence of clay-solute interactions on aqueous copper ion levels. Water, Air and Soil Pollution 5, 63-69.
- 108. Peijnenburg, W., Baerselman, R., de Groot, A., Jager, T., Leenders, D., Posthuma, L., Van Veen, R., 2000. Quantification of metal bioavailability for lettuce (*Lactuca sativa* L.) infield soils. Archives of Environmental Contamination and Toxicology 39, 420-430.
- 109. Pinochet, D.T., Aguirre, J.A., Quiroz, E.R., 2002. Estudio de la lixiviación de cadmio, mercurio y plomo en suelos derivados de cenizas volcánicas. Agro Sur 30, 51-58.
- 110. Poschenrieder, C., Tolrà, R., Barceló, J., 2006. Can metals defend plants against biotic stress? Trends in Plant Science 11,288-295.
- 111. Qiao, L., Ho, G., 1996. The effect of clay amendment on speciation of heavy metals in sewage sludge. Water Science and Technology 34, 413-420.
- 112. Rama-Devi, S., Prasad, M.N.V., 1998. Copper toxicity in *Ceratophyllum demeresum* L. (Coontail), a free floating macrophyte: Response of antioxidant enzymes and antioxidants. Plant Science 138, 157-165.

- 113. Redjala, T., Zelko, I., Sterckeman, T., Legué, V., Lux, A., 2011. Relationship between root structure and root cadmium uptake in maize. Environmental and Experimental Botany 71, 241-248.
- 114. Ribeiro, C., Cambraia, J., Henrique, P., Peixoto, P., Meira, E., 2012. Antioxidant system response induced by aluminum in two rice cultivars. Brazilian society of plant physiology 24, 107-116.
- 115. Rout, G.R., Samantaray, S., Das, P., 2001. Aluminium toxicity in plants: a review. Agronomie 21, 3-21.
- 116. Ryan, P.R., Kochian, L.V., 1993. The interaction between aluminum toxicity and calcium uptake at the root apex in near-isogenic lines of wheat (*Triticum aestivum* L.) differing in aluminum tolerance. Plant Physiology 102, 975-982.
- 117. Ryan, P.R, Delhaize, E., Jones, D.L., 2001. Function and mechanism of organic acid excretion from plant roots. Annual Review of Plant Biology. Plant Molecular Biology 52, 527–560.
- 118. Sakihama, Y., Cohen, M.F., Grace, S.F., Yamasaki H., 2002. Plant phenolic antioxidant and prooxidant activities: phenolics-induced oxidative damage mediated by metals in plants. Toxicology 177, 67–80.
- 119. Salt, D.E., Smith, R.D., Raskin, I., 1998. Phytoremediation. Annual Review of Plant Physiology and Molecular Biology 49, 643–668.

- 120. Sandalio, L.M., Dalurzo, H.C., Gomez, L.A., Romero, M.C., Puertas, M.C., del Río, M., 2001. Cadmium-induced changes in the growth and oxidative metabolism of pea plants. Journal of Experimental Botany 52, 2115-2126.
- 121. Sanitá di Toppi, L., Gabbrielli, R., 1999. Response to cadmium in higher plants. Environmental and Experimental Botany 41, 105-130.
- 122. Sauve, S., McBride, M.B., Norvell, W.A., Hendershot, W.H., 1997. Copper solubility and speciation of in situ contaminated soils: effects of copper level, pH and organic matter. Water, Air and Soil Pollution 100, 133-149.
- 123. Sauve, S., Cook, N., Hendershot, W.H. and McBride, M.B., 1996. Linking plant tissue concentration sand soil copper pools in urban contaminated soils. Environmental Pollution 94, 153-157.
- 124. Schutzendubel, A., Polle A., 2002. Plant responses to abiotic stresses: Heavy metal induced oxidative stress and protection by mycorrhization. Journal of Experimental Botany 53, 1351-1365.
- 125. Scora, R. W., Chang, A. C., 1997. Essential oil quality and heavy metal concentrations of peppermint grown on a municipal sludge-amended soil. Journal of Environmental Quality 26, 975-979.
- 126. Silva, I.R., Smyth, T., Barros, H., Novais, R., 2002. Physiological aspects of aluminum toxicity and tolerance in plants. Tópicos em Ciencia do Solo 2, 277-335.
- 127. Smeets, K., Cuypers, A., Lambrechts, A., Semane, B., Hoet, P., Laerve, A.V., Vangronsveld, J., 2005. Induction of oxidative stress and antioxidative mechanisms

- in Phaseolus Vulgaris after Cd application. Plant Physiology and Biochemistry 43, 437-444.
- 128. Stevenson, F.J., 1991. Organic matter-micronutrient reactions in soil. In 'Micronutrients in Agriculture'. 2<sup>nd</sup>. edn. (ed. Mortvedt, J.J., Cox, F.R., Shuman, L.M. and Welch, R.M.). pp. 145-186. Soil Science Society of America, Madison.
- 129. Tam, P.C.F., 1995. Heavy metal tolerance by ectomycorrhizal fungi and metal amelioration by Pisolithus tinctorius. Mycorrhiza 5, 181-187.
- 130. Taylor, G.J., Blamey, F.P.C., Edwards, D.G., 1998. Antagonistic and synergistic interactions between aluminum and manganese on growth of Vigna unguiculata at low ionic strength. Physiologia Plantarum 104, 183-194.
- 131. Tolrà, R., Poschenrieder, C., lupi, P., Barceló, J., 2005. Aluminium-induced changes in the profiles of both organic acids and phenolic substances underlie al tolerance in *Rumex acetosa* L. Environmental and Experimental Botany 54, 231-238.
- 132. Tolrà, R., Barceló, J., Poschenrieder, C., 2009. Constitutive and aluminium induced patterns of phenolic compounds in two maize varieties differing in aluminium tolerance. Journal of Inorganic Biochemistry 103, 1486-1490.
- 133. Turner, A.P., 1994. The responses of plants to heavy metals. In 'Toxic Metals in Soil-Plant Systems'. (ed. Ross, S.M.). pp. 153-187.John Wiley and Sons, Chichester.

- 134. Valencia, R., Rubén, A., Ligarreto, M., Gustavo, A., 2012. Differential response of plants to aluminum. A review. Agronomía colombiana 30, 71-77.
- 135. Vega, S., Calisay, M., Hue, N.V., 1992. Manganese toxicity in cowpea as affected by soil pH and sewage sludge amendments. Journal of Plant Nutrition 15, 219-231.
- 136. Wang, A.S., Angle, J.S., Chaney, R.L., Delorme, T.A., Reeves, R.D., 2006. Soil pH effects on uptake of Cd and Zn by Thlaspi caerulescens. Plant and Soil 281, 325-337.
- 137. Welch, R.M., 1995. Micronutrient nutrition of plants. Critical Reviews in Plant Science14, 49-82.
- 138. Whitehead, D.C., 2000. Nutrient Elements in Grasslands: Soil-Plant-Animal Relationships. CABI Publishing, Wallingford.
- 139. Wu, F.B., Zhang G.P., Dominy, P., 2003. Four barley genotypes respond differentily to cadmium: Lipid peroxidation and activities of antioxidant capacity. Environmental and Experimental Botany 50, 67-78.
- 140. Wulff, M., Cellini, A., Masia, A., Marino, G., 2010. Aluminium-induced effects on growth, morphogenesis and oxidative stress reactions in *in vitro* cultures of quince. Scientia Horticulturae 125, 151-158.
- 141. Yamamoto, Y., Kobayashi, Y., Devi, S.R., Rikiishi, S., Matsumoto, H., 2002. Aluminum toxicity is associated with mitochondrial dysfunction and the production of reactive oxygen species in plant cells. Plant Physiology 128, 63 72.

142. Yamamoto, Y., Kobayashi, Matsumoto, H., 2001. Lipid Peroxidation Is an Early Symptom Triggered by Aluminum, But Not the Primary Cause of Elongation Inhibition in Pea Roots. Plant Physiology 125, 199-208.

# **Chapter 2**

# Effect of aluminum on antioxidant activity and phenolic compounds content in *in vitro* cultured blueberries

<sup>†</sup>The chapter was published in *Boletín Latinoamericano y del Caribe de Plantas Medicinales y*Aromáticas, 2013, 12(6): 603-611 (ISI).

#### **Abstract**

Blueberry is a popular natural food product consumed worldwide. Acid soils are found throughout the world. A significant problem of acid soils is the active aluminum content, which may result toxic to plant. The present study was undertaken to assess the toxicities of Al for Blueberry (*Vaccinium corymbosum* L.) cultivated *in vitro* and treated with 100 and 200 µM Al. The effects of Al concentration on malondialdehyde (MDA) content, antioxidant activity and phenolic compounds of blueberry after 7, 14 and 21 days of treatment were established. The analysis of the MDA accumulated in the tissues of the blueberry plantlets indicates that Al concentration increases the damage caused by lipidic peroxidation, for both treatments, after 14 days. The highest antioxidant activity in the extracts was observed at 200 µM Al after 14 days of treatment, being chlorogenic and ellagic acids the most significant metabolites involved in the antioxidant properties. Then, the content of Al in soil could be modulate the content of bioactive compounds in blueberry plants.

**Keywords:** Blueberry, aluminum, antioxidant capacity, phenolic compounds.

# Resumen

El Arándano es un popular alimento natural consumido en todo el mundo. Los suelos ácidos se encuentran en todo el mundo. Un problema significativo de suelos ácidos es el contenido de aluminio activo, que puede resultar tóxico para la planta. Este estudio se realizó para evaluar la toxicidad del aluminio en plantas de arandano, cultivadas in vitro y tratadas con 100 y 200 mM de Al.Se establecieron los del aluminio en el contenido de malodialdehido (MDA), capacidad antioxidante y contenido de compuestos fenolicos en plantulas de arandano luego de 7, 14 y 21 dias de tratamiento. El análisis del MDA

acumulado en los tejidos de las plantulas de arándanos indica que la concentración de Al aumenta el daño causado medido como peroxidación de lípidos, para ambos tratamientos, después de 14 días. La actividad antioxidante más alta de los extractos se obseva a 200 mM de Al después de 14 días de tratamiento, siendo los ácidos clorogénico y elágico los metabolitos más importantes que participan en las propiedades antioxidantes. Entonces, el contenido de Al en el suelo podría modular el contenido de compuestos bioactivos en plantas de arándanos, alterando sus propiedades medicinales.

**Palabras Clave**: Arándano, aluminio, capacidad antioxidante, compuestos fenólicos.

#### **INTRODUCTION**

Aluminum (Al) phytotoxicity is one of the major agronomic problems in acid soils (Guo *et al.*, 2007; Tolrá *et al.*, 2009). Its availability and activity in the soil solution is heightened to pH less than 5.5, where concentrations in the order μM quickly may inhibit the elongation of roots and subsequently the capture of water and nutrients (Alvarez *et al.*, 2005).

A mechanism of tolerance to this element in plants is the exudation of organic acids from the roots, which trap the free Al present in soil solution (Kochian *et al.*, 2005; Pineros *et al.*, 2008; Poschenrieder *et al.*, 2008; Liu *et al.*, 2009; Giannakoula *et al.*, 2010). However, despite this defense mechanism, the Al plant uptake induces oxidative stress due to their high affinity with phosphate and carboxylic groups present in the plasma membrane. (Ma *et al.*, 2001; Ryan *et al.*, 2001; Devi-Rama *et al.*, 2003). This phenomenon increases the amount of reactive oxygen species (ROS), which affect various physiological parameters of the plant and may even induce cell death (Yamamoto *et al.*, 2002; Boscolo *et al.*, 2003; Corrales *et al.*, 2008).

Plants show efficient systems for scavenging active oxygen species that protect them from destructive oxidative reactions (Munné-Bosch *et al.*, 2001). As part of this system, antioxidative enzymes are key elements in the defense mechanisms. Garratt *et al.*, (2002) has listed some of these enzymes as catalase (CAT EC 1.11.1.6), glutathione reductase (GR EC 1.6.4.2), superoxide dismutase (SOD EC 1.15.1.1) and glutatione-S-transferase (GST EC 2.5.1.18). Superoxide dismutase, for example, metabolizes oxygen (O<sub>2</sub>) radicals to hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), thus protecting cells from damage. Catalase, ascorbate peroxidase, and a variety of peroxidases catalyze the subsequent breakdown of H<sub>2</sub>O<sub>2</sub>to

water and oxygen (Garratt *et al.*, 2002). Plants with high levels of antioxidants have been reported to have greater resistance to this oxidative damage (Koca *et al.*, 2007).

It is proposed that the phenolic compounds, increase plant tolerance to Al, due to its ability to detoxify through the formation of stable complexes of Al and also for their antioxidant capacity (Tolrá *et al.*, 2009).

A plant species highly valued for its high content of phenolic compounds, is the blueberry (Kähkönen *et al.*, 2001; Dastmalchi *et al.*, 2010). Blueberries are native to North America and have a rich folklore history of medicinal uses by the native American Indians. For centuries, native American tribes have used the leaves, roots, and fruits from the blueberry plant for medicinal purposes (Sanchez-Moreno *et al.*, 2003), and blueberries continue to be used in many types of dietary health products as pharmaceutical or food supplements in modern society (Kalt and Dufour, 1997). Many of the uses, once thought to be anecdotal, are now the subject of intensive scientific research. Research on blueberries, which originally focused on antioxidant activity, has now expanded into the areas of anti-inflammation, and cell signaling (Howell, 2009). Blueberry grows well in acid soils of southern Chile, where the Al content is significantly high (Inostroza-Blancheteau *et al.*, 2012).

The free Al content in soil solution can vary by natural (rain) and anthropogenic (liming, fertilization and organic amendment) processes (Inostroza-Blancheteau *et al.*, 2012), which could modify the total content of phenolic compounds in *V. corymbosum*, affecting their antioxidant capacity. For this reason, it is important to determine which might be the ideal conditions in the management of this crop to maximize antioxidant metabolite production.

The aim of this study was to determine the effects of Al on the antioxidant capacity and profile of phenolic compounds in blueberries grown *in vitro*.

# MATERIALS AND METHODS

# Plant material, growth conditions and treatments

In vitro cultures of V. corymbosum cv. Legacy was started from shoot tips of free-pathogen certified plants and its sterilized in 10% of sodium hypochlorite solution and rinsed with sterilized and destilled water, for culture using a Lloyd-McCown media base (Lloyd and McCown, 1980) supplemented with 2.76 mg·L<sup>-1</sup> of hormone 2-iP and 3.0 g·L<sup>-1</sup> of agar phytagel, it mixture was place in a glass flask and was sterilizing in autoclave at 121 °C during 15 minutes. The Al treatments (AlCl<sub>3</sub>), was applied as follow: (1) pH 5.2 (control); (2) Al 100 μM pH 5.2; (3) Al 200 μM pH 5.2.

The cultures were maintained during 7, 14 and 21 days at  $23 \pm 2$  °C with photoperiodicity 16/8 h (day/night). *In vitro* cultivated plantlets with more than 30 days of adaptation to cultivation conditions were defined as starting control (time=0 days), and used to define the physiological base line for all the studied parameters.

# Lipid peroxidation

The level of lipid peroxidation was determined in terms of MDA concentration according to the method of Heath and Packer (1968) with modifications. The concentration of MDA was calculated from the difference of the absorbance at 532 and 600 nm using the extinction coefficient of 155 mmol·L<sup>-1</sup> cm and expressed as nmol·g<sup>-1</sup> FW.

#### Extracts preparation

Fresh materials (0.1g·mL<sup>-1</sup>) were used to prepare the extract using 85% v/v of hydroethanolic solution; the samples were sonicated at 50-60 Hz of frequency during two hours at 25 °C according to the method Rostagno *et al.*, (2002).

# Antioxidant activity

1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical-scavenger spectrophotometric assay

The removal capacity of free radicals of different extracts was evaluated using the radical DPPH technique, described by Shyu *et al.*, 2002. An aliquot of the ethanol extracts was added to DPPH solution, the absorbance decrease was continuously monitored at a wavelength of 517 nm with an UV-visible spectrophotometer (Agilent 8453 UV-Vis), for 240 seconds (Brand-Williams *et al.*, 1995). Results were expressed as % of consumed DPPH.

*Ferric reducing Antioxidant power (FRAP)* 

The FRAP assay measure the ability of the sample to reduce Fe III to Fe II (Benzie and Strain, 1996), through the formation of a blue complex with tripyridyltriazine (TPTZ) which show a maximum absorbance at 593 nm. FRAP reagent was prepared by mixing in the ratio 10:1:1 of acetate buffer (300 mM), TPTZ and FeCl<sub>3</sub> solutions. The FRAP reagent was maintained at 37 °C. Absorbance was measured at 593 nm in a spectrophotometer (Agilent 8453 UV-Vis) to the sample containing 900  $\mu$ L of FRAP reagent, 80  $\mu$ L of water sample and 20  $\mu$ L of ethanolic extract. The measures were expressed in Ascorbic acid equivalents.

#### Total phenolic content (TPC)

The total phenolic content of ethanolic extracts was determined based on the method described by Singleton and Rossi (1965). Results were expressed as Gallic acid equivalents.

#### Analysis of extracts HPLC-DAD

High performance liquid chromatography with diode array detector (HPLC-DAD) was used to separate, identify and determine phenolic compound in extracts ethanolic blueberry tissue. The ethanolic extract was filtered through a 0.45- $\mu$ m membrane and analyzed by HPLC-DAD.

Agilent HPLC-DAD 1100 series equipped with a RP-C18 column at 25 °C was used. The mobile phase is a gradient of acetonitrile (A) and 1% phosphoric acid (B), using the program: time = 0 minutes 10% of A, 5 minutes 25% of A, 8 minutes 35% of A, 15 minutes 60% of A, 17 minutes 35% of A and finally 20 minutes 10% of A; with 120 bar approximately pressure at start, 1 mL/min of flow and 20  $\mu$ L of injection volume using a Reodyne valve, registering the signals at 254, 280, 314 and 340 nm.

#### Statistics analysis

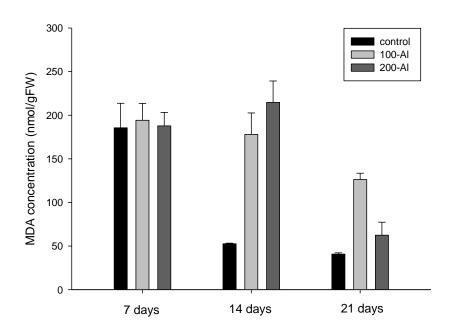
The analysis was carried out using analysis of variance (ANOVA) with Fisher (F) test in all samples to determine significant difference with  $n \ge 3$  in all measurements.

## **RESULTS**

Effect of Al on malondialdehyde (MDA) content

Variations in the content of MDA, as a result of the presence of Al in the growth medium are shown in Figure 1. The results show no significant effect on the MDA content for both treatments after 7 days of exposure to the metal. The greatest accumulation of MDA over the control was observed after 14 days of treatment with 100 and 200  $\mu$ M Al to decrease after 21 days of treatment, even when it remains significantly higher than control.

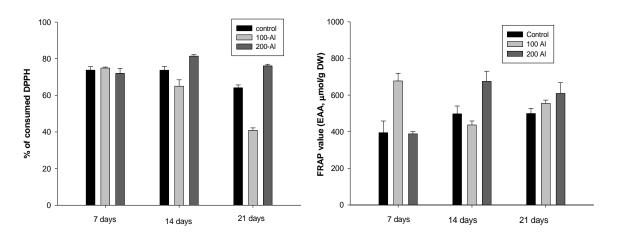
These results suggest that in early stages of exposure to Al (14 days), the variations in the content of MDA in *V. corymbosum* are associated with the presence of Al and not necessarily with the concentration of this element in the growth medium. When the exposure period is extended, MDA levels decreased, possibly as a result of the mechanisms of the plant to counteract the damage caused by lipid peroxidation. The effect of different concentrations of Al is appreciable when the exposure time is prolonged, where higher concentrations of Al generate a better defense against oxidative damage in blueberries, suggesting that the content of MDA in *V. corymbosum*, depends on the exposure time and subsequently the dose of Al in a first stage, showing a differential response to oxidative damage level.



**Figure 1**. Effect of the Al concentration and exposure time on the content of MDA in plantlets of *V. corymbosum* L. Each value is a mean of three samples  $\pm$  1 s.d.

# Antioxidant activity

The antioxidant capacity of ethanolic extracts of *V. corymbosum*, subjected to 100 and 200 uM of Al was evaluated from ethanolic extracts using DPPH and FRAP assays (Figure 2A and B). The antioxidant activity (DPPH assay) showed no significant difference between treatments and control for the first 7 days.



**Figure 2.** Antioxidant capacity of extracts of plantlets of V. corymbosum L. treated with Al. DPPH assay (A) and FRAP assay (B). Each value is a mean of three samples  $\pm$  1 s.d.

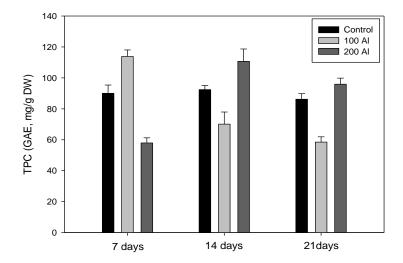
However, clear differences were observed at 14 and 21 days of exposure. 100 uM With Al, DPPH consumption decreased with respect to control up to 21 days, the highest antioxidant activity for the 3 conditions studied, was observed with 200 uM Al which peaked at 14 days (Figure 2A). This behavior suggests that the presence of Al generates a significant variation in consumption of DPPH in plantlets of *V. corymbosum*, which depends on the concentrations of Al present in the medium (Figure 2A).

The evaluation of the reducing power of the ethanolic extracts was performed by the FRAP assay (Figure 2B). The results obtained from this test also show a different response with respect to the Al concentration and exposure time, showing the greatest variations of

this parameter during the first two weeks of exposure. With 100 uM Al it was observed a higher FRAP value at 7 and 21 days, with a minimum at 14 days. With 200 uM Al it was observed a higher FRAP value at 14 and 21 days with a maximum at 14 days (Figure 2B).

# Total phenolic content (TPC)

The content of total phenolic compounds for each extract of blueberry is shown in Figure 3. Control plants showed a slight decrease at day 21, while for the treatment at 100  $\mu$ M, it was observed a maximum at day 7 (113.9 mg·g<sup>-1</sup>DW). At 21 days the total phenolic content was lower than in the control. For treatment at 200  $\mu$ M of Al, it was observed a maximum at day 14 of treatment (110.7 mg·g<sup>-1</sup>DW) and a minimum at day 7, with a value below the control

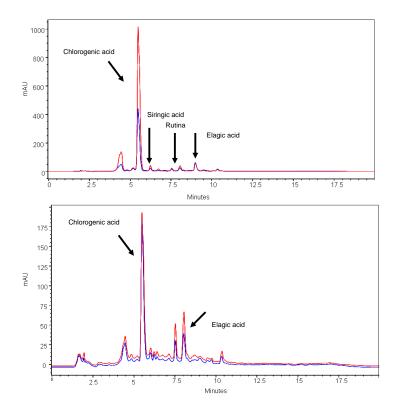


**Figure 3.** Variation in the content of total phenolic compounds (TPC) in plantlets of *V. corymbosum* L. treated with Al. Each value is a mean of three samples  $\pm 1$  s.d.

The different response observed in the TPC content in both treatments, may indicate that the mechanisms of antioxidant response in *V. corymbosum* are more effective when it exceeds a threshold concentration of Al, what would happen about 100 uM of Al in the growth medium.

# Identification of phenolic compound by HPLC-DAD

The identification of phenolic compounds in the blueberry extract was performed using HPLC-DAD. Four compounds were identified, two of them (chlorogenic acid and ellagic acid) were also quantified (Figure 4).



**Figure 4.** HPLC profiles of phenolic compound in blueberries cultivated *in vitro* at 314 nm (up) and 254 nm (bottom). Blue line: control; Red line: 200μM Al<sup>3+</sup>. In both cases evaluation was done after 14 days of treatment.

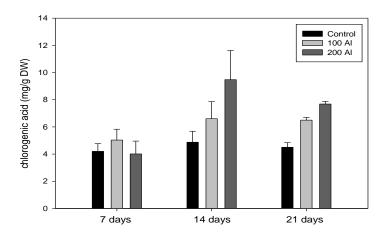
Compared with the control, there was an increase in the content of chlorogenic acid for both AL levels after 7 days with a peak on day 14 (Figure 5). Similar results were reported by Zheng *et al.*, (2003), when tested blueberry plants to high oxygen content, suggesting that these phenolic compounds are biosynthesized in response to stress conditions (Zheng *et al.*, 2003).

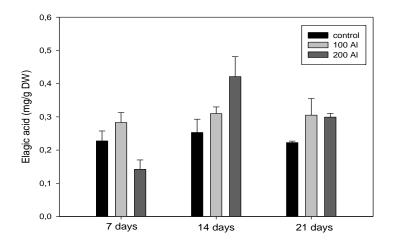
Moreover, with 100  $\mu$ M Al, ellagic acid concentration remained relatively stable over time. In contrast, with a treatment of 200  $\mu$ M of Al this compound significantly increases, with a maximum at 14 days.

#### **DISCUSSION**

The phenomenon of Al toxicity in plants has been extensively studied in nutrient solutions, showing that the first symptom of Al toxicity is inhibition of root elongation, which directly impacts the absorption of nutrients into the plant (Yamamoto *et al.*, 2003; Kochian *et al.*, 2005). However, no research has been reported on the effect of Al in the nutrient medium, the antioxidant capacity of the extracts and phenolic composition of seedlings of blueberry cultured *in vitro*.

The effects of exposure to high Al levels were evaluated in a kinetics study of 21 days. The MDA, a byproduct of lipid peroxidation of membranes, it accumulates in the tissues of plants when these are subjected to stress. Treatments at 100 and 200 mM of Al, induced lipid peroxidation in blueberry cv. Legacy at 14 days of exposure, increasing the content of MDA more than three times with respect to control. This damage would be associated to the presence of the metal in the solution, since no significant differences were observed in the content of MDA between treatments, although it is highly probable that there is a threshold concentration. Research by Yamamoto *et al.*, (2002) found similar behavior in pea plants, where an increase in lipid peroxidation after exposure for 4 hours with Al was observed. Similar behavior was described by Cakmak and Horst (1991), who observed an increase in the MDA concentration in soybean, after 2 days of treatment with different doses of Al.





**Figure 5**. Variation in the content of chlorogenic acids (up) and ellagic acid (bottom), in function of exposure time and the concentration of Al in plantlets of V. corymbosum. Each value is a mean of three samples  $\pm 1$  s.d.

During the exposure period of blueberry plantlets to different doses of Al, a decrease in lipid peroxidation was observed, suggesting that plants were able to acclimate to the presence of Al. Studies by Reyes-Diaz *et al.*, (2010) showed an increase in the content of MDA in two blueberry varieties cv. Legacy and Bluegold growing in Hogland solution modified with Al, concluding that cv. Legacy is Al-tolerant. Our results allow us to suggest

that Al tolerance mechanisms reported by Reyes-Diaz *et al.*, (2010), for this variety of blueberry, mainly be triggered by exposure time.

The ability of a plant to enhance its capacity to remove ROS is a key factor in the mechanism of oxidative stress tolerance. It was observed that the antioxidant capacity of blueberry plantlets varied according to the Al treatment applied. The increase in antioxidant capacity of the extracts on DPPH and FRAP assays could be due to the increase in the total content of phenolic compounds. The smaller effect observed in seedlings treated with 100  $\mu$ M Al, suggests that the non-enzymatic antioxidant defense mechanism is differentially activated according to Al concentration and exposure time to this element, which could be an indication that the effective antioxidant answer occur at high concentrations of Al, as was the case of 200  $\mu$ M Al treatment.

By using HPLC it was possible to identify four phenolic compounds, syringic acid, chlorogenic acid, ellagic acid and rutin. We observed a significant increase in the content of chlorogenic acid at 14 days of treatment with 200 μM Al. Similar results were found by Wang *et al.*, 2009, who describe an increase in the content of chlorogenic acid in blueberry cultures exposed to UV-C radiation. The ellagic acid content increased in both treatments, but the greatest accumulation of this compound was observed at 200 μM Al. The antioxidant capacity of a molecule is reflected in the IC 50 value corresponding to the concentration of compound needed to consume 50 % of the DPPH radical. The IC 50 value for the chlorogenic acid in this study was 4.2 μg, while for the ellagic acid was 11.1 μg, indicating that the chlorogenic acid has a greater ability to remove radicals (data not shown), then, the observed variations in the antioxidant capacity of blueberry plantlets would be attributed mainly to the accumulation of chlorogenic acid (Zheng *et al.*, 2003).

In conclusion, the application of different concentrations of Al, to culture medium of blueberry produced significant effects on the content of phenolic compounds, with a greater response to a higher concentration of the metal in the culture media. Due to the importance of the levels of chlorogenic and ellagic acid in blueberry, it is very important to know factors that regulate its content, as a way to produce fruits with high antioxidant capacity. Then, soils with high levels of aluminum would help to increase the content of antioxidants in blueberry.

#### ACKNOWLEDGMENTS

Financial support of this work by the Center for the Development of Nanoscience and Nanotechnology (CEDENNA), and DICYT-DGT (Universidad de Santiago de Chile, GE. Zúñiga) is gratefully acknowledged

#### REFERENCES

- Álvarez, E., Fernández-Marcos, M.L., Monterroso, C., Fernández-Sanjurjo, M.J., 2005.
   Application of aluminum toxicity indices to soils under various forest species. Forest Ecology and Managment 211, 227 - 239.
- 2. Benzie, I. F. F., Strain, J.J., 1996. The Ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power" the FRAP assay. Analytical Biochemistry 23, 70 76.
- 3. Boscolo, P., Menossi, M., Jorgea, R., 2003. Aluminum-induced oxidative stress in maize. Phytochemistry 62, 181 189.
- 4. Brand-Williams, W., Cuvelier, M.E., Berset, C., 1995. Use of free radical method to evaluate antioxidant activity. LebensmittelWissenschaft und Technology 28, 25 30.

- 5. Cakmak, I., Horst, W.J., 1991. Effect of aluminum on lipid peroxidation, superoxide dismutase catalase and peroxidase activities in roots tips of soybean (*Glicine max*). Plant Physiology 83, 463 468.
- Corrales, I., Poschenrieder, C., Barcelo, J., 2008. Boron-induced amelioration of aluminum toxicity in a monocot and a dicot species. Journal of Plant Physiology 165, 504 - 513.
- Dastmalchi, K., Flores, G., Petrova, V., Pedraza-Peñalosa, P., Kennelly, E.J., 2010.
   Edible neotropical blueberries: antioxidant and compositional fingerprint analysis.
   Journalof Agricultural and Food Chemistry 59, 3020 3026.
- 8. Devi-Rama, S., Yamamoto, Y., Matsumoto, H., 2003. An intracellular mechanism of aluminum tolerance associated with high antioxidant status in cultured tobacco cells.

  Journal of Inorganic Biochemistry 97, 59 68.
- Garratt, L.C., Janagoudar, B.S., Lowe, K.C., Anthony, P., Power, J.B., Davey, M.R.,
   2002. Salinity tolerance and antioxidant status in cotton cultures. Free Radical Biology
   and Medicine 33, 502 511
- 10. Giannakoula, A., Moustakas, M., Syros, T., Yupsanis, T., 2010. Aluminum stress induces up-regulation of an efficient antioxidant system in the Al-tolerant maize line but not in the Al-sensitive line. Environmental and Experimental Botany 67, 487 494.
- 11. Guo, T. R., Zhang, G. P., Zhang, Y. H., 2007. Physiological change in barley plants under combined toxicity of aluminum, copper and cadmium. Colloids and Surface B: Biointerfaces 57, 182 188.
- Heath, R., Packer, L., 1968. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. Archives of Biochemistry Biophysics 125, 189 - 198.

- 13. Howell, A.B., 2009. Update on health benefits of cranberry and blueberry. Acta Horticulturae 810, 779 784.
- 14. Inostroza-Blancheteau, C., Rengel, Z., Alberdi, M., Mora, M. L., Aquea, F., Arce-Johnson, P., Reyes-Díaz, M., 2012. Molecular and physiological strategies to increase aluminum resistance in plants. Molecular Biology Report 39, 2069 2079.
- 15. Kähkönen, M. P., Hopia, A. I., Heinonen, M., 2001. Berry phenolics and their antioxidant activity. Journal of Agricultural and Food Chemistry 49, 4076 4082.
- 16. Kalt, W., Dufour, D., 1997. Health functionality of blueberries. Horticulture Technology 7, 216 221.
- 17. Kochian, L.V., Piñeros, M.A., Hoekenga, A.O., 2005. The physiology, genetics and molecular biology of plant aluminum resistance and toxicity. Plant and Soil 274, 175 195.
- 18. Koca, H., Bor, M., Özdemir, F., Türkan, I., 2007. The effect of salt stress on lipid peroxidation, antioxidative enzymes and proline content of sesame cultivars. Environmental and Experimental Botany 60, 344 351
- 19. Liu, J.W., Magalhaes, J.V., Shaff, J.E., Kochian, L.V., 2009. Aluminum-activated citrate and malate transporters from the MATE and ALMT families function independently to confer Arabidopsis aluminum tolerance. The Plant Journal 57, 389 399.
- 20. Lloyd, G., McCown, B., 1980. Commercially feasible micropropagation of mountain Laurel, Kalmialatifolia, by use of shoot tip culture. Combined Proceedings of the International Plant Propagators Society 30, 421 - 427.
- 21. Ma, J. F., Ryan, P. R., Delhaize, E., 2001. Aluminium tolerance in plants and the complexing role of organic acids. Trends in Plant Science 6, 273 278.

- 22. Munné-Bosch, S., Jubany-Marí, T., Alegre, L., 2001. Drought-induced senescence is characterized by loss of antioxidant defences in chloroplasts. Plant Cell and Environment 24, 1319 1327.
- 23. Poschenrieder, C., Gunse, B., Corrales, I., Barcelo, J., 2008. A glance into aluminum toxicity and resistance in plants. Science of the Total Environment 400, 356 368.
- 24. Reyes-Díaz, M., Inostroza-Blancheteau, C., Millaleo, R., Cruces, E., Alberdi, M., Mora, M. L., 2010. Long-term aluminum exposure effects on physiological and biochemical features of highbush blueberry cultivars. Journal of American Society for Horticulture Science 135, 212 222.
- 25. Rostagno, M.A., Araujo, J. M. A., Sandi, D., 2002. Supercritical fluid extraction of isoflavones from soybean flour. Food Chemistry 78, 111 117.
- 26. Ryan, P. R., Delhaize, E., Jones, D. L., 2001. Function and mechanism of organic acid excretion from plant roots. Annual Review of Plant Physiology and Plant Molecular Biology 52, 527 560.
- 27. Sanchez-Moreno, C., Cao, G., Ou, B., Prior, R. L., 2003. Anthocyanin and proanthocyanidin content in selected white and red wines. Oxygen radical absorbance capacity comparison with nontraditional wines obtained from highbush blueberry. Journal of Agricultural and Food Chemistry 51, 4889 4896.
- 28. Shyu, Y. S., Hwang, L. S., 2002. Antioxidant activity of the crude extract of lignan glycosides from unroasted Burma black sesame meal. Food Research International 35, 357 365.
- 29. Singleton, V.L., Rossi, J. A., 1965. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. American Journal of Enology and Viticulture 16, 144 158.

- 30. Tolrá, R., Barceló, J., Poschenrieder, C., 2009. Constitutive and aluminum-induced patterns of phenolic compounds in two maize varieties differing in aluminum tolerante.

  Journal of Inorganic Biochemistry 103, 1486 1490.
- 31. Yamamoto, Y., Kobayashi, Y., Devi, S. R., Rikiishi, S., Matsumoto, H., 2002. Aluminum toxicity is associated with mitochondrial dysfunction and the production of reactive oxygen species in plant cells. Plant Physiology 128, 63 72.
- 32. Yamamoto, Y., Kobayashi, Y., Devi, S. R., Rikiishi, S., Matsumoto, H., 2003.

  Oxidative stress triggered by aluminum in plant roots. Plant and Soil 255, 239 243.
- 33. Wang, C.Y., Chena, C.T., Wang, S. Y., 2009. Changes of flavonoid content and antioxidant capacity in blueberries after illumination with UV-C. Food Chemistry 117, 426 431
- 34. Zheng, Y., Wang, C. Y., Wang, S. Y., Zheng, W., 2003. Effect of high-oxygen atmospheres on blueberry phenolics, anthocyanins, and antioxidant capacity. Journal of Agricultural and Food Chemistry 51, 7162 7169.

# **Chapter 3**

Effect of the availability of Cd<sup>2+</sup> on the non-enzymatic antioxidant response and the phenolic compounds profile of blueberry (*Vaccinium corymbosum* L.) plantlets cultivated *in vitro*.

<sup>†</sup>The chapter will be subbmited to Molecules (ISI).

# **Abstract**

The presence of trace elements in the soil at levels that can cause toxicity in plants is an important problem in agriculture. One of the main elements that affect the soil-plant system is cadmium, due to its great mobility in the system, in addition to the fact that it is not an essential element for the plants. With the purpose of evaluating the effects of Cd<sup>2+</sup> on antioxidant response and phenolic compounds production, blueberry (*Vaccinium corymbosum* L.) plantlets were cultivated *in vitro* and exposed to 50 and 100 µM Cd<sup>2+</sup> concentrations in the growth medium during periods of 7, 14 and 21 days. A significant effect was seen on the antioxidant response by means of the FRAP test, in addition to an increase in the content of MDA, showing that the Cd<sup>2+</sup> present in the medium caused oxidative damage.

The profile of phenolic compounds in the treated plantlets was established by LC-MS. The main phenolic compound found in the blueberry plantlets was chlorogenic acid, whose abundance in the tissues increased due to the addition of Cd<sup>2+</sup> in the medium. The presence of lower abundance of vanillic acid and quercetin was also detected. These results show that the presence of Cd<sup>2+</sup> in the culture medium results in changes of phenolic compounds profile.

Key words: Vaccinium corymbosum, cadmium, phenolic compounds, oxidative stress.

#### Introduction

Mining and agriculture are important sources of incorporation of trace elements (TEs) in the environment. Mining, on the one hand, through the residues generated by its extractive and purification processes, and agriculture on the other, through the constant

application of fertilizers, biosolids and amendments, seriously modify the ecosystem's biogeochemical cycles (Basu *et al.*, 2006; Molina *et al.*, 2010; Wu *et al.*, 2010).

Among the TEs that affect the soil and water, cadmium (Cd) is often mentioned because of its high toxicity and mobility in the soil/plant system (Sanita and Gabbrielli, 1999; Hassan and Aarts, 2011). The World Health Organization (WHO) has established that the maximum concentration of this element in water for human consumption must not exceed 0.003 mg·L<sup>-1</sup> (WHO, 2013). However, industrial and agricultural activities have enriched the soils and water courses with this element (Molina *et al.*, 2009; Lux *et al.*, 2011). In Chile, Bonomelli *et al.* (2002) analyzed the presence of Cd in phosphate fertilizers imported to Chile during 1999-2000, and found a high Cd content in fertilizers of the Triple superphosphate (TSP). Similar results were reported by Molina et al., concluding that the use of phosphate fertilizers over long periods could increase Cd levels in agricultural soils and plants uptake (Molina *et al.*, 2009; 2010, 2013).

In contrast with other TEs (e.g. Cu, Zn, Mn), Cd is not an essential element for plant growth and development, and it is considered highly toxic to them, even at trace levels (Das *et al.*, 1997; Rascio and Navari-Izzo, 2011). The symptoms associated with high Cd concentrations in plants are biomass reduction, inhibition of root elongation, in addition to perturbations in the capture and distribution of nutrients (Milone *et al.*, 2003; Chamseddine *et al.*, 2009).

It has been reported that the presence of Cd in plants generates, at the cellular level, reactive oxygen species (ROS), which are responsible of membrane damages and degradation of proteins and nucleic acids (Polge *et al.*, 2009). Depending on its concentration, Cd can inhibit or stimulate the activity of various antioxidant enzymes before the symptoms of toxicity become visible in the plant (Fu and Huang, 2001; Liu *et* 

al., 2007). The oxidative stress produced by Cd has been observed as an increase of the lipid peroxidation products measured from the accumulation in the tissues of malondialdehyde (MDA), which is a by-product of the oxidation of the polyunsaturated fatty acids of the membranes, changes in the activity of antioxidant enzymes like superoxide dismutase or ascorbate peroxidase, in addition to the degradation of chlorophyl (Djebali et al., 2005; Gratao et al., 2008; Fidalgo et al., 2011). It has been reported that plants have different mechanisms to tolerate the presence of Cd, which include the metal's exclusion from the roots and retention of the metal in the vacuole through complexes formed with organic ligands and detoxification processes (Clements, 2006; Hédiji et al., 2010).

Other studies indicate that the plants have strategies that allow them to control ROS, one of which is based on the production of secondary metabolites with antioxidant properties (Mourato *et al.*, 2012). The secondary metabolites are synthesized by the plant as a defense mechanism against biotic stress, but they may also play a role in the plant's physiological functioning when it is in the presence of a high concentration of heavy metals (Close and McArthur, 2002; Fernández *et al.*, 2013).

Participation of antioxidant molecules as a response of the plant to intoxication by Cd may be a defense mechanism to the stress caused by the metal, but there are no important reports on the effects caused by changes of the levels and profiles of antioxidant molecules.

Blueberries (*V. corymbosum* L.) have become important in human nutrition in recent years, due to the high content of antioxidant molecules in their fruits and leaves (Giovanelli and Buratti, 2009; You *et al.*, 2011). In Chile the cultivation of blueberries occurs mainly in the central and southern zones (33° to 41° south latitude), developing well

in acid soils with pH between 4.8 and 5.5. Because this species has positioned itself in recent years in world agriculture, it is important to study the effect of its exposure to high Cd concentrations, managing the conditions of the plant's growth medium.

The objective of this work was to evaluate the effects on the antioxidant capacity and the profile of phenolic compounds, in the early growth stages, and the generation of MDA as an indicator of the oxidative damage in blueberry plantlets cultivated *in vitro* exposed to Cd in the nutritive solution.

#### 2. MATERIALS AND METHODS

# 2.1 Plant material, growth conditions and treatments

To start the *in vitro* culture, Blueberry plantlets were obtained through the sterilized foliar segments replication (2 cm long) of two months of growth pre-cultivated plantlets. 25 mL of Woody Plant culture medium (Lloyd and McCown, 1980) were added to each flask of culture (6 cm in diameter, 10 cm high and 197.82 cm³) containing the following components: (mg·L¹): NH<sub>4</sub>NO<sub>3</sub> 400 mg, CaCl<sub>2</sub>·2H<sub>2</sub>O 96 mg, MgSO<sub>4</sub>·7H<sub>2</sub>O 370 mg, K<sub>2</sub>SO<sub>4</sub> 990 mg, KH<sub>2</sub>PO<sub>4</sub> 170 mg, Ca(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O 556 mg, H<sub>3</sub>BO<sub>3</sub> 6.2 mg, Na<sub>2</sub>EDTA·2H<sub>2</sub>O 37.2 mg, CuSO<sub>4</sub>·5 H<sub>2</sub>O 0.25 mg, FeSO<sub>4</sub>·7 H<sub>2</sub>O 27.8 mg, MnSO<sub>4</sub>·4 H<sub>2</sub>O 22.3 mg, ZnSO<sub>4</sub>·7 H<sub>2</sub>O 8.6 mg, Na<sub>2</sub>MoO<sub>4</sub>·2 H<sub>2</sub>O 0.25 mg. The culture medium was supplemented with 2.76 mg·L¹ of 2-iP hormone (2-isopentenyl adenine), sucrose was used as carbon source (15 g·L¹) and as a gelling agent agar Phytagel (3 g·L¹). The pH was adjusted to 5.2. The culture medium was sterilized in an autoclave for 15 minutes at 121 °C (394 K). Cultures were maintained in cultivation cabinet at 21±2 °C (294±2 K) and photoperiod of 16 hours of light/8 hours of darkness. Processes of sterilization and explants sowing were conducted in a laminar flow cabinet in aseptic conditions. Once established *in vitro* culture (care and treatment), we

used a completely randomized design, where the experimental unit corresponded to 6 clones of plantlets of blueberries (6 plantlets: 1 sample), each sample was carried out in triplicate. *In vitro* cultivated plantlets with more than 30 days of adaptation to cultivation conditions were defined as starting control (time=0 days), and used to define the physiological base line for all the studied parameters.

To calculate the chemical speciation of Cd and Al in the culture medium of the plantlets, the software GEOCHEM PC was used (Parker *et al.*, 1995).

# 2.2. Extracts preparation.

Fresh materials  $(0.1\text{g}\cdot\text{mL}^{-1})$  are used to prepare the extract using 85% v/v of hydroethanolic solution; the samples were sonicated at 50-60 Hz of frequency during two hours at 25±2 °C (298±2 K) according to the method described by Adam (2009).

# 2.3. Antioxidant activity

2.3.1. 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical-scavenger spectrophotometric assay.

The free radicals removal capacity of each extract was evaluated using the radical DPPH technique, described by Brand-Williams *et al.* (1995) with some modifications. Briefly, 20 μL of the ethanol extract was added to 980 μL of DPPH solution (25 mg·L<sup>-1</sup>), the absorbance was continuously monitored at 517 nm using a UV-visible spectrophotometer (Agilent 8453 UV-Vis), for 240 seconds (Brand-Williams *et al.*, 1995; Huang *et al.*, 2005). Results were expressed as % of consumed DPPH.

# 2.3.2. Ferric reducing antioxidant power (FRAP).

The FRAP assay measures the ability of the sample to reduce Fe<sup>3+</sup> to Fe<sup>2+</sup> (Benzie and Strain, 1996), followed by the absorbance at 593 nm of the blue complex formed with tripyridyltriazine (TPTZ). FRAP reagent was prepared by mixing acetate buffer (300 mM), TPTZ solution (10 mM in HCl) and FeCl<sub>3</sub> solution (20 mM) at a 10:1:1 ratio.

The FRAP reagent was maintained at  $37\pm2^{\circ}$  C ( $310\pm2$  K). Absorbance of samples containing 900  $\mu$ L of FRAP reagent, 80  $\mu$ L of water and 20  $\mu$ L of ethanolic extract, was measured at 593 nm in an Agilent 8453 UV-Vis spectrophotometer. Results were expressed in ascorbic acid equivalents.

#### 2.4. Total phenolic content (TPC)

The TPC of ethanolic extracts was determined through the method described by Singleton and Rossi (1965). Results were expressed as mg Gallic acid equivalents.

2.5 Analysis of extracts by Liquid Chromatography-Triple Quadrupole Mass Spectrometry (LC-MS).

Extracts were analyzed using a 1120 Compact LC-MS, Agilent Technologies (Santa Clara, Ca). LC was in-line with a 6400 Triple quadrupole (Agilent Technologies) with electrospray (ESI). Compounds were separated using a SUPELCOSIL LC-PAH RP-C18 column (15 cm x 4.6 mm; particle size 5μm) and a mobile phase of acetonitrile as eluent A and 0.1% formic acid as eluent B. All solutions were filtered through a 0.45 μm filter and degassed for 15 min. Elution was performed using a linear gradient from 10:90 (A: B) to 70:30 over 20 min at a flow rate of 0.3 ml·min<sup>-1</sup>. For analysis of the phenolic compounds

the MS was operated in negative ion mode, capillary temperature of 200°C (473 K), spray voltage 4 kV, and data were acquired in MS and MS/MS scanning modes.

The multiple reactions monitoring method (MRM) was used for phenolic compounds analysis; the method involves a phenolic compounds data base considering their corresponding molecular ions and precursors.

## 2.6. Lipid peroxidation

The lipid peroxidation level was determined in terms of MDA concentration, according to the method of Heath and Parker (1968) with modifications. The concentration of MDA was calculated from the difference of the absorbance at 532 and 600 nm using the extinction coefficient of 155 mmol·L<sup>-1</sup> cm and expressed as nmol·g<sup>-1</sup> FW.

# 2.7. Statistics analysis.

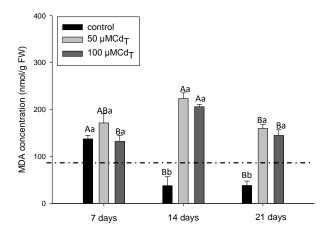
All data shown in the study are mean  $\pm$  sd. The measurement was performed in sextuple for all measured parameters. Statistical analysis was performed by two-way analysis of variance (Two-way ANOVA) and comparisons between culture medium were performed using the Tukey Averages Multiple Comparison Test.

#### 3. RESULTS AND DISCUSSION

To achieve the objectives of this study and evaluate the behavior of blueberry plantlets when exposed to Cd, the study parameters considered were MDA content, the response mechanisms to oxidative stress estimated from the determination of the antioxidant capacity (DPPH and FRAP), and the production of phenolic compounds determined by LC-MS.

# 3.1 Cd-induced changes in malondialdehyde (MDA) content

The effects of oxidative stress caused by the presence of Cd in the growth medium were determined by the accumulation of MDA in the plants' tissues.



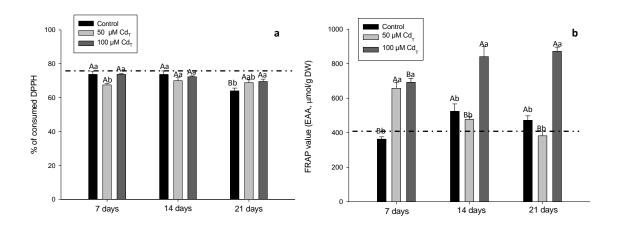
**Fig. 1.** Effect of Cd concentration on oxidative stress in blueberries measured as MDA. Data are mean  $\pm$  sd; n = 6. Different letters (lower case) indicate statistical differences among means from treatments and time after ANOVA and Tukey HSD post hoc test (P< 0.05). The dashed line indicates the level of MDA measured for control when initiating treatment.

Figure 1 shows the results of the Cd treatments and the control at the different treatment times. It is seen that the control shows a decrease of MDA content with time. This behavior suggests that during the first 7 days of treatment the blueberry plantlets present a degree of stress because of the change to the *in vitro* culture medium, although no significant variation between treatments (p < 0.05). After the acclimation period, MDA content decreases, staying stable during the rest of the study.

In the case of plantlets exposed to Cd, an effect of the presence of the metal on the MDA content is seen after 14 days of exposure, with MDA levels higher than those of the control (Fig. 1) (p < 0.05). The presence of Cd in the growth medium seems to be more important than its concentration in the response of blueberry plantlets in the early stages of Cd exposure (14 days), since no significant differences were found between the treatments (p < 0.05). At longer exposure times (21 days), the MDA content decreases in the plantlets exposed to Cd, but it remains significantly higher than in the control (p < 0.05). This behavior agrees with the observations of Ge et al. (2012), and Chaoui et al. (1997), who found that MDA content reaches its peak after 2 week of exposure to Cd in populus nigra and *Phaseolus vulgaris*. Another effect reported in the literature and observed in our study is that MDA content significantly decreases as exposure time and Cd concentration increase, indicating that blueberries develop better defense mechanisms against lipid peroxidation at higher Cd concentrations (Metwally et al., 2005; Hegedus et al., 2001, Liu et al., 2011) These results suggest that the action of these enzymatic and non-enzymatic mechanisms strongly depends on two factors: i) exposure time (early stages) and ii) initial total Cd concentration (Cd<sub>T</sub>) added to the culture medium (late stages), with a differentiated response at oxidative damage level. In this context it has been reported that the MDA content in the plant's tissues due to the presence of Cd in the growth medium depends on both the genotype and the mechanisms presented by the species to defend itself from the stress (Metwally et al., 2005; Saidi et al., 2013; Fernández et al., 2013).

# 3.2 Antioxidant activity

The antioxidant activity of blueberries ethanol extracts was measured by the DPPH and FRAP tests, which evaluate the ability to trap the diphenylpicrylhydrazyl free radical (DPPH) and to reduce Fe<sup>3+</sup> to Fe<sup>2+</sup>, respectively.



**Fig. 2.** Effect of Cd concentration on antioxidant activity of ethanolic extracts. (a) DPPH free radical scavenging (DPPH consumed), (b) FRAP test (ascorbic acid equivalents). Data are mean  $\pm$  sd; n = 6. Different letters (lower case) indicate statistical differences among means from treatments and time after ANOVA and Tukey HSD post hoc test (P< 0.05). The dashed lines indicate the level of % DPPH and FRAP value respectivily measured for control when initiating treatment.

The results of the DPPH test only show changes of the antioxidant capacity with respect to the control after 7 days for 50  $\mu$ M Cd<sub>T</sub> (p < 0.05), where a decrease of the antioxidant capacity is seen by DPPH, and after 21 days for the 100  $\mu$ M Cd<sub>T</sub> treatment, with respect to the control, with a slight increase of the antioxidant capacity of the extract. No significant differences were found after 14 days of treatment (p > 0.05) for 50 and 100  $\mu$ M Cd<sub>T</sub> (Fig. 2a). These results show that the ethanol extracts have a high capacity for

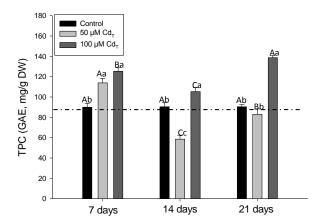
trapping free radicals, and this is not largely affected by the presence and concentration of Cd.

The FRAP test showed significant differences between treatments; the reducing capacity significantly increases in the presence of Cd during the first 7 days (p < 0.05). As the study period got longer, the response to the oxidative damage generated by the presence of Cd is now associated to the metal's concentration. Both, the control as well as the samples exposed to 50  $\mu$ M Cd<sub>T</sub>, did not show significant differences between them (p > 0.05), while in the seedlings subjected to 100  $\mu$ M Cd<sub>T</sub>, the antioxidant capacity measured by the FRAP test increased 1.8 and 2.0 times after 14 and 21 days of exposure with respect to the control and the treatment with 50  $\mu$ M Cd<sub>T</sub> (Fig. 2b) (p < 0.05). These results suggest that Cd induces the formation of compounds with reducing ability, as a possible defense mechanism. Different studies indicate that this parameter is sensitive to the presence of Cd, with positive and negative relations in terms of exposure time and metal concentration, showing that reducing ability depends on the variety of plants studied (Das *et al.*, 1997; Cherif *et al.*, 2011; Douchiche *et al.*, 2012; Panitlertumpai *et al.*, 2013).

# 3.3 Total phenolic content (TPC) and LC-MS analysis

TPC in each blueberry extract is shown in Fig. 3. In the control it remains without major variations during the whole exposure time (p > 0.05), under the treatment with 50  $\mu$ M of Cd<sub>T</sub> there is a maximum at 7 days (113.87 mg·g<sup>-1</sup> DW), and under the treatment with 100  $\mu$ M of Cd<sub>T</sub> the TPC exceeds the control during the whole exposure time. The response seen in TPC in the blueberry extracts, under both treatments, may indicate more than one antioxidant response mechanism. One occurs when a threshold concentration is

exceeded, in this case at  $100~\mu M$  Cd<sub>T</sub> in the growth medium, but there may also be a fast response to the presence of the metal that was seen at 7 days of exposure at both concentrations, in addition to a response over time (Fig. 3) Results shown that TPC decreases at 14 days for both treatments and increases at 21 days of treatment, suggesting that synthesis mechanism of phenolic compounds is reduced at 14 days due to their high production during the first days of treatment (early response), while at 21 days of exposure the blueberry again shows an increase of the synthesis of these metabolites in both treatments (p < 0.05).



**Fig. 3**. Effect of Cd concentration on TPC variation in Vaccinium corymbosum L. Values are mean  $\pm$  standard deviation; n = 6. Different letters (lower case) indicate statistical differences among means from treatments and time after ANOVA and Tukey HSD post hoc test (P< 0.05). The dashed line indicates the level of TPC measured for control when initiating treatment.

The LC-MS analysis allowed the identification of 16 different phenolic compounds (Table 1). The MRM method is extensively used as an analytical tool in these studies (Soong and Barlow, 2005; Romani *et al.*, 2006; Serrano *et al.*, 2013). The relative

abundance of the identified phenolic compounds changes with the presence of and the exposure time to Cd in the growth medium. (Table 1).

 Table 1. Phenolic compounds in ethanolic extracts for control and Cd exposed blueberry plantlets.

-							Control			50 μM Cd <sub>1</sub>	r		100 μM Cd	Т
						7	14	21	7	14	21	7	14	21
Phenolic Compound	Molecular	Molecular	t <sub>R</sub> (min)	m/z ES (-)	Main fragment ion (M-H -X)					Days				
	Formula	Weight			(NI-II -A)				Re	lative abun	dance			
										$((\mathbf{M}^{-}) m/z)$	:)			
Gallic acid	$C_7H_6O_5$	170	1.25	169	125 [(M-H)-CO <sub>2</sub> ]	0.25	0.23	0.27	-	-	-	0.08	0.15	0.17
Vaillinic acid	$C_8H_8O_4$	168	1.25	167	123 [(M-H)- CO <sub>2</sub> ]	-	-	-	0.69	0.44	0.65	0.56	0.53	0.15
Caffeic acid hexoside	$C_{15}H_{18}O_9$	342	1.53	341	$179[(M-H)-C_6H_{10}O_5]$	1.52	1.58	1.45	-	-	-	1.12	1.55	0.87
Chlorogenic acid	$C_{15}H_{18}O_{9}$	354	3.91	353	$191[(M-H)-C_7H_{11}O_6]$	85.5	82.8	85.12	83.6	82.55	83.1	96.3	92.78	95.75
p-Coumaric acid	$C_9H_8O_3$	164	3.97	163	119[(M-H)- CO <sub>2</sub> ]	0.77	0.81	0.74	1.41	0.72	0.67	0.35	0.56	0.87
4-Hydroxybenzoic acid	$C_7H_6O_3$	138	4.23	137	93 [(M-H)- CO <sub>2</sub> ]	0.46	0.48	0.44	0.33	0.24	0.28	0.3	0.39	0.14
Quercetin-3-O-hexoside	$C_{21}H_{20}O_{12} \\$	464	4.83	463	$301[(M-H)-C_6H_{10}O_5]$	5.86	6.11	5.61	4.63	7.02	6.96	0.35	1.51	0.49
Hesperidin	$C_{28}H_{34}O_{15}$	610	5.00	609	$301[(M-H)-C_{12}H_{20}O_9]$	2.32	2.42	2.22	4.97	5.16	4.49	0.17	0.77	0.19
Sinapic acid	$C_{11}H_{12}O_5$	224	6.80	223	179 [(M-H)- CO <sub>2</sub> ]	0.24	0.25	0.23	-	-	-	0.48	0.70	0.50
Ferulic acid	$C_{10}H_{10}O_4$	194	8.98	193	178 [(M-H)-CH <sub>3</sub> ]	0.15	0.15	0.14	0.27	0.09	0.14	-	-	-
Phloridzin	$C_{21}H_{24}O_{10} \\$	436	9.38	435	273 [(M-H)- $C_6H_{10}O_5$ ]	0.45	0.47	0.43	0.72	0.32	0.35	-	-	-
Quercetin	$C_{15}H_{10}O_7$	302	16.84	301	151 [(M-H)-C <sub>8</sub> H <sub>6</sub> O <sub>3</sub> ]	0.37	0.38	0.35	0.99	0.40	0.42	-	-	-
Hydroxybenzoic acid- O-hexoside	$C_{13}H_{15}O_{8}$	300	22.64	299	137 [(M-H)- $C_6H_{10}O_5$ ]	0.46	0.48	0.44	1.03	0.60	0.58	0.08	0.77	0.50
Methoxy carnosol	$C_{21}H_{28}O_5$	360	22.70	359	329 [(M-H)-CH <sub>2</sub> O]	0.15	0.15	0.14	0.32	0.22	0.20	-	-	-
Luteolin	$C_{15}H_{10}O_6$	286	22.95	285	$267[(M-H)-H_2O]$	-	-	-	-	-	-	0.21	0.31	0.36
Catechin	$C_{15}H_{14}O_6$	290	27.00	289	$109 [(M-H)-C_9H_8O_4]$	2.53	2.64	2.42	1.03	2.23	2.16	-	-	-

For a better discussion of the effects of the presence of Cd on the production of phenolic compounds, they were divided into three groups according to their presence and relative abundance when compared to the control.

- **Group I:** Phenolic compounds whose relative abundance increases: p-coumaric acid, quercetin-3-O-hexoside, hesperidin, and hydroxybenzoic acid-O-hexoside, and chlorogenic acid, which is the most representative compound found in the blueberry plantlets (Prior *et al.*, 2001; You *et al.*, 2011).
- **Group II:** Phenolic compounds absent in the control but detected in one of the treatments: vanillic acid and luteolin.
- **Group III:** Compounds absent in one or both treatments (50 μM Cd<sub>T</sub> and/or 100 μM Cd<sub>T</sub>) but present in the control: gallic, ferulic, and sinapic acids, phloridzin, quercetin, methoxycarnosol, catechin, and caffeic acid hexoside.

In this context the relative abundance increasing of chlorogenic acid (Group I) allows to state that Cd increases its production (100 µM Cd<sub>T</sub>), understanding that the function of chlorogenic acid is a protective response to the oxidative damage caused by the metal. Similar behaviors have been reported in *Matricaria chamomilla* treated with Cu (Kovacik and Klejdus, 2008), while in *Brassica juncea* chlorogenic acid decreases when it is exposed to Cd (Irtelli and Navari-Izzo, 2006). It was found that in blueberry plantlets Cd induces the biosynthesis of some phenolic compounds (Group II), mainly vanillic acid, which has antioxidant characteristics, although in a lower degree than chlorogenic acid. The results indicate that the presence of Cd<sup>2+</sup> modifies the phenolic compound profile in blueberries. The absence of some of them (Group III) like gallic and ferulic acids, which

are known as antioxidants (Kanski *et al.*, 2002; Irtelli and Navari-Izzo, 2006), may be related to processes of consumption or inhibition of their synthetic route.

The biosynthesis of phenolic compounds in blueberries is conditioned by the exposure time to Cd and its presence in the culture medium.

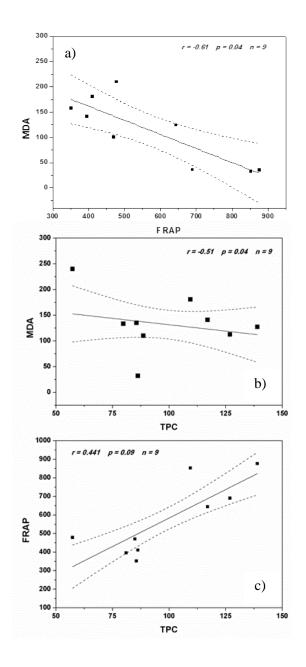
The profile variation of phenolic metabolites may be a defense mechanism against induced stress, observing that during the study time the control treatment showed no significant variations in the relative abundance of detected compounds (Table 1). It cannot be established whether the presence of Cd in the metabolism of blueberries affects the different biosynthetic routes of these phenolic compounds, such as, for example, those produced as derivatives of cinnamic or benzoic acids.

### 3.5 Correlation between antioxidant response parameters and the content of MDA

Possible correlations between oxidative stress (MDA) and antioxidant response (FRAP, DPPH and TPC) generated by Cd exposure were evaluated (Fig. 4). From these results, it is possible to establish that the presence of Cd in the culture medium affected the MDA content and the cranberry seedlings antioxidant response.

In general the lipid peroxidation measured by changes in the content of MDA significantly correlated with antioxidant response measured by the FRAP assay (p < 0.05), while MDA and DPPH no significant relationship between the two variables exists (p > 0.05). The total phenolic content (TPC) showed a negative correlation with MDA (p < 0.05), whereas the values obtained from the FRAP assay, in this parameter a high correlation and significance (p > 0.05) was observed, which suggests that biosynthesized

phenolic compounds in studies conditions play an important role in cranberry seedlings protection with the presence of Cd in the culture medium.



**Fig. 4.** Relationship between the lipid peroxidation and FRAP (a), MDA and TPC (b) and FRAP and TPC (c). Pearson's coefficients are indicated.

## 3.4 Chemical speciation of Lloyd-McCown nutrient medium

Every culture medium, because of its composition, has a high complexing capacity, and therefore it tends to modify significantly the distribution of the chemical forms in which the compounds present are found. To know the actual form in which the Cd added to the nutrient medium is found it is necessary to know its chemical speciation (Table 2), for which the GEOCHEM-PC program (Parker *et al.*, 1995) was used. From those results, the concentration of free Cd<sup>2+</sup> was 1.168 and 16.88  $\mu$ M (for a total supplied concentration of 50 and 100  $\mu$ M), while it shows 0.5889 and 8.475  $\mu$ M Cd activity.

**Table 2.** Percent distribution of Cd in the Lloyd-McCown medium culture, for a concentration of 50 and 100  $\mu$ M of total Cd (Cd<sub>T</sub>) added (Cd-anion means Cd complexed by the corresponding anion, Parker *et al*, 1995)

Charina	50 μM Cd	l <sub>T</sub>	$100~\mu M~Cd_T$				
Species	Concentration µM	Activity µM	Concentration µM	Activity µM			
Free Cd <sup>2+</sup>	1.17	0.59	16.88	8.48			
Cd-SO <sub>4</sub> <sup>2</sup> -	0.56	0.57	8.13	8.23			
Cd-EDTA	47.86	24.1	74.13	36.7			
Cd-Cl	0.08	0.07	1.29	1.07			
Cd-NO <sub>3</sub>	0.01	0.01	0.17	0.14			
Cd-PO <sub>4</sub> <sup>3-</sup>	0.01	0.01	0.14	0.15			

It was determined that the cadmium is found mostly complexed by EDTA and  ${\rm SO_4}^{2-}$  achieving around 80% - 95% formation of highly stable complexes. Through the

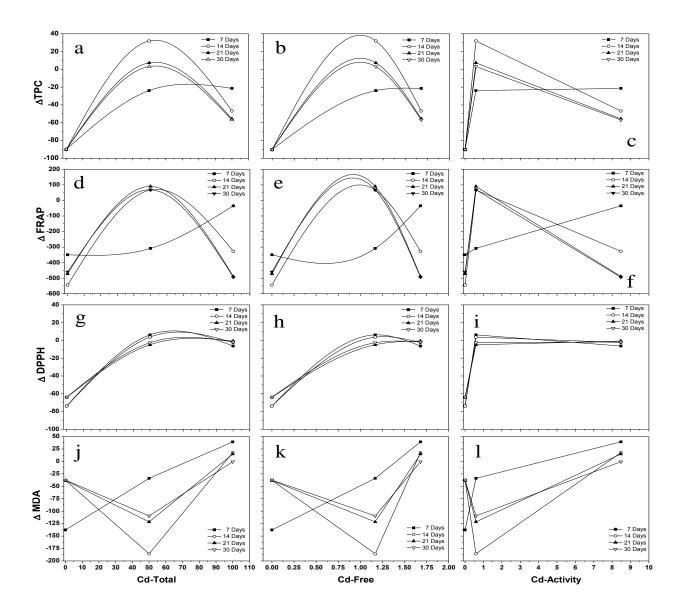
GEOCHEM-PC program, no formation of any complex in which Cd and other cations present in the Lloyd-McCown solution participate simultaneously could be established. The symbol  $Cd_F$  was used to refer to the concentration of free Cd in the medium and its activity by  $Cd_A$ .

3.5 Evidence of the impact of Cd species on the response and damage in Vaccinium corymbosum L.

A more significant way of looking the changes associated with the total concentration of added Cd (Cd<sub>T</sub>) and with the most significant chemical forms present in the medium (Cd<sub>F</sub> and Cd<sub>A</sub>) on the effects observed for blueberries. (TPC, FRAP, DPPH and MDA) is to represent the derivative in the plant's response ( $\Delta$ TPC,  $\Delta$ FRAP,  $\Delta$ DPPH) and the damage ( $\Delta$ MDA), respectively, as a function of Cd<sub>T</sub>, Cd<sub>F</sub> and Cd<sub>A</sub> (Fig. 5).

Fig. 5 shows similar responses between antioxidant capacity and oxidative damage when the differences between them are analyzed vs.  $Cd_T$  and  $Cd_F$ , while  $Cd_A$  did not show the same behavior. Under any condition, the observed response is conditioned by the species or reagent that is in the lowest concentration. Therefore, from the results it is possible to establish that  $Cd_F$  is the Cd species responsible for its effects on blueberries, and not  $Cd_T$ , which can be explained by the size of the formation constant of the Cd-EDTA complex ( $K_f$  =  $2.9 \times 10^{16}$ ), Cd form which is not easily available to the plant, but represents anyway the most significant chemical form of Cd in the growth medium.

In general, the effect on the physiological parameters of blueberries will depend on the amount of Cd (or another metal) that is in its free form, and this is directly related with the complexing agents available in the growth medium which, under natural conditions, will include the different organic compounds produced by blueberries exudation.



**Fig. 5.** Representation of derivative of the antioxidant response ( $\Delta$ TPC,  $\Delta$ FRAP,  $\Delta$ DPPH) and the damage ( $\Delta$ MDA) as a function of Cd in the growth medium.

#### **CONCLUSIONS**

The results obtained in this research suggest that phenolic compounds play an important role in response to the damage produced by the different species of Cd present in the growth medium, mainly free Cd<sup>3+</sup>.

Chemical speciation allowed showing the importance of carrying out this analysis in multiple composition systems, like culture medium, in nutrient solutions and/or in *in vitro* cultivation, where complexing agents significantly reduce the free concentration of cationic elements in the system.

The presence of Cd to the growth medium resulted in a variation in the MDA content when control and the Cd treatments were compared. The greatest oxidative damage occurs at 14 days of exposure at 50 and 100  $\mu$ M Cd<sub>T</sub>. The response to oxidative damage, quantified by the DPPH and FRAP tests, suggest that one of the defense mechanisms used by the blueberry plantlets is the production of reducing molecules, and this would explain the high FRAP values, but this response was not directly proportional to the damage, showing an early response in both treatments at 7 days of exposure compared to the control, and this was not seen at 14 and 21 days for a 50  $\mu$ M Cd<sub>T</sub> concentration, where no important differences with respect to the control were observed. The high FRAP values seen in the blueberry seedlings treated with 100  $\mu$ M Cd<sub>T</sub> may correspond to a selective late response mechanism that is triggered at a higher concentration range, leading to the conclusion that blueberryhad two responses: a first one at 7 days due to the presence of the metal rather than to its concentration, and a second selective response to concentration at 14 and 21 days for 100  $\mu$ M Cd<sub>T</sub>, where the Cd<sub>F</sub> concentration is 13 times greater than at 50  $\mu$ M Cd<sub>T</sub>.

The application of different  $Cd_T$  concentrations to the blueberry culture medium resulted in noticeable effects on the contents of phenolic compounds. The plant's response to the presence of the metal was different with respect to the concentration of  $Cd_T$ , showing a greater response at higher concentrations in the medium.

The  $Cd_F$  induces changes in the production of phenolic compounds, as shown by an increase of their proportion compared to the control medium. However, further research is proposed in this relation and concerning the presence of other metals, to see if there is synergism or antagonism with the plant's response in the production of phenolic compounds.

## Acknowledgements

Financial support of this work by the Center for the Development of Nanoscience and Nanotechnolgy (CEDENNA). K. Manquián acknowledges a scholarship from UFRO (Chile) and DICYT-DGT (Universidad de Santiago de Chile, GE. Zúñiga).

#### REFERENCES

- Adam, M., Dobiáš, P., Eisner, A., Ventura, K., 2009. Extraction of antioxidants from plants using ultrasonic methods and their antioxidant capacity. Journal of Separation Science 32, 288-294.
- Basu, A.J. and Van Zyl, D.J.A., 2006. Industrial ecology framework for achieving cleaner production in the mining and minerals industry. Journal of Cleaner Production 14, 299-304

- 3. Benzie, I. F. F. and Strain, J.J., 1996. The Ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power" the FRAP assay. Analytical-Biochemistry 239, 70-76.
- 4. Bonomelli, C., Bonilla, C., Valenzuela, A., Saavedra, N., 2002. Presencia de Cadmio en fertilizantes fosfatados de diferente procedencia comercializados en Chile, segunda temporada. Simiente 72, 9-16.
- 5. Brand-Williams, W., Cuvelier, M. E., Berset, C., 1995. Use of free radical method to evaluate antioxidant activity. Lebensmittel Wissenschaft und Technologie 28, 25–30.
- 6. Chamseddine, M., Wided, B.A., Guy, H., Marie-Edith, C., Fatma, J., 2009. Cadmium and copper induction of oxidative stress and antioxidative response in tomato (*Solanum lycopersicon*) leaves. Plant Growth Regulator 57, 89–99.
- 7. Cherif, J., Mediouni, C., Ammar, W. B., Jemal, F., 2011. Interactions of zinc and cadmium toxicity in their effects on growth and in antioxidative systems in tomato plants (*Solanum lycopersicum*). Journal of Environmental Sciences. 23, 837–844.
- 8. Chaoui, A., Mazhoudi, S., Ghorbal, M.H., El Ferjani, E., 1997. Cadmium and zinc induction of lipid peroxidation and effects on antioxidant enzyme activities in bean (*Phaseolus vulgaris* L). Plant Science 127, 139-147.
- 9. Close, D. C., and McArthur, C., 2002. Rethinking the role of many plant phenolics Protection from photodamage not herbivores? Oikos 99, 166–172.
- Clements, R., 2006. Organic Acids in Citrus Fruits. II. Seasonal Changes in the Orange. Journal of Food Science 29, 281-286.
- 11. Das, P., Samantaray, S., Rout, G.R., 1997. Studies on cadmiun toxicity in plants: A Review. Environmental Pollution 1, 29-36.

- 12. Djebali, W., Zarrouk, M., Brouquisse, R., Kahoui, S., Limam, F., Ghorbel, M.H., Chaïbi, W., 2005. Ultrastructure and lipid alterations induced by cadmium in tomato (*Lycopersicon esculentum*) chloroplast membranes. Plant Biology 7, 258-368.
- 13. Douchiche, O., Chaibi, W., Morvan, C. 2012. Cadmium tolerance and accumulation characteristics of mature flax, cv. Hermes: Contribution of the basal stem compared to the root. Journal of Hazardous Materials 235, 101-107.
- 14. Fernández, R., Bertranda, A., Reisc, R., Mourato, M.P., Martinsc, L.L., González, A., 2013. Growth and physiological responses to cadmium stress of two populations of *Dittrichia viscosa* L. Journal of Hazardous Materials 244, 555-562.
- Fidalgo, F., Freitas, R., Ferreira, R., Pessoa, A. M., Teixeira, J., 2011. Solanum nigrum
   L. antioxidant defence system isozymes are regulated transcriptionally and posttranslationally in Cd-induced stress. Environmental and Experimental Botany 72, 312–319.
- 16. Fu, J. and Huang, B., 2001. Involvement of antioxidants and lipid peroxidation in the adaptation of two cool-season grasses to localized drought stress. Environmental and Experimental Botany 45, 105–114.
- 17. Ge, W., Jiao, Y.Q., Su, B.L., Qin, R., Jiang, W.S., Liu, D.H. 2012. Cadmium-mediated oxidative stress and ultrastructural changes in root cells of poplar cultivars. South African Journal of Botany 83, 98–108.
- 18. Giovanelli, G. and Buratti, S., 2009. Comparison of polyphenolic composition and antioxidant activity of wild Italian blueberries and some cultivated varieties. Food Chemistry 112, 903–908.

- 19. Gratao, P.L., Monteiro, C.C., Antunes, A.M., Peres, L.E.P., Azevedo, R.A., 2008. Acquired tolerance of tomato (*Lycopersicon esculentum* cv.Micro-Tom) plants to cadmium induced stress. Annual of Applied Biology 153, 321–333.
- 20. Hassan, Z., and Aarts, M.G.M., 2011. Opportunities and feasibilities for biotechnological improvement of Zn, Cd or Ni tolerance and accumulation in plants. Environmental and Experimental Botany 72, 53–63.
- 21. Heath, R. and Packer, L., 1968. Photoperoxidation in isolated chloroplasts. I. Kinetics and stoichiometry of fatty acid peroxidation. Archives of Biochemistry and Biophysics 125, 189-1986.
- 22. Hédiji, H., Djebali, W., Cabasson, C., Maucourt, M., Baldet, P., Bertrand, A., Zoghlami, L.B., Deborde, C., Moing, A., Brouquisse, R., Chaïbi, W., Gallusci, P., 2010. Effects of long-term cadmium exposure on growth and metabolomic profile of tomato plants. Ecotoxicology and Environmental Safety 73, 1965-1974.
- 23. Hegedüs, A., Erdei, S., Horváth, G., 2001. Comparative studies of H<sub>2</sub>O<sub>2</sub> detoxifying enzymes in green and greening barley seedlings under cadmium stress. Plant Science 160, 1085–1093.
- 24. Huang, D., Ou, B. and Prior R.L., 2005. The chemistry behind the antioxidant capacity assays. Journal of Agricultural and Food Chemistry 53, 1841-1856.
- 25. Irtelli, B. and Navari-Izzo, F., 2006. Influence of sodium nitrilotriacetate (NTA) and citric acid on phenolic and organic acids in *Brassica juncea* grown in excess of cadmium. Chemosphere 65, 1348-1354.
- 26. Kanski, J., Aksenova, M., Stoyanova, A., Butterfield, D. A., 2002. Ferulic acid antioxidant protection against hydroxyl and peroxyl radical oxidation in synaptosomal

- and neuronal cell culture systems *in vitro*: structure–activity studies. Journal of Nutritional Biochemistry 13, 273–281.
- 27. Kovácik, J. and Klejdus, B., 2008. Dynamics of phenolic acids and lignin accumulation in metal-treated Matricaria chamomilla roots. Biotic and abiotic stress 27, 605-615.
- 28. Liu, X.L., Zhang, S.Z., Shan, X.Q., Christie, P., 2007. Combined toxicity of cadmium and arsenate to wheat seedlings and plant uptake and antioxidative enzyme responses to cadmium and arsenate co-contamination. Ecotoxicology and Environmental Safety 68, 305–313.
- 29. Liu, Z., Chen, W., He, X., 2011. Cadmium-induced changes in growth and antioxidative mechanisms of a medicine plant (*Lonicera japonica* Thunb.). Journal of Medicinal Plants Research 5, 1411-1417.
- 30. Lloyd, G. and McCown, B., 1980. Combined Proceedings of the International Plant Propagators Society 30, 421-427.
- 31. Lux, A., Martinka1, M., Vaculík, M., White, P., 2011. Root responses to cadmium in therhizosphere: a review. Journal of Experimental Botany 62, 21–37.
- 32. Metwally, A., Safronova, V.I., Belimov, A.A., Dietz, K-J., 2005. Genotypic variation of the response to cadmium toxicity in *Pisum sativum* L. Journal of Experimental Botany 56, 167–178.
- 33. Milone, M.T., Sgherri, C., Clijsters, H., Navari-Izzo, F., 2003. Antioxidative responses of wheat treated with realistic concentration of cadmium. Environmetal and Experimental Botany 50, 265–276.

- 34. Molina, M., Aburto, F., Calderón, R., Escudey, M., Cazanga, M., 2009. Trace element composition of selected fertilizers used in Chile with special regard to phosphorus fertilizers. Soil and Sediment Contamination 18, 497-511.
- 35. Molina, M., Manquian-Cerda, K., Escudey, M., 2010. Sorption and selectivity sequences of Cd, Cu, Ni, Pb, and Zn in single- and multi-component systems in a cultivated Chilean mollisol. Soils and Sediments Contamination 19, 405-418.
- 36. Molina, M., Escudey, M., Chang, A. C., Chen, W., Arancibia-Miranda, N., 2013. Trace element uptake dynamics for maize (*Zea mays* L.) grown under field conditions. Plant and Soil 370, 471-483.
- 37. Mourato, M., Reis, R. and Martins, L.L., 2012. Characterization of plant antioxidative system in response to abiotic stresses: a focus on heavy metal toxicity, in: G. Monta-naro, B. Dichio (Eds.), Advances in Selected Plant Physiology Aspects, Intech, Rijeka, pp. 23–44.
- 38. Panitlertumpai, N., Nakbanpote, W., Sangdee, A., Thumanu, K., Nakai, I., Hokura, A., 2013. Zinc and/or cadmium accumulation in *Gynura pseudochina* (L.) DC. studied *in vitro* and the effect on crude protein. Journal of Molecular Structure 1036, 279-291.
- 39. Parker, D.R., Norvell, W.A., Chaney R.L., 1995. GEOCHEM-PC: A chemical speciation program for IBM and compatible personal computers. p. 253-269. In R. H. Loeppert et al. (ed.) Chemical equilibrium and reaction models. SSSA Spec. Publ. 42, ASA and SSSA, Madison, WI.

- 40. Polge, C., Jaquinod, M., Holzer, F., Bourguignon, J., Walling, L., Brouquisse, R., 2009. Evidence for the existence in Arabidopsis thaliana of the proteasome proteolytic pathway-Activation in response to cadmium. Journal of Biology Chemistry 284, 35412-35424.
- 41. Prior, R.L., Lazarus, S.A., Cao, G.H., Muccitelli, H., Hammerstone, J.F., 2001. Identification of procyanidins and anthocyanins in blueberries and cranberries (*Vaccinium* spp.) using high-performance liquid chromatography/mass spectrometry. Journal of Agricultural and Food Chemistry 49, 1270-1276.
- 42. Rascio, N., and Navari-Izzo, F., 2011. Heavy metal hyperaccumulating plants: How and why do they do it? And what makes them so interesting? Plant Science 180, 169-181.
- 43. Romani, N., Ebner, S., Tripp, C.H., Flacher, V., Koch, F., Stoitzner, P., 2006. Epidermal Langerhans cells—changing views on their function in vivo. Immunology Letter 106,119-125.
- 44. Sanita di Toppi, L., Gabbrielli, R., 1999. Response to cadmium in higher plants. Environmental and Experimental Botany 41, 105–130.
- 45. Serrano, A.B., Font, G., Mañes, J., Ferrer, E., 2013. Comparative assessment of three extraction procedures for determination of emerging *Fusarium* mycotoxins in pasta by LC-MS/MS. Food Control 32,105-114.
- 46. Singleton, V.L. and Rossi, J.A., 1965. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. American Journal of Enology Viticulture 16, 144-58.

- 47. Saidi, I., Ayouni, M., Dhieb, A., Chtourou, Y., Chaibi, W., Djebali, W., 2013.

  Oxidative Damage induced by short-term exposure to cadmium in bean plants:

  Protective role of salicylic acid. South African Journal of Botany 85, 32-38.
- 48. Soong, Y. Y., Barlow, P. J., 2005. Isolation and structure elucidation of phenolic compounds from longan (*Dimocarpus longan* L) seed by high performance liquid chromatography-electrospray ionization mass spectrometry. Journal of Chromatography A, 1085, 270-277.
- 49. WHO, 2011. Cadmium in drinking-water. Background document for preparation of WHO Guidelines for drinking-water quality. Geneva, World Health Organization (WHO/SDE/WSH/03.04/80/Rev/1).
- 50. Wu, G., Kanga, H., Zhangc, X., Shaob, H., Chuc, L., Ruand, Ch., 2010. A critical review on the bio-removal of hazardous heavy metals from contaminated soils: Issues, progress, eco-environmental concerns and opportunities. Journal of Hazardous Materials 174, 1-8.
- 51. You, Q., Wang, B.W., Chen, F., Huang, Z.L., Wang, X., Luo P.G., 2011. Comparison of anthocyanins and phenolics in organically and conventionally grown blueberries in selected cultivars. Food Chemistry 125, 201–208.

# **Chapter 4**

Oxidative stress and production of compounds of phenolic origin in blueberry plantlets cultivated in vitro under a combined cadmium and aluminum treatment

<sup>†</sup>The chapter will be subbmited to Environmental Experimental Botany (ISI).

#### **ABSTRACT**

Blueberries are a widely spread crop around the world, which develops in acid soil where free Al (Al<sup>3+</sup>) can be found at phytotoxic levels. Their production requires constant application of fertilizers that have significant concentrations of cadmium (Cd), an element which under those pH conditions can be very highly bioavailable to the blueberries. In the present study the effect of Al and Cd combined at different concentrations on oxidative stress, antioxidant capacity, and concentration of phenolic compounds in blueberry seedlings cultivated in vitro during four weeks was evaluated. The results showed that the application of binary (Al + Cd) systems caused increased oxidative stress, quantified by the accumulation of MDA, with the greatest damage seen at three weeks of exposure. In the same time period an increase of the antioxidant capacity of blueberries was seen, the same as of the total concentration of phenolic compounds. The results of the HPLC study of the extracts showed increased chlorogenic and elagic acid content with all the treatments applied, suggesting that these phenolic compounds play an important role in the antioxidant response of blueberries. Our results suggest that the addition of metals to the growth medium produce changes of the profile and concentration of phenolic compounds, which play an important rolein the response of blueberries to stress by metals.

#### Introduction

Soil is the most important factor in the plant's growth and development, depending on its origin and physical, chemical and biological characteristics, and it conditions the availability and concentration of nutrients as well as contaminants, largely governing the nutrient flow in the plant.(Yamamoto et al., 2002; Boscolo et al., 2003; Corrales et al., 2008).

Soils of acid origin are found throughout the world, and it is estimated that 40% of the world's soils are used for agriculture and 12% of these have pH below 5.5. The soils in the center south zone of Chile are of volcanic origin, and they are characterized by having a high capacity to retain chemical elements of agricultural and environmental importance (nutrients, trace elements, pesticides). These kinds of soils have pH below 6.0, in addition to a low concentration of exchange bases (Ca<sup>2+</sup>, Mg<sup>2+</sup>, K<sup>+</sup>, Na<sup>+</sup>).

Soil acidity is accelerated by the abundant rainfall in the zone and the use of acidic fertilizers (urea, ammonium phosphate), with the consequent high activity of aluminum, which can reach phytotoxic levels.

Toxicity by Al<sup>3+</sup> limits the productivity of crops in acid soil, and it has been found that micromolar concentrations in the soil solution can rapidly inhibit the elongation of the root and later the absorption of water and nutrients (von Uexküll and Mutert, 1995; Giannakoula *et al.*, 2010). Research has shown that the stress generated by Al<sup>3+</sup> increases the production of reactive oxygen species (ROS) that affect the physiological functioning of the plants and can even induce cell death(Yamamoto *et al.*, 2002; Boscolo *et al.*, 2003; Corrales *et al.*, 2008, Bontempo *et al.*, 2013, Feng *et al.*, 2013). The mechanisms of Al<sup>3+</sup> toxicity in plants has not been completely elucidated, but it is estimated that the Al<sup>3+</sup>induces oxidative stress because of its high affinity for ligands like phosphate and carboxyl groups that have donor oxygen atoms, bonding easily to the phospholipidic membrane and making it more rigid (Devi *et al.*, 2003; Ma *et al.*, 2001; Ryan *et al.*, 2001). There are plants that have developed tolerance mechanisms to Al<sup>3+</sup>, both internally and

externally, allowing them to develop in acid soil (Kochian *et al.*, 2005; Giannakoula *et al.*, 2010; Poschenrieder *et al.*, 2008). Among the tolerance to Al<sup>3+</sup> mechanisms there is exudation of short chain organic acids from the roots of the plants, which is an Al<sup>3+</sup> exclusion mechanism, in the soil solution before it enters the root (Pineros *et al.*, 2008; Liu *et al.*, 2009). However, the production of compounds with a phenolic origin has also been studied trying to help in the tolerance to Al<sup>3+</sup>, stating the hypothesis that phenolic compounds play a role in the detoxification as stable complexing agents of the Al present in the plants (Tolrá *et al.*, 2005).

The application of fertilizers as sources of nitrogen, phosphorus and potassium, among which the most widely used are urea, triple superphosphate (TSP), and mono- and diammonium phosphate,represents also an important source of trace elements like Zn, Cd, Pb, Cr, Cu, and Ni (Chang and Page, 2000; Nicholson *et al.*, 2003; Molina *et al.*, 2009) Among the elements that affect soil and water, cadmium (Cd) is often mentioned because of its high toxicity and its mobility in the soil/plant system (Sanita and Gabbrielli, 1999). Industrial activities, and agriculture through the application of phosphate fertilizers and biosolids, have enriched the soil with this element (Zheljazkov *et al.*, 2005).

Exposure of plants to cadmium induces various symptoms of phytotoxicity, such as chlorosis, biomass reduction, inhibition of root elongation until cell death occurs (Milone *et al.*, 2003). It has been found that the presence of Cd generates free radicals that harm the plants' tissues, and depending on its concentration and the kind of plant that is exposed to cadmium, it can inhibit or stimulate the activity of various antioxidant enzymes before the toxicity symptoms become visible (Fu and Huang, 2001; Liu *et al.*, 2007). Another effect

of the presence of cadmium is related to perturbations of the capture and distribution of nutrients in the plants (Sandalio *et al.*, 2001).

It has been described that plants have strategies that allow them to control reactive oxygen species. One of these strategies is based on the production of secondary metabolites with antioxidant properties which are synthesized by the plant as a method of defense against biotic stress, but it is possible that they also play a role in the physiological functioning of the plant when it is in the presence of a high concentration of heavy metals (Close and McArthur, 2002).

Studies made with blueberry plantlets cultivated *in vitro* showed higher concentrations of phenolic compounds, mainly chlorogenic acid, when they were subjected to different concentrations of Al<sup>3+</sup> (Manquián *et al.*, 2013). On the other hand, studies made with blueberry plants subjected to the presence of Cd<sup>2+</sup> in the growth medium have shown that this element causes damage by oxidative stress which has been quantified by means of the increased content of malondialdehyde (MDA), which is a by-product of the degradation by oxidation of the polyunsaturated fatty acids in the membranes, and it has also been shown that the presence of Cd causes a change in the profile of phenolic compounds generated by the plant (Ge *et al.*, 2012, Guo *et al.*, 2007, Irtelli and Navari-Izzo, 2006). However, most of the research is centered on the physiological effects of the plants when exposed to heavy metals, and there are not many studies that evaluate the effect of the metals on the production of phenolic compounds and the generated oxidative stress.

Therefore, it is important to carry out a study to evaluate the effects of exposure to both metals when they are present in the growth medium on the production of phenolic compounds and their implication oxidative stress. For that purpose, blueberry plantlets cultivated *in vitro* were exposed to the combined action of Al and Cd to determine the effects of the presence of both metals on the damage by oxidative stress, on the plant's response by means of its antioxidant activity, and on the concentration of phenolic compounds as a consequence of their presence in the growth medium.

#### 2. MATERIALS AND METHODS

### 2.1 Plant material, growth conditions and treatments

To start the in vitro culture, blueberry plantlets were obtained by replication of sterilized leaf segments (2 cm long) from two-month old precultivated plantlets. To each culture flask (6 cm diameter, 10 cm tall, and 200 cm³) was added 25 mL of Woody Plant culture medium (Lloyd and McCown, 1980) which contains the following components (mg·L¹): NH<sub>4</sub>NO<sub>3</sub> 400, CaCl<sub>2</sub>·2H<sub>2</sub>O 96, MgSO<sub>4</sub>·7H<sub>2</sub>O, K<sub>2</sub>SO<sub>4</sub> 990, KH<sub>2</sub>PO<sub>4</sub> 170, Ca(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O 556, H<sub>3</sub>BO<sub>3</sub> 6.2, Na<sub>2</sub>EDTA·2H<sub>2</sub>O 37.2, CuSO<sub>4</sub>·5H<sub>2</sub>O 0.25, FeSO<sub>4</sub>·7H<sub>2</sub>O 27.8, MnSO<sub>4</sub>·4H<sub>2</sub>O 22.3, ZnSO<sub>4</sub>·7H<sub>2</sub>O 8.6, Na<sub>2</sub>MoO<sub>4</sub>·2H<sub>2</sub>O 0.25. The culture medium was supplemented with 2.76 mg·L¹lof hormone 2-iP (2-isopentenyl adenine) purine, using sucrose as a source of carbon (15 g·L¹l) and Phytagel agar (3 g·L¹l) as gelling agent. The pH was adjusted to 5.2. The media were sterilized in an autoclave during 15 minutes at 121 °C. and the culture medium was modified by adding Al and/or Cd, setting the following conditions: (1) Control (original culture medium); (2) 100 μM of total Al (designated as 100Al), (3) Al + Cd (100 μM total Al + 50 μM total Cd, designated as 100Al/50Cd), (4) and Al + Cd (100 μM total Al + 100 μM total Cd, designated as 100Al/100Cd). The

concentration of Cd and Al was adjusted using the corresponding 0.01 M stock solutions of CdCl<sub>2</sub>·2H<sub>2</sub>O and AlCl<sub>3</sub>.

The cultures were kept in culture chambers at  $21 \pm 2$  °C and photoperiods of 16 hours light/8 hours darkness. The sterilization and planting procedures were carried out in a laminar flow chamber under aseptic conditions. Once the *in vitro* culture had become established (control and treatments), a completely random design was used in which the experimental unit included six blueberry plantlets clones (6 plantlets: 1 sample), and every sample was ran in triplicate. *In vitro* cultivated plantlets with more than 30 days of adaptation to cultivation conditions were defined as starting control (time=0 days), and used to define the physiological base line for all the studied parameters.

The computer program GEOCHEM-PC (Parker *et al.*, 1995) was used to calculate the chemical speciation of Cd and Al in the growth medium of the plantlets.

#### 2.2. Extracts preparation.

Fresh materials (0.1g·mL<sup>-1</sup>) are used to prepare the extract using 85% v/v of hydroethanolic solution; the samples were sonicated at 50-60 Hz of frequency during two hours at 25 °C (298 K) according to the method described by Adam (2009).

### 2.3. Antioxidant activity

2.3.1. 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical-scavenger spectrophotometric assay.

The free radicals removal capacity of each extract was evaluated using the radical DPPH technique, described by Brand-Williams *et al.* (1995) with some modifications. Briefly, 20  $\mu$ L of the ethanol extract was added to 980  $\mu$ L of DPPH solution (25 mg·L<sup>-1</sup>), the absorbance was continuously monitored at 517 nm using a UV-visible

spectrophotometer (Agilent 8453 UV-Vis), for 240 seconds (Brand-Williams *et al.*, 1995; Huang *et al.*, 2005). Results were expressed as % of consumed DPPH.

## 2.3.2. Ferric reducing antioxidant power (FRAP).

The FRAP assay measures the ability of the sample to reduce Fe<sup>3+</sup> to Fe<sup>2+</sup> (Benzie and Strain, 1996), followed by the absorbance at 593 nm of the blue complex formed with tripyridyltriazine (TPTZ). FRAP reagent was prepared by mixing acetate buffer (300 mM), TPTZ solution (10 mM in HCl) and FeCl<sub>3</sub> solution (20 mM) at a 10:1:1 ratio. The FRAP reagent was maintained at  $37\pm2^{\circ}$  C ( $310\pm2$  K). Absorbance of samples containing 900  $\mu$ L of FRAP reagent, 80  $\mu$ L of water and 20  $\mu$ L of ethanolic extract,was measured at 593 nm in an Agilent 8453 UV-Vis spectrophotometer. The measures were expressed in ascorbic acid equivalents.

# 2.4. Total phenolic content (TPC)

The total phenolic content of ethanolic extracts was determined based on the method described by Singleton and Rossi (1965). Results were expressed as mg Gallic acid equivalents.

### 2.5 HPLC-DAD analysis of extracts

High performance liquid chromatography with diode array detector (HPLC-DAD) was used to separate and determine the phenolic compounds in the ethanolic extracts of blueberry tissues. The ethanolic extract was passed through a 0.45-µm membrane filter and analyzed by HPLC-DAD. An Agilent HPLC-DAD 1100 series chromatograph equipped with a RP-C18 column at 25 °C, was used. The mobile phase is a gradient of acetonitrile

(A) and 1% phosphoric acid (B), using these program: time=0 minutes 10% of A, 5 minutes 25% of A, 8 minutes 35% of A, 15 minutes 60% of A, 17 minutes 35% of A and finally 20 minutes 10% of A; with an initial pressure of about 120 bar, 1 mL/min of flow and 20  $\mu$ L of injection volume using a Reodyne valve, the register was carried out at 254, 280, 314 and 340 nm.

## 2.6. Lipid peroxidation

The level of lipid peroxidation was determined in terms of MDA concentration according to the method proposed by Heath and Parker (1968) slightly modified. The concentration of MDA was calculated from the difference of the absorbance at 532 and 600 nm using the extinction coefficient of 155 mmol·L<sup>-1</sup> cm<sup>-1</sup> and expressed as nmol·g<sup>-1</sup> FW.

# 2.7. Statistics analysis.

All the data shown in the study correspond to  $X\pm1sd$ . The tests were made three times for the HPLC measurements and six times for the other measurements. The statistical analysis was made by two-way Anova and the comparisons of averages were made by means of the Tukey Multiple Comparison of Means test. To determine the correlation between physiological variables, Pearson's test was performed. Statistical significance was set at p<0.05.

#### 3. RESULTS

# 3.1 Chemical speciation of the Woody-Plant nutrient medium

The availability and mobility of Cd and Al in environmental systems like soil-soil solutions is not only related to total concentration, but to the concentration of the different Cd and Al species present in the solution, forming either inorganic complexes, organic

complexes, and precipitates, or as free species. Using computer programs like GEOCHEM-PC it was possible to determine the chemical forms in which Cd and Al are found in the culture medium.

**Table 1.** Percent distribution of Al and Cd in Lloyd-McCown culture medium for the different conditions studied ("complexed with" means Al and Cd complexed by the respective anion, Parker *et al.*, 1995).

Condition		$\mathbf{S}_{1}$	peciation (%	<b>(o)</b>			
Condition	100Al	100Al 100Al/50Cd			100Al/100Cd		
Species	Al	Al	Cd	Al	Cd		
Free metal	0.66	1.05	8.63	1.26	20.81		
Complexed with							
Sulfate	2.05	3.24	4.19	3.88	10.09		
Borate	0.12	0.19	0	0.22	0		
EDTA	51.5	23.46	86.23	9.06	66.82		
Phosphate	44.8	70.83	0.07	84.09	0.17		
Cloride	0.00	0.00	0.79	0.00	1.9		
Nitrate	0.00	0.00	0.08	0.00	0.2		
OH (Hydroxides)	0.79	1.24	0.00	1.49	0.00		

According to the chemical speciation obtained from GEOCHEM-PC (Table 1), it was found that free Al<sup>3+</sup>increases with the addition of Cd to the culture medium. The Cd is found in greater proportion as free Cd<sup>2+</sup> in both binary treatments (8.63% for 100Al/50Cd and 20.81% for 100Al/100Cd), with a lower free Al<sup>3+</sup>proportion in solution (<1.5% of the total in both binary treatments).

In Table 1 it is seen that Al in the absence of Cd is found mainly forming complexes with EDTA and to a smaller degree with phosphate. In the presence of cadmium, Al is

found mostly complexed by  $PO_4^{3-}$  (70-90%), while Cd is forming complexes mostly with EDTA (> 60%).

Table 2 shows the free concentration and the activity of each metal in solution. For both Cd and Al the GEOCHEM-PC program showed that the free concentration as well as the activity in solution increased in the binary system.

**Table 2.** Molar concentration and activity of Al and Cd in Lloyd-McCown culture medium for the different treatments applied.

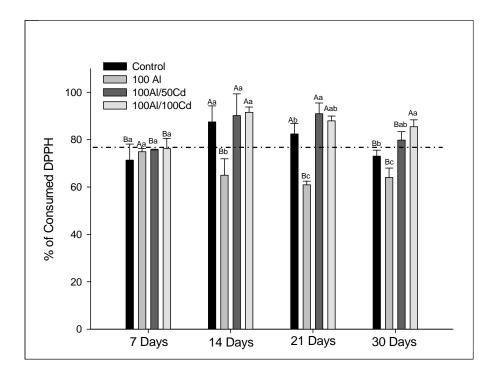
Tratamiento	Specie	Concentration (M)	Activity (M)
100Al	Free Al <sup>3+</sup>	$5.01 \times 10^{-7}$	1.48x10 <sup>-7</sup>
50Cd	Free Cd <sup>2+</sup>	$1.17 \times 10^{-6}$	$5.90 \times 10^{-7}$
100Cd	Free Cd <sup>2+</sup>	$1.69 \times 10^{-5}$	$8.48 \times 10^{-6}$
100Al/50Cd	Free Al <sup>3+</sup>	$1.04 \times 10^{-6}$	$2.23 \times 10^{-7}$
100AI/30Ca	Free Cd <sup>2+</sup>	$4.32 \times 10^{-6}$	$2.18 \times 10^{-6}$
100 41/1000 4	Free Al <sup>3+</sup>	$1.26 \times 10^{-6}$	$2.68 \times 10^{-7}$
100Al/100Cd	Free Cd <sup>2+</sup>	$2.08 \times 10^{-5}$	$1.05 \times 10^{-5}$

## 3.2 Antioxidant activity

The antioxidant activity of the ethanol extracts of blueberries was measured by means of the diphenylpicrylhydrazyl (DPPH) and ferric reducing antioxidant power (FRAP) assays, which evaluate the capacity of the extract for trapping the DPPH free radical and for reducing Fe(III) to Fe(II), respectively.

The results obtained from the DPPH showed that the response to the application of the binary Al + Cd system (in its different concentrations) in the growth solution did not affect significantly the antioxidant capacity of blueberries in the first three weeks of

treatment with respect to the control (Fig. 1), but after 30 days of exposure significant differences were seen in the DPPH values between the control and the Al + Cd treatments applied. However, a significant decrease of the DPPH values was seen in the blueberry seedlings treated only with aluminum (100Al) compared to the other treatments and at different times during the study (Fig. 1)

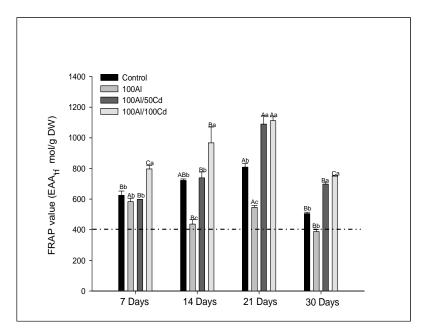


**Fig. 1.** DPPH scavenging, the result is expressed in % of consumed DPPH (mean  $\pm$  standard deviation n= 6). A different lower case letters indicate statistically significant differences (Tukey, P < 0.05) between same day and different treatment. A different upper case letters indicate differences (Tukey, P  $\leq$  0.05) between different day and same treatment. The segmented line shows the % of DPPH consumed for the t=0 days control.

The measurement of the reducing capacity of the ethanol extracts was made with the FRAP test (Fig. 2). The data obtained from this assay show a trend similar to that obtained from the DPPH test. The 100Al/50Cd treatment did not show significant differences with respect to the control after the first 7 and 14 days de exposure, while the 100Al/100Cd

treatment showed a significant increase of the FRAP value during the whole study (p < 0.05) with respect to the values obtained for the control and 100Al. Similar results have been reported for plantlets exposed to only 100  $\mu$ M Cd, in which the FRAP value increased at 7, 14 and 21 days (Manquián et al., 2013).

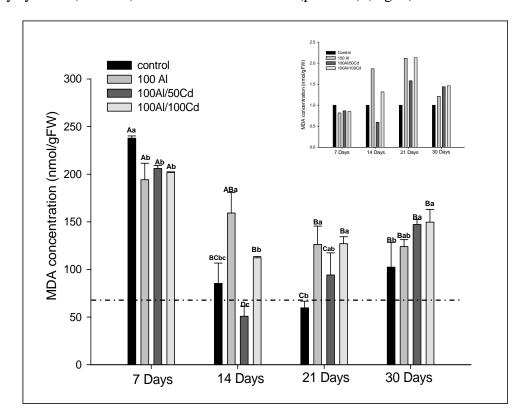
In the case of the 100Al treatment the FRAP values were smaller over the whole exposure time compared to the Al + Cd systems, and this is consistent with what was seen with the DPPH assay (Fig. 1).



**Fig. 2**. Ferric reducing/Antioxidant power (FRAP) the results as expressed in ascorbic acid equivalents (mean  $\pm$  standard deviation, n= 6). A different lower case letters indicate statistically significant differences (Tukey, P < 0.05) between same day and different treatment. A different upper case letters indicate differences (Tukey, P  $\leq$  0.05) between different day and same treatment. The segmented line shows the FRAP value for the t=0 days control.

# 3.3 Cd-induced changes in malondialdehyde (MDA) content

The effects of oxidative stress due to the presence of heavy metals can be quantified from the content of MDA, which is a by-product of the degradation of polyunsaturated fatty acids in the membranes (Fig. 3). In the first week of exposure the highest MDA content was found in the control, while no significant differences in the MDA content between the binary systems (Al + Cd) and the 100Al treatment (p < 0.05) (Fig. 3).



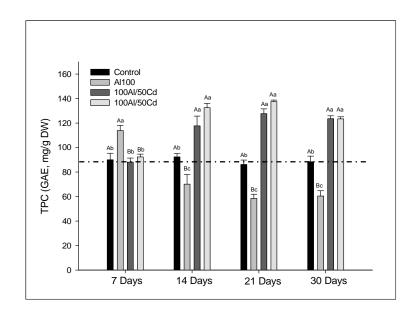
**Fig. 3.** Effects of the different stress treatments on MDA contents (mean  $\pm$  standard deviation, n= 6). A different lower case letter indicate statistically significant differences (Tukey, P < 0.05) between same day and different treatment. A different upper case letters indicate differences (Tukey, P  $\leq$  0.05) between different day and same treatment. Inset: Normalized data with respect to control (MDA<sub>sample</sub>/MDA<sub>control</sub>). The segmented line shows the MDA level for the t=0 days control.

This behavior would account for the degree of stress generated by transferring the plantlets to the *in vitro* culture medium (high damage in the control) and the kind of response of the plant as a defense mechanism when it is under stress by metals, finding that in those culture media where there are heavy metals (Al + Cd and 100Al) the blueberries increase their antioxidant defense system in order to reduce the damage from the presence of the metals, but the plants are unable to respond quickly to the stress generated by the culture medium change. At 14 days of treatment a decrease of the MDA content was seen in both binary systems, with the lowest point obtained with the 100Al/50Cd treatment (p < 0.05), while the largest MDA values were seen with the 100Al treatment. In the last two weeks of the study (21 and 30 days), the damage by stress was significantly greater for all the treatments with respect to the control (p < 0.05).

# 3.4 Total phenolic content (TPC) and HPLC-DAD phenolic compounds identification.

Total phenolic compound content is shown in Fig. 4. Compared to the control, during the first seven days a significant increase of TPC is seen in the 100Al treatment, which decreases with increasing exposure time (14, 21 and 30 days) (p < 0.005).

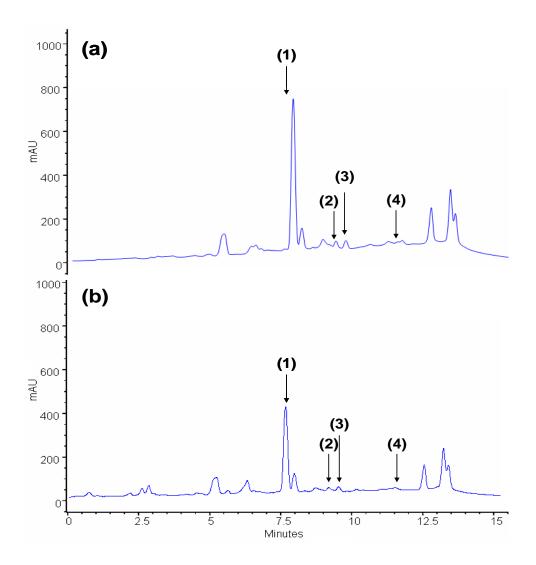
No significant changes are seen with both binary Al + Cd treatments during the first 7 days of exposure. In the following 14, 21 and 30 days of the study a significant increase of the TPC is seen for the binary treatments compared to the control and 100Al treatments.



**Fig. 4**. Effect of different stress treatments on the total content of phenolic (TPC) compounds (mean  $\pm$  standard deviation). A different lower case letter indicates statistically significant differences (Tukey, P < 0.05) between same day and different treatment. A different upper case letters indicate differences (Tukey, P  $\leq$  0.05) between different day and same treatment. The segmented line shows the TPC for the t=0 days control.

The HPLC analysis of the ethanol extracts showed that the predominant compounds in the controls are chlorogenic and elagic acid, compounds that have been identified in previous studies in blueberry fruits at different ripening stages (Rivera *et al.*, 2011). It was also possible to identify compounds such as rutin and quercetin, which are found in significantly lower concentrations compared to the two most important ones (Fig. 5).

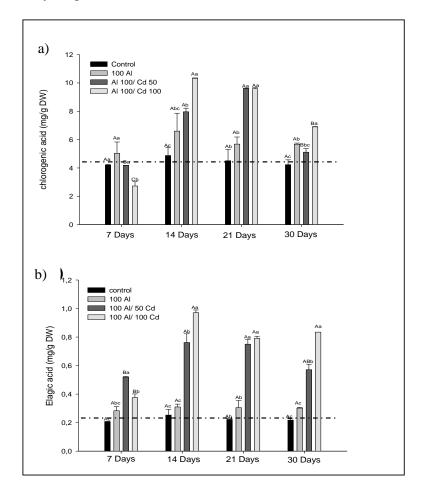
Fig. 5 shows the HPLC chromatograms of the blueberry ethanol extracts. In the seedlings subjected to the two Al + Cd treatments it was seen that the areas corresponding to chlorogenic and elagic acid increase with increasing Cd concentration in each binary combination.



**Fig. 5**. Chromatogram of blueberry extracts of 14 days measured at 314 nm, for the treatment 100Al/100Cd (a) and control (b). The numbers correspond to identified phenolic compounds (1) chlorogenic acid, (2) Rutin, (3) ellagic acid and (4) Quercetin.

The Fig. 6 shows the concentration of chlorogenic and elagic acids obtained with the applied treatments. It is seen that no significant changes occurred in the control over the whole study period (p < 0.05). At 14 days of exposure the chlorogenic acid concentration increased with the binary treatments and a significant increase only with the 100Al/100Cd treatment compared with 100Al. At 21 days there was a significant increase in the

chlorogenic acid concentration with the binary (Al + Cd) mixture. It should be stressed that between 14 and 21 days of exposure there were higher chlorogenic acid concentrations with all the treatments except with the control. For the 100Al/50Cd and 100Al/100Cd conditions an important decrease of the chlorogenic acid concentration (p < 0.005) was seen during the fourth week of the study (Fig.6a).



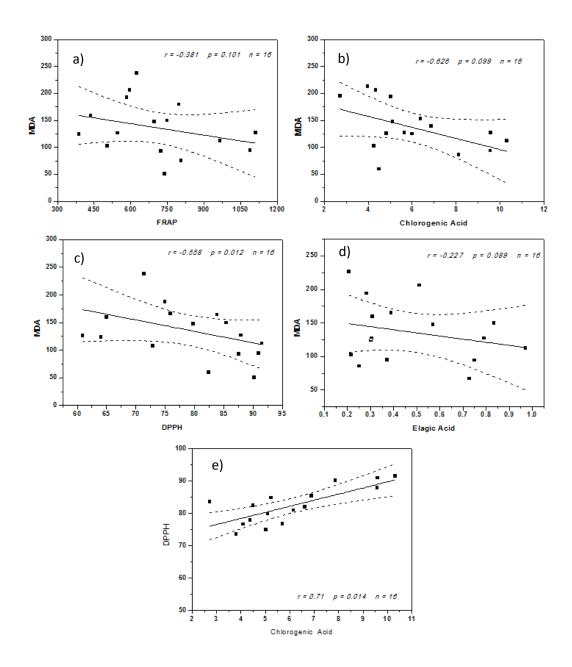
**Fig. 6.** Variation of (a) chlorogenic and (b) elagic acid content (mean  $\pm$  standard deviation) as a function of time and the concentration of the different treatments in blueberries (thegraphs have different scales. A different lower case letter indicates statistically significant differences (Tukey, P < 0.05) between same day and different treatment. A different upper case letter indicates differences (Tukey, P  $\leq$  0.05) between different day and same treatment. The segmented line shows the elagic and chlorogenic acids concentration for the t=0 days control.

Similar results were seen in the elagic acid concentration over time, with the peak concentration occurring between 14 and 21 days of exposure to the binary treatments (p <0.005). The differences with respect to the control and the 100Al treatment remained until the end of the study (Fig. 6b).

## 3.5 Correlation between the antioxidant response parameters and MDA content

The combined presence of Al and Cd causes oxidative stress and changes in the profile of phenolic compounds biosynthesized by blueberries; this is shown by the correlation between the MDA, DPPH and chlorogenic acid content during exposure of the plantlets to the different conditions considered (Fig. 7). From these results it can be suggested that MDA content decreases when the capacity to capture free radicals (expressed as a higher DPPH content) increases (Fig. 7c, r = -0.56, p < 0.05). A significant negative correlation (Fig. 8b, r = -0.626, p = 0.099) between lipid peroxidation and chlorogenic acid content indicates that the latter plays a critical role in the decrease of MDA, which is justified by the high correlation (Fig. 7e, r = 0.70, p < 0.05) found between the chlorogenic acid content and DPPH. Similar results were reported by Manquián et al., for a study made with blueberry plantlets subjected to different concentrations of Al (Manquián et al., 2013).

The reducing power of the extracts determined by means of FRAP essays and by the elagic acid content did not correlate significantly with MDA (Fig.7a, d), indicating that this protection mechanism is not very effective to reduce the lipid peroxidation undergone by blueberries under the studied conditions.



**Fig. 7**. Relations between lipid peroxidation (MDA) and the antioxidant response measured by FRAP, DPPH, Chlorogenic acid, and ellagic acid addition ratio DPPH and chlorogenic acid. Pearson's coefficients are indicated.

#### 4.-DISCUSSION

Aluminum is one of the most limiting factors of cultivation in acid soils, and furthermore the use of phosphate fertilizers, particularly thoseof natural origin like triple superphosphate (TSP),increase the concentration of trace elements (Cd, Cu, Ni, etc.) in the soil. The acidity of these soils favors the mobility of the metals in the soil solution, which results ingreater activityof metals like Al and Cd. While at the usual pH of these acid soils (4.5-5.5) 29-15% of the total Al and more than 99.8% of the total Cd are found in solution (as free species or forming some soluble complex), in a neutral or alkaline soil (pH 7.0-8.0) more than 98% of the total Al and Cd are found as precipitates. Therefore, in view of this greater availability, the presence of trace elements in soil derived from volcanic materials will have a greater impactonthe behavior of plants compared to what can happen in neutral or alkaline soils.

The increased concentration of trace elements or heavy metals in the growth solution has as a consequence a saturation of the complexing capacity of the medium, and this results in increase of the free concentration and the total activity of the trace elements, in turn increasing the availability of these elements to the plant. An equivalent situation can occur in real terms, with an excessive increase of the total concentration of trace elements in the soil solution, with the result of the inability of the plant to generate short chain organic acids which can complex elements, resulting in increased availability that can finally reach phytotoxic levels.

The application of metals to the culture medium produced effects in the antioxidant response of the plant, showing an increase of the antioxidant capacity of the blueberry extracts treated with the binary combination.

Similar results were found in a study in which only the effectof Cd on the antioxidant activity of blueberry extracts was determined (Manquián *et al.*, 2013). These results suggest that the joint presence of both metals in the culture medium produces a greater response of the blueberry non-enzymatic antioxidant mechanisms (FRAP and DPPH).

The presence of trace elements like Al and Cd can induce oxidative stress, whose effects are determined by means of the analysis of lipid peroxidation (MDA). The increased MDA content in the first seven days, for all the treatments, including the control, is a consequence of the stress to which the plant is subjected at the time of being transplanted to the *in vitro* medium, but no differences are seen between the treatments and the control. However, once the plantlets have become climatized, after 14 days, a significant decrease in MDA content was seen in the binary treatments. At 21 and 30 days of exposure to the binary treatments no significant differences were seen in the accumulation of MDA with respect to the 100Al treatment, but they did appear with respect to the control, where the presence of the metals causes increased lipid peroxidation. These results may indicate that there was no synergism in the damage produced by the presence of both metals in solution, as could have been expected, because through chemical speciation there was increased activity of the metals in solution in the binary composition for both treatments. But the results of the MDA content in the combined treatments can be reflected in an increased FRAP value in the combined treatments and in the values of the concentration of phenolic compounds, which increases at days 14 and 21 of combined treatment. Tolerance to abiotic stress is often related to increased antioxidant activity, so our study would be indicating that aluminum tolerant plants would increase their antioxidant components to counteract

the damage by the metal (Fig. 7). The induction of phenolic compounds due to the presence of metals in the growth medium has been observed in different studies (Michalak *et al.*, 2006). In the phenolic compounds found in blueberry seedlings, chlorogenic and elagic acids were quantified, both of them well known for their antioxidant power (Niggeweg *et al.*, 2004). The results showed similarities between antioxidant capacity and phenolic compounds, whose concentration increased after 14 days. The concentration of chlorogenic acid was stimulated at 14 days of treatment mainly in the 100Al/100Cd binary mixture, while at 21 days both combinations, 100Al/50Cd, and 100Al/100Cd, increased the concentration of chlorogenic acid. These results agree with those obtained for MDA and decreased damage by lipid peroxidation at 14 and 21 days, showing that chlorogenic acid has a protective role in the damage caused by lipid peroxidation (Niggeweg *et al.*, 2004).

The quantification of chlorogenic acid in the presence of metals has been studied by various authors (Kováčic, 2009; Irtelli and Navari-Izzo, 2006; Manquián *et al.*, 2013), and the trend has always beenincreased chlorogenic acid concentration in the presence of the metal in the growth medium.

Elagic acid is a polyphenol found in berries like strawberries, raspberries, blackberries and blueberries (Jiménez-García et al., 2013; Da Silva et al., 2008, Rivera et al., 2010). In strawberries it has been found that elagic acid content represents 51% of the total acids of phenolic origin (Aiyer et al., 2008). In our study we found a significant increase of elagic acid content after 14 days with the binary treatments, and yet this amount is significantly low compared to the chlorogenic acid content. Similar results have been obtained in studies on phenolic compounds, where the main compound found in blueberries

is chlorogenic acid and to a smaller extent other compounds like elagic acid (Ribera *et al.*, 2010; Može *et al.*, 2011).

The binary study also allowed us to detec to ther compounds that were only identified but not quantified, mainly rutin and quercetin. Rivera *et al.*, (2010), determined the presence of rutin and quercetin in blueberries at different ripening stages. Studies made by Kovacik *et al.*, (2010), on chamomile plants exposed to 60 and 120 µM Al showed increased content of different phenolic compounds. Our results show that there is an variation in the profile of the chromatograms, indicating that there is an change of these compounds when the plantlet grows in the presence Al + Cd, indicating that phenolic compounds may be affected by the amount and kind of metal to which the plants are exposed during growth.

#### 5.- CONCLUSION

In general, the results of this study showed the stress-causing effect of trace elements in the growth medium, which was reflected in increased damage by lipid peroxidation, finding that the greatest oxidative damage occurs after 14 days of exposure to both treatments. The antioxidant response increased in the binary treatments, where the largest responses were seen after 14 days, and this increase is related to an increase in the content of quantified phenolic compounds, with chlorogenic acid as the main component, concluding that antioxidant activity plays an important role as a defense mechanism against damage by trace elements.

In conclusion, the Al + Cd mixture induces changes in the production of phenolic compounds, evidenced by an increase of their proportion compared to those in the control, indicating that it would actually be possible to modulate plantlets to favor the production of

compounds of interest, in this case phenolic compounds, but the impact on the composition and incorporation of trace elements at later growth stages, when the leaves of the plants can be used as infusions, must be evaluated.

### Acknowledgements

Financial support of this work by the Center for the Development of Nanoscience and Nanotechnolgy (CEDENNA). K. Manquián-Cerda acknowledges a scholarship from UFRO (Chile) and DICYT-DGT (Universidad de Santiago de Chile, GE. Zúñiga).

#### REFERENCES

- Aiyer, H.S., Srinivasan, C., Gupta, R.C., 2008. Dietary berries and ellagic acid diminish estrogen-mediated mammary tumorigenesis in ACI rats. Nutrición and Cancer 60, 227-234.
- 2. Benzie, I. F. F. and Strain, J.J., 1996. The Ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power" the FRAP assay. Analytical Biochemistry 239, 70-76.
- 3. Boscolo, P., Menossi, M. and Jorgea, R., 2003. Aluminum-induced oxidative stress in maize. Phytochemistry 62, 181–189.
- Botempo, P., Carafa, V., Grassi, R., Basile, A., Tenore, G.C., Formisano, C., Rigano,
   D. and Altucci, L., 2013. Antioxidant, antimicrobial and anti-proliferative activities of
   Solanumtuberosum L. var. Vitelotte. Food and Chemical Toxicology 55, 304-312.
- 5. Brand-Williams, W., Cuvelier, M. E., Berset, C., 1995. Use of free radical method to evaluate antioxidant activity. Lebensmittel Wissenschaft und Technology 28, 25–30.

- 6. Chang, A.C., Page, A.L., 2000. Trace element slowly accumulating, depleting in soils. California Agriculture 54, 49-55.
- 7. Close, D. C., McArthur, C., 2002. Rethinking the role of many plant phenolics Protection from photodamage not herbivores? Oikos 99, 166–172.
- 8. Corrales, I., Poschenrieder, C., Barcelo J., 2008. Boron-induced amelioration of aluminium toxicity in a monocot and a dicot species. Journal of Plant Physiology 165, 504–513.
- 9. Da Silva, M., Lajolo, F.M., Genovese, M.I., 2008. Bioactive compounds and quantification of total ellagic acid in strawberries (*Fragaria x ananassa* Duch.). Food Chemistry 107, 1629-1635.
- 10. Dragović-Uzelac, V., Savić, Z., Brala, A., Levaj, B., Kovaĉević, D.B., Biŝko, A., 2010.
  Evaluation of phenolic content and antioxidant capacity of blueberry cultivars
  (Vaccinium corymbosum L.) grownin the northwest Croatia. Food Technology and Biotechnology 48, 214–221.
- 11. Feng, R., Wei, C., Tu, S., 2013. The roles of selenium in protecting plants against abiotic stresses. Environmental and Experimental Botany 87, 58–68.
- 12. Fu, J., Huang, B., 2001. Involvement of antioxidants and lipid peroxidation in the adaptation of two cool-season grasses to localized drought stress. Environmental and Experimental Botany 45, 105–114.
- 13. Ge, W., Jiao, Y.Q., Sun, B.L., Qin, R., Jiang, W.S., Liu, D.H., 2012. Cadmium-mediated oxidative stress and ultrastructural changes in root cells of poplar cultivars. South African Journal of Botany 83, 98–108.

- 14. Giannakoula, A., Moustakas, M., Syros, T., and Yupsanis, T., 2010. Aluminum stress induces up-regulation of an efficient antioxidant system in the Al-tolerant maize line but not in the Al-sensitive line. Environmental and Experimental Botany 67, 487-494.
- 15. Guo, T. R., Zhang, G.P.and Zhang, Y.H., 2007. Physiological change in barley plants under combined toxicity of aluminum, copper and cadmium. Colloids and Surface B: Biointerfaces 57, 182-188.
- 16. Heath, R. and Packer, L., 1968. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. Archives of Biochemistry and Biophysics 125, 189-198.
- 17. Huang, D., Ou, B. and Prior, R.L., 2005. The chemistry behind antioxidant capacity assay. Journal of Agricultural and Food Chemistry 53, 1841-1856
- 18. Irtelli, B., and Navari-Izzo, F., 2006. Influence of sodium nitrilotriacetate (NTA) and citric acid on phenolic and organic acids in *Brassica juncea* grown in excess of cadmium. Chemosphere 65, 1348-1354.
- 19. Jimenez-García, S., Guevara-Gonzalez, R., Miranda-Lopez, R., Feregrino-Perez, A., Torres-Pacheco, I., Vasquez-Cruz, M., 2013. Functional propierties and quality characteristics of bioactive compounds in berries: Biochemistry, biotechnology and genomics. Food Research International 54, 1195-1207.
- 20. Jung, M.J., Heo, S., Wang, M.H., 2008. Free radical scavenging and total phenolic contents from methanolic extracts of *Ulmus davidiana*. Food Chemistry 108, 482-487.
- 21. Kováčik, J., Klejdus, B., Bačkor, M., 2009. Phenolic metabolism of Matricaria chamomilla plants exposed to nickel. Journal of plant physiology 166, 1460-1464.

- 22. Kováčik, J., Klejdus, B., Hedbavnyb, J., 2010. Effect of aluminium uptake on physiology, phenols and amino acids in Matricaria chamomilla plants. Journal of Hazardous Materials 178, 949–955
- 23. Liu, X.L., Zhang, S.Z., Shan, X.Q., Christie, P., 2007. Combined toxicity of cadmium and arsenate to wheat seedlings and plant uptake and antioxidative enzyme responses to cadmium and arsenate co-contamination. Ecotoxicology and Environmental Safety 68, 305–313.
- 24. Lloyd, G., McCown, B., 1980. Commercially feasible micropropagation of mountain Laurel, *Kalmialatifolia*, by use of shoot tip culture. Combined Proceedings of the International Plant Propagators' Society 30, 421-427.
- 25. Milone, M.T., Sgherri, C., Clijsters, H., Navari-Izzo, F., 2003. Antioxidative responses of wheat treated with realistic concentration of cadmium. Environmenal and Experimental Botany 50, 265–276.
- 26. Molina, M.A., Aburto, F.A., Calderón, R.A., Cazanga, M., and Escudey, M., 2009.
  Trace Element Composition of Selected Fertilizers Used in Chile with Special Regard to Phosphorus Fertilizers. Soil and Sediments Contamination 18, 497-511.
- 27. Molina, M. Manquian-Cerda, K. Escudey. M., 2010. Sorption and selectivity sequences of Cd, Cu, Ni, Pb, and Zn in single- and multi-component systems in a cultivated Chilean mollisol. Soil and Sediments Contamination 19, 405-418.
- 28. Može, S., Polak, T., Gasperlin, L., Koron, D., Vanzo, A., Ulrih, N.P., Abram, V., 2011.
  Phenolics in slovenian bilberries (*Vaccinium myrtillus* L.) and blueberries (*Vaccinium corynbosum* L.). Journal of agricultural and food chemistry 59, 6998-7004.

- 29. Nicholson, F.A., Smith, S., Alloway, B. J., Carlton-Smith, C., and Chambers, B.J., 2003. An inventory of heavy metals inputs to agricultural soils in England and Wales. The Science of the Total Environment 311, 205–219.
- 30. Niggeweg, R., Michael, A.J., Martin, C., 2004. Engineering plants with increased levels of the antioxidant chlorogenic acid. Nature Biothecnology 22, 746-754.
- 31. Poschenrieder, C., Gunse, B., Corrales, I. and Barcelo, J., 2008. A glance into aluminum toxicity and resistance in plants. Science Total Environmental 400, 356–368.
- 32. Parker, D. R., Norvell, W. A., Chaney, R. L., 1995. GEOCHEM-PC: A chemical speciation program for IBM and compatible personal computers. p. 253-269. In R. H. Loeppert et al. (ed.) Chemical equilibrium and reaction models. SSSA Spec. Publ. 42, ASA and SSSA, Madison, WI.
- 33. Prasad K. N., Yang B., Yang S., Chen Y., Zhao M., Ashraf M., Jiang Y., 2009. Identification of phenolic compounds and appraisal of antioxidant and antityrosinase activities from litchi (*Litchi sinensis Sonn.*) seeds. Food Chemistry 116, 1–7.
- 34. Ribera, A.E., Reyes-Díaz, M., Alberdi, M., Zuñiga, G.E., Mora, M.L., 2010. Antioxidant compounds in skin and pulp of fruits change among genotypes and maturity stages in highbush blueberry (*Vaccinium corymbosum* L.) grown in southern Chile. Journal soil science and plant nutrition 10, 509-536.
- 35. Sakihama, Y., Cohen, M.F., Grace, S.F., Yamasaki H., 2002. Plant phenolic antioxidant and prooxidant activities: phenolics-induced oxidative damage mediated by metals in plants. Toxicology 177, 67–80.

- 36. Sandalio L.M., Dalurzo H.C., Gómez M., Romero-Puertas M.C., Del Río, L.A., 2001. Cadmium-induced changes in the growth and oxidative metabolism of pea plants. Journal of Experimental Botany 52, 2115-2126.
- 37. Sanita di Toppi, L., Gabbrielli, R., 1999. Response to cadmium in higher plants. Environmental and Experimental Botany 41, 105-130.
- 38. Seeram N.P., Lee, R., Sheuller, S., Heber, D., 2005. Identification of phenolic compound in strawberries by liquid chromatography ionization mass spectroscopy. Food Chemistry 97, 1-11.
- 39. Singleton, V. L., Rossi, J. A., 1965. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. American Journal of Enology and Viticulture 16, 144-158.
- 40. Szőllősi, R., Varga, I.Sz., Erdei, L., Mihalik, E., 2009. Cadmiun-induced oxidative stress and antioxidative mechanisms in germinating Indian mustard (*Brassica juncea* L.) seeds. Ecotoxicology and Environmental Safety 72, 1337-1342.
- 41. Tolrá, R., Barceló, J., Poschenrieder, C., 2009. Constitutive and aluminium-induced patterns of phenolic compounds in two maize varieties differing in aluminium tolerante. Journal of Inorganic Biochemistry 103, 1486-1490.
- 42. Vardar, F. and Ünal M., 2007. Aluminum toxicity and resistance in higher plants.

  Advances in Molecular Biology 1, 1-12.
- 43. Wang, C., Chen, C., Wang, S., 2009. Changes of flavonoid content and antioxidant capacity in blueberries after illumination with UV-C. Food Chemistry 117, 426-431.
- 44. Williams, C.H., David, D.J., 1973. The effect of Superphosphate on the cadmium content of soils and plants. Australian Journal of Soil Research 11, 43-56.

- 45. Yamamoto, Y., Kobayashi, Y., Devi, S.R., Rikiishi, S. and Matsumoto, H., 2002. Aluminum toxicity is associated with mitochondrial dysfunction and the production of reactive oxygen species in plant cells. Plant Physiology 128, 63-72.
- 46. Zheljazkov, V. D., Craker, L. E., Baoshan, X., 2005. Effects of Cd, Pd, Cu on growth and essential oil contents in dill, pepermint and basil. Environmental and Experimental Botany 58, 9-16.

## DISCUSIÓN GENERAL

La deficiencia o exceso de nutrientes produce en las plantas diferentes desórdenes metabólicos (Liu et al., 2007; Benavides et al., 2005; Gaballah et al., 2012). En general las plantas son más resistentes al incremento de la concentración de nutrientes, que a la insuficiencia de los elementos esenciales. En el desarrollo de esta tesis se describió (*Capítulo I*), que elementos como Zn, Mn, Fe, Cu, Mg entre otros, son esenciales para el crecimiento y desarrollo normal de la planta, sin embargo elementos como Cd y Al que se encuentran por diferentes fuentes presentes en los suelos, pueden resultar tóxicos para las plantas (Guo et al., 2007; Wulff et al., 2010).

Los sistemas de cultivo alternativos como solución nutritiva, hidropónica o cultivo *in vitro* pueden ser considerados modelos del sistema suelo o más específicamente de la solución suelo. La ventaja principal de los medios de cultivo es que la composición de la solución de cultivo se puede definir, manipular y diseñar con un alto nivel de precisión (Wulff *et al.*, 2010).

Los sistemas de cultivo en solución son una simplificación del sistema suelo, donde las raíces de las plantas obtienen de manera ideal los nutrientes, mientras que en el suelo las raíces deben explorar continuamente para obtener los nutrientes y el agua. Sin embargo, este sistema ideal puede ser alterado cuando se desea investigar los efectos de elementos que no son componentes de estas soluciones. Cada medio de cultivo, debido a su composición, tiene una alta capacidad complejante, y por lo tanto tiende a modificar de manera significativa la distribución de las formas químicas en las que se encuentran los compuestos presentes cuando se adiciona un elemento que pueda interaccionar con los

agentes complejantes. De esta manera la aplicación de metales como Cd y Al al medio de crecimiento altera la composición química de la solución, dejando en algunos casos mayor disponibilidad de elementos esenciales para la planta, la cual puede ver alterado su metabolismo por el aumento de la concentración de nutrientes en el medio.

El arándano, tanto sus frutos como sus hojas poseen una alta concentración de compuestos activos antioxidante de lo hace de este cultivo importante desde el punto de vista nutricional y económico (Castrejón *et al.*, 2008). Esta especie en los últimos años ha aumentado su cultivo en las zonas central y sur de Chile, donde se caracterizan los suelos por tener un pH ácido donde se presenta una alta concentración de Al libre (Mora *et al.*, 2009), además de aumentar la actividad fitotóxica del Cd presente en los suelos por diferentes factores como la fertilización (Molina *et al.*, 2009).

La aplicación de Al y Cd al medio de cultivo (*Capítulo II y III*) produjo efectos similares en las plántulas de arándano, daño oxidativo medido mediante el contenido de MDA. Diversas investigaciones han demostrados que ambos metales producen peroxidación lipídica en diversas especies (Ribeiro *et al.*, 2012; Ge *et al.*, 2012; Cavusoglu *et al.*, 2010). Sin embargo el estrés oxidativo es generados por un aumento de las especies reactivas de oxigeno (ROS) en el sistema, las cuales pueden verse aumentada por diversos factores abióticos, como estrés por metales, cambio en la disponibilidad de nutrientes en el sistema, estrés hídrico y salino (Arora *et al.*, 2002). Esta información nos da cuenta que el daño observado por la aplicación de Al y Cd al medio podría ser una resultante de los estreses producidospor al cambio en disponibilidad de nutrientes debidola adición de metales al medio. Para determinar las especies químicas de los componentes que

constituyen la solución nutritiva, se realizó una especiación química utilizando el programa GEOCHEM (Parker *et al.*, 1995). Se determinó que entre un 40-70 % de los metales aplicados (Al y Cd), formaron complejos con EDTA, modificando la disponibilidad nutrientes como Fe, Zn y Cu disponibles para la planta.

Esta tesis además contempló analizar el efecto combinado de Al+Cd aplicados en medios de cultivos, donde una de las principales diferencias con los estudios individuales fue la mayor respuesta antioxidantes, el cual se vio reflejado en una disminución en el daño por peroxidación lipídica (Capítulo IV). La aplicación de ambos metales al medio de modificó la disponibilidad de nutrientes, observándose una variación de Zn libre, aumentando 7 veces su disponibilidad en el tratamiento binario de Al+Cd con respecto al medio enriquecido con solo un metal. El Zn es un micronutriente esencial, el cual forma parte diferentes enzimas, jugando un rol crítico en los sistemas de defensa en contra las especies reactivas de oxígeno que inducen peroxidación lipidica (Cakmak, 2000; Smeets et al., 2005). Se ha observado que desbalances en el contenido de Al y Cd de manera independiente provocan un aumento en la peroxidación lipídica. En el caso Cd este puede ingresar a la planta, donde genera un incremento en la actividad de lipoxigenasa (LOXs), enzima encargada de oxidación de los ácidos grasos, (Smeets et al., 2008), mientras que el Al puede producir peroxidación lipidica debido a la rigidificación de la membrana al unirse este a los fosfolopídos presentes en ella (Devi et al., 2003).La aplicación binaria de metales aumento la disponibilidad de Zn en el medio de crecimiento, lo cual sugiere que este elemento, ayudaría a disminuir los posibles daños asociados a la presencia de elementos como Cd y Al en el medio de crecimiento, aumentando la respuesta antioxidante del arándano. Resultados similares se han encontrado en plantas tratada con Cd, donde al

aumentar la disponibilidad de Zn en el medio, se observa una reducción del daño por peroxidación lipidica (Cherif *et al.*, 2011).

La acumulación de compuestos fenólicos en plantas puede ser inducida por estreses bióticos y abióticos. Diversos estudios indican que estrés abiótico producido por metales pesados promueven la producción de ROS. La capacidad antioxidante de los compuestos fenólicos se encuentra estrechamente ligada al número de grupos hidróxilos que posea la molécula (donde la actividad antioxidante se incrementa en aquellos compuestos con una alta sustitución de grupos -OH), de su estructura (grado de plimerización) y los sustituyentes (Sgherii et al., 2003; Caillet et al., 2006). Los posibles mecanismos involucrados establecen donación de electrones o átomos de hidrógeno a los radicales libres, generándose un radical fenoxilo que es menos reactivo ya que se estabiliza por resonancia en el anillo aromatico (Procházková et al., 2011). Además de la función antioxidantes que poseen estos compuestos, estos también participan como quelantes de metales (Vasconcelos et al., 1999). Sin embargo existe poca información que dé cuenta de los cambios en el perfil de compuestos fenólicos y su rol en la desintoxicación de ROS y el impacto de los metales pesados. Una investigación realizada por Tolra et al., (2005) detectaron cambios en el contenido de ácido ferúlico y p-cumárico en plantas de Rumex acetosela tratadas con aluminio, sin embargo no fue posible establecer si estos compuestos están implicados en la desintoxicación de aluminio.

En nuestra investigación la presencia de Al y Cd provocó cambios en la biosíntesis de compuestos fenólicos en arándano, principalmente ácido clorogénico y elágico, los cuales son reconocidos por ser metabolitos con propiedades antioxidantes e inhibidores de la

actividad en los tumores (Charurin *et al.*, 2002; Yen *et al.*, 2005; Tavani and Vecchia, 2004, Koshiro *et al.*, 2006). Estos cambios pueden ser consecuencia de una serie de procesos desencadenados por los contenidos de Cd y Al, tales como:

- 1. La enzima Fenilalanina Amonio-Liaza (PAL por sus siglas en inglés), una de las enzimas que forma parte de la vía Fenilpropanoide, principal ruta de biosíntesis de compuesto fenólicos, ve incrementada su actividad como consecuencia del aumento de especies reactivas de oxígeno (ROS), los cuales juegan un rol de señalizadores para esta enzima, frente a condiciones de peroxidación lipídica. Estudios han encontrado una correlación positiva entre la actividad de la PAL y la acumulación de compuestos fenólicos y flavonoides solubles, en hojas tratadas con Cd de *Azolla imbricata* (Dai *et al.*, 2006) y en las raíces de *Panax ginseng* sometidas a la presencia de Cu (Ali *et al.*, 2006).
- 2. La competencia que se genera entre los micronutrientes y elementos trazas por agentes complejantes, los cuales pueden estar presente en los medios de crecimiento (EDTA) o ser exudados radicales, ocasionan cambios en las disponibilidades de micronutrientes. En esta tesis se determinó que la adición de Al y Cd al medio de cultivo modificó la disponibilidad de nutrientes. Estas modificaciones alterarían el funcionamiento de la enzima PAL, debido a que esta enzima responde de manera diferenciada a desequilibrios en el contenido de micronutrientes,incrementando su actividad, lo que explicaría el incremento en los contenidos de los ácidos clorogénico y elágico, en las plántulas tratadas (Luo et al., 2010).

El alto contenido de compuestos fenólicos que contiene el arándano posee una marcada respuesta antioxidante, lo cual se vio reflejado en los valores de DPPH y FRAP medidos a los extractos etanólicos. Nosotros sugerimos que la respuesta de las plántulas de arándano frente a la aplicación de elementos trazas en la actividad antioxidante, puede estar directamente relacionada con el aumento de estrés oxidativo generado por los cambios en el medio de crecimiento.

La adición de Cd y Al en sistemas mono y multicomponentes en el medio de cultivo *in vitro*, han permitido correlacionar positivamente los compuesto fenólicos con la respuesta antioxidante y negativamente con el daño por peroxidación lipídica, siendo estos resultados la base para establecer posibles modulaciones de compuestos fenólicos, principalmente ácido clorogénico y elágico, cuyas importancias nutricionales y médicas son relevantes para las proyecciones del arándano.

## **REFERENCES**

- 1. Ali, M. B., Singh, N., Shohael, A. M., Hahn, E. J., Paek, K. Y., 2006. Phenolics metabolism and lignin synthesis in root suspension cultures of *Panax ginsengin* response to copper stress. Plant Science 171, 147–154.
- 2. Arora, A., Sairam, R.K., Srivastava, G.C., 2002. Oxidative stress and antioxidative system in plants. Current Science 82, 1227-1238.
- 3. Benavides, M.P., Gallego, S.M., Tomaro, M., 2005. Cadmium toxicity in plants. Brazilian Journal of Plant Physiology 17, 21-34.

- 4. Caillet, S., Salmiéri, S., Lacroix, M., 2006. Evaluation of free radical-scavenging properties of commercial grape phenol extracts by a fast colorimetric method. Food Chemistry 95, 1-8.
- 5. Cakmak, I., 2000. Possible roles of zinc in protecting plant cells form damage by reactive oxygen species. New Phytologist 146, 185-205.
- Castrejón, A., Eichholz, I., Rohn, S., Kroh, L., Huyskens-Keil, S., 2008. Phenolic profile and antioxidant activity of highbushblueberry (*Vaccinium corymbosum*L.) during fruitmaturation and ripening. Food Chemistry 109, 564-572.
- 7. Cavusoglu, K., Yalcin, E., 2010. Detection of lipid peroxidation and cytotoxicity induced by aluminium (Al) and cobalt (Co) ions in barbunia root tip cells. Journal of Environmental Biology 31, 661-666.
- 8. Charurin, P., Ames, J. M., Del Castillo M. D., 2002, Antioxidant activity of coffee model systems. Journal of Agriculture and Food Chemistry 50, 3751-3756.
- 9. Cherif, J., Mediouni, Ch., Ammar W. B., Jemal, F., 2011. Interactions of zinc and cadmium toxicity in their eects on growth and inantioxidative systems in tomato plants (*Solanum lycopersicum*). Journal of Environmental Sciences 23, 837-844.
- 10. Dai, L.P., Xiong, Z. T., Huang, Y., Li, M. J., 2006. Cadmium-induced changes in pigments, total phenolics, and phenylalanine ammonia-lyase activity in fronds of *Azolla imbricata*. Environmental Toxicology 21, 505–512.

- 11. Devi, R. S., Yamamoto, Y., Matsumoto, H., 2003. An intracellular mechanism of aluminum tolerance associated with high antioxidant status in cultured tobacco cells. Journal of Inorganic Biochemistry 97, 59-68.
- 12. Gaballah, M., Rady, M.M., 2012. Salicylic Acid Mitigated Cadmium Toxicity by Attenuating the Oxidative Stress in Pea (*Pisum sativum* L.) Plants. International Journal of Biological. Ecological and Environmental Sciences 4, 159-165.
- 13. Ge, W., Jiao, Y.Q., Sun, B.L., Qin, R., Jiang, W.S., Liu, D.H., 2012. Cadmium-mediated oxidative stress and ultrastructural changes in root cells of poplar cultivars. South African Journal of Botany 83, 98-108.
- 14. Guo, T. R., Zhang, G.P.and Zhang, Y.H., 2007. Physiological change in barley plants under combined toxicity of aluminum, copper and cadmium. Colloids and Surface B: Biointerfaces 57, 182-188.
- 15. Koshiro, Y., Zheng, X.Q., Wang, M., Nagai, C., Ashihara, H., 2006. Changes in content and biosynthetic activity of caffeine and trigonelline during growth and ripening of Coffea Arabica and Coffea canephora fruits. Plant Science 171, 242-250.
- 16. Liu, X.L., Zhang, S.Z., Shan, X.Q., Christie, P., 2007. Combined toxicity of cadmium and arsenate to wheat seedlings and plant uptake and antioxidative enzyme responses to cadmium and arsenate co-contamination. Ecotoxicology and Environmental Safety. 68, 305-313.

- 17. Luo, Z.B, He, X.J, Chin, L., Tang, L., Gao, S., Chin, F., 2010. Effects of zinc on growth and antioxidant response in *Jatropha curcas* seedlings. International Journal of Agriculture and Biology 12, 119-124.
- 18. Molina, M., Aburto, F., Calderón, R., Escudey, M., Cazanga, M., 2009. Trace element composition of selected fertilizers used in Chile with special regard to phosphorus fertilizers. Soil and Sediment Contamination 18, 497-511
- 19. Mora, M.L., Rosas, A., Ribera, A., Rengel, Z., 2009. Differential tolerance to Mn toxicity in perennial ryegrass genotypes: involvement of antioxidative enzymes and root exudation of carboxylates. Plant Soil 320, 79-89.
- 20. Parker, D.R., Norvell, W.A., Chaney R.L., 1995. GEOCHEM-PC: A chemical speciation program for IBM and compatible personal computers. p. 253-269. In R. H. Loeppert et al. (ed.) Chemical equilibrium and reaction models. SSSA Spec. Publ. 42, ASA and SSSA, Madison, WI.
- 21. Procházková, D., Boušová, I., Wilhelmová, N., 2012. Antioxidant and prooxidant properties of flavonoids. Fitoterapia 82, 513-523.
- 22. Ribeiro, C., Cambraia, J., Henrique, P., Peixoto, P., Meira, E., 2012. Antioxidant system response induced by aluminum in two rice cultivars. Brazilian Society of Plant Physiology 24, 107-116.
- 23. Sgherri, C., Cosi, E., Navari-Izzo, F., 2003. Phenols and antioxidative status of Raphanus sativus grown in copper excess. Physiologia Plantarum 118, 21–28.

- 24. Smeets, K., Cuypers, A., Lambrechts, A., Semane, B., Hoet, P., Laerve, A.V., Vangronsveld, J., 2005. Induction of oxidative stress and antioxidative mechanisms in Phaseolus Vulgaris after Cd application. Journal of Plant Physiology and Biochemistry 43, 437-444.
- 25. Smeets, K., Ruytinx, J., Semane, B., Van Belleghem, F., Remans, T., Van Sanden, S., 2008. Cadmiun-induced transcriptional and enzymatic alterations related to oxidative stress. Environmental and Experimental Botany 63,1-8.
- 26. Tavani A., Vecchia C. L., 2004. Coffee, decaffeinated coffee, tea and cancer of the colon and rectum: A review of epidemiological studies, 1990-2003. Cancer Causes and Control 15, 743-757.
- 27. Tolrà, R.P., Poschenrieder, Ch., Luppi, B., Barceló, J., 2005. Aluminium induced changes in the profiles of both organic acids and phenolic substances underlie Al tolerance in *Rumex acetosela* L. Environmental and Experimental Botany 54, 231–238.
- 28. Vasconcelos, M.T., Azenha, M., de Freitas, V., 1999. Role of polyphenols in copper complexation in red wines. Journal of Agriculture and Food Chemistry 47, 2791–2796.
- 29. Wulff, M., Cellini, A., Masia, A., Marino, G., 2010. Aluminium induced effects on growth, morphogenesis and oxidative stress reaction in *in Vitro* cultures of quince. Scientia Horticulturae 125, 151-158.

30. Yen, W. J., Wang, B.S., Chang, L.W., Duh, P.D., 2005. Antioxidant properties of roasted coffee residues. Journal of Agriculture and Food Chemistry 53, 2658-2663.

#### **CONCLUSIONES GENERALES**

En este estudio se logró desarrollar e implementar un sistema de propagación *in vitro* de arándano y aplicar diferentes tratamientos destinados a generar estrés por metales (Al<sup>3+</sup>y Cd<sup>2+</sup>), modificando la composición de las soluciones de crecimiento y evaluando los cambios en diferentes parámetros fisiológicos, ocasionados por la presencia de agentes inductores de estrés, con el fin de estudiar la producción de compuestos fenolicos producidos bajo condiciones de estrés.

Mediante un estudio de especiación química realizada a las soluciones del medio de cultivo, fue posible establecer que especie(s) de los diferentes metales aplicados como tratamientos (Al<sup>3+</sup> y Cd<sup>2+</sup>), tienen un rol importante en los efectos observados en arándano. Adicionalmente fue posible determinar que la concentración libre del metal es más importante que su actividad, en relación con el comportamiento a nivel de daño oxidativo yrespuesta antioxidante en arándano. Este procedimiento demostró la importancia de los estudios de especiación química en matrices complejas, ya sea en solución nutritiva y/o cultivos *in vitro*, en donde existen componentes acomplejantes, que modifican la distribución de las especies presentes de cada elemento en el sistema.

Se investigó la aplicación de Al en plántulas de arándanos cultivados *in vitro (capitulo II)*. El estudio mostró que los extractos de arándano poseen un alto contenido de compuestos fenólicos, además de una alta actividad antioxidante. La aplicación de Al<sup>3+</sup> al medio de cultivo produjo efectos en la capacidad antioxidante y un aumento en el daño por peroxidación lipídica donde los mayores efectos se observaron a los 14 días de tratamiento. El contenido de compuestos fenólicos aumento, principalmente ácido clorogénico y ácido

elagico, este comportamiento se observó independiente de la concentración de aluminio aplicado, sugiriendo que estos compuestos fenólicos juegan un rol importante en la respuesta ante el estrés producido por este metal.

Las plántulas de arándano que crecieron en presencia de Cd mostraron un daño significativo por peroxidación lipídica en todas las concentraciones aplicadas, siendo el mayor daño a los 14 días de exposición. En términos de la respuesta antioxidante, si bien se observó un aumento respecto del tratamiento, no fue suficiente para controlar el daño oxidativio generado, pues para ambas concentraciones de Cd, los niveles de MDA fueron mayores que en el control respectivo. Esto sugiere que las plantas de arandano cultivadas in vitro, presentan una alta sensibildiad al Cd.

La aplicación de diferentes concentraciones de Cd<sup>2+</sup>, al medio de cultivo de plantulas de arándano produjo efectos notorios en los contenidos de compuestos fenólicos. La respuesta de la planta frente a la presencia del metal fue diferenciada con respecto a la concentración de Cd, observándose una mayor respuesta a una mayor concentración presente en el medio.

Los efectos de la presencia de Cd + Al en el medio de cultivo de plantulas de arandano se estudiaron en el capitulo IV. Los resultados obtenidos mostraron que se induce daño oxidativo en las etapas iniciales del tratamiento, sin embargo, luego a partir de los 14 días, se produce un aumento en la respuesta antioxidante, lo que estaría relacionado con un aumento en el contenido de compuestos fenólicos cuantificados, donde el componente principal es el ácido clorogénico, lo que permite concluir que la actividad antioxidante juega un rol importante como mecanismo de defensa frente al daño por elementos trazas.

Por lo tanto nosotros podemos concluir luego del desarrollo de esta tesis, que la disponibilidad de nutrientes y elementos trazas como Cd y Al, en arándano se encuentra estrechamente relacionada con los componentes totales de la solución de crecimiento, los cuales pueden aumentar su toxicidad o disminuirla, dependiendo de los agentes complejantes y el pH en el que se encuentra el medio de crecimiento (solución nutritiva, cultivos *in vitro* o suelos). De esta manera la aplicación de elementos trazas como Cd y Al producen efectos importantes en elperfil de compuestos fenolicos, aumentando en algunos casos su concentración, como por ejemplo el ácido clorogénico y ácido elégico, concluyendo que si es posible manipular el medio de crecimiento de manera de modular a la planta para la generación de compuestos bioactivos de interés.

### **Futuras Proyecciones**

Las proyecciones que surgen de este trabajo son:

- El desarrollo de una metodología de estudios basados en la producción *in vitro* de arándano u otras especies vegetales, donde es posible modificar y simular factores medioambientales, permitiendo investigar la respuesta de las plantas desde un punto de vista metabolómico, conociendo como posibles alteraciones en los factores ambientales modifican la regulación de la biosíntesis de compuestos bioactivos antioxidantes.
- El sistema de cultivo *in vitro* de arándano o cualquier especie a la cual se propague y desarrolle en óptimas condiciones, permitirá desarrollar estudios de parámetros fisiológicos, bioquímicos y moleculares, orientados a identificar mecanismos de tolerancia o resistencia a condiciones externas como la aplicación de diferentes estreses medioambientales.
- Considerando que las plantulas de arándano cultivados *in vitro* poseen un alto contenido de compuestos fenólicos, los cuales pueden ser en algún grado modificados por la presencia de metales pesados. Entre los principales compuestos fenólicos se destaca el ácido clorogénico, el cual es un compuesto de gran interés comercial debido a su alto poder antioxidante. De esta manera el uso de los extractos de arándano a nivel comercial podría significar un aporte para la generación individuos *in vitro* con un alto poder antioxidante.
- El poder reductor de los extractos de arándano obtenidos en cultivos *in vitro*, sometidos a diferentes tratamientos con metales pesados, están siendo evaluados como agentes reductores para la síntesis verde de diferentes nanopartículas

metálicas de valencia cero (nMVC), lo que abre perspectivas en la revalorzación de los residuos generados en el proceso de cosecha y poda de arándano, permitiendo el cruce entre diferentes áreas de la ciencia. Este trabajo está siendo desarrollado bajo el marco del proyecto CORFO 12IDL2-16251 y el Centro de Nanociencia y Nanotecnología (CEDENNA)



Artículo Original | Original Artícle

# Effect of aluminum on antioxidant activity and phenolic compounds content in in vitro cultured blueberries

[Efectos de aluminio en la capacidad antioxidante y en el contenido de compuestos fenólicos en plántulas de arándano cultivadas in vitro]

Karen MANQUIÁN\*, Gustavo E. ZÚÑIGA1, Herna BARRIENTOS1, Mauricio ESCUDEY1,2 & Mauricio MOLINA3

Facultad de Química y Biología, Universidad de Santiago de Chile, Av. B. O'Higgins, 3363, Santiago, Chile. <sup>2</sup>Center for the Development of Nanoscience and Nanotechnology, CEDENNA, Santiago, Chile. Departamento de Industrias, Universidad Técnica Federico Santa María, Av. Santa María 6400, Santiago, Chile. \*Ph.D programm in Ciencia de Recursos Naturales. Universidad de la Frontera, Temuco, Chile. Contactos | Contacts: Gustavo E. ZÚÑIGA - E-mail address: gustavo.zuniga@usach.cl Contactos | Contacts: Mauricio ESCUDEY - E-mail address: mauricio.escudey@usach.cl

#### Abstract

Blueberry is a popular natural food product consumed worldwide. Acid soils are found throughout the world. A significant problem of acid soils is the active aluminum content, which may result toxic to plant. The present study was undertaken to assess the toxicities of Al for Blueberry (Vacclutum corymbosum L.) cultivated in vitro and treated with 100 and 200 µM Al. The effects of Al concentration on malondialdehyde (MDA) content, antioxidant activity and phenolic compounds of blueberry after 7, 14 and 21 days of treatment were established. The analysis of the MDA accumulated in the tissues of the blueberry seedlings indicates that AI concentration increases the damage caused by lipid peroxidation, for both treatments, after 14 days. The highest antioxidant activity in the extracts was observed at 200 µM Al after 14 days of treatment, being chlorogenic and ellagic acids the most significant metabolites involved in the antioxidant properties. Then, the content of Al in soil could be modulate the content of bioactive compounds in blueberry plants.

Keywords: Blueberry, aluminum, antioxidant capacity, phenolic compounds.

El Arándano es un popular alimento natural consumido en todo el mundo. Los suelos ácidos se encuentran en todo el mundo. Un problema significativo de suelos ácidos es el contenido de aluminio activo, que puede resultar tóxico para la planta. Este estudio se realizó para evaluar la toxicidad del aluminio en plantas de arandano, cultivadas in vitro y tratadas con 100 y 200 mM de Al. Se establecieron los del aluminio en el contenido de malodialdehido (MDA), capacidad antioxidante y contenido de compuestos fenolicos en plantulas de arandano luego de 7, 14 y 21 dias de tratamiento. El análisis del MDA acumulado en los tejidos de las plantulas de arándanos indica que la concentración de Al aumenta el daño causado medido como peroxidación de lípidos, para ambos tratamientos, después de 14 días. La actividad antioxidante más alta de los extractos se obseva a 200 mM de Al después de 14 días de tratamiento, siendo los ácidos clorogênico y elágico los metabolitos más importantes que participan en las propiedades antioxidantes. Entonces, el contenido de Al en el suelo podría modular el contenido de compuestos bioactivos en plantas de arindanos, alterando sus propiedades medicinales.

Palabras Clave: Arándano, aluminio, capacidad antioxidante, compuestos fenólicos.

Recibido | Received: April 7, 2013

Aceptade en versión corregida | Acceptad in revised form: April 23, 2013 Publicado en línea | Published online: November 30, 2013

Declaración de intereses | Declaración of interests: Financial support of this work by the Center for the Development of Nanoscience and Nanotechnology (CEDENNA), and DICYT-DGT (Universidad de Santiago de Chile, GE. Zúñiga) is gratefully acknowledged

Este artículo puede ser citado como / This artícle must be cited as: K Manquian, G Zúñiga, H Barrientos, M Escudey, M Molina. 2013. Effect of aluminum on antioxidant activity and phenolic compounds content in in vivo cultured blueberries. Bol Latinoam Caribo Plant Med Aromat 12(6): 603 – 611.